# Survey on relationship between acute phase proteins (serum amyloid A, milk amyloid A and serum haptoglobin) in inflammatory diseases of dairy cattle

Ezzatollah Fathi<sup>1</sup>\*, Raheleh Farahzadi<sup>2</sup>

<sup>1</sup>Department of Clinical Sciences, Faculty of Veterinary Medicine, University of Tabriz, Tabriz, Iran <sup>2</sup>Department of Clinical Biochemistry, Faculty of Medical Sciences, Tarbiat Modarres University, Tehran, Iran

Received: November 21, 2012

Accepted: February 12, 2013

#### Abstract

The objective of this study was to evaluate the concentration of some acute phase proteins (serum amyloid A, milk amyloid A and serum haptoglobin) and alteration in hematological values in some inflammatory diseases in dairy cattle. Fifty-six Holstein dairy cows with inflammatory diseases were used in this study including: subclinical mastitis, hematological infections (theileriosis and anaplasmosis), metritis, pneumonia and displacement of abomasum (n= 8 each). Clinically healthy cows were selected as control group (n= 8). Blood samples were collected from jugular vein of both groups. Milk samples were taken from four quarters of each cow and mixed together and then one sample of pooled milk from both groups was taken. There were significant differences in mean concentrations of serum and milk amyloid A between clinically healthy cows and diseased cows and also, between abomasal displacement and other infectious diseases (p < 0.05). There were also significant differences in haptoglobin concentrations between cattle suffering from subclinical mastitis, theileriosis and pneumonia compared to those with anaplasmosis, abomasal displacement and metritis (p < 0.05). Our results indicated the application of serum haptoglobin, serum amyloid A and milk amyloid A measurements as indicators of inflammatory diseases in dairy cattle. In conclusion, the current study shows that haptoglobin, serum amyloid A, and milk amyloid A measurements, could be successfully applied in cattle as valuable indicators of inflammatory diseases.

Keywords: Acute phase protein, serum amyloid A, haptoglubin, inflammatory diseases

<sup>\*</sup>Corresponding author: Ezzatollah Fathi

Email: ez.fathi@tabrizu.ac.ir, fathi\_vet79@yahoo.com

Tel: +98 411 3392351

Fax: +98 411 3357834

P.O.: 51666-16471

## Introduction

Detailed functions of the acute phase proteins (APPs) are not fully understood, however, it is suggested that they are mainly engaged in processes like opsonization and trapping of microorganisms and their products, binding of cellular remnants (Whicher and Westacott, 1992), complement activation, neutralization of enzymes, and scavenging of free radicals and hemoglobin (Niewold et al., 2003). In the recent studies concerning new indicators of mastitis, acute phase proteins are strongly implicated (Eckrsall et al., 2001 Grönlund et al., 2003, Nielsen et al., 2004, Grönlund et al., 2005, , O'Mahony et al., 2006, Ihnat et al., 2007, Akerstedt et al., 2007, Hiss et al., 2007). APPs are often determined in serum of animals to monitor their health (Kostro et al., 2001; Murata et al., 2004, Petersen et al., 2004). APPs can be valuable to the veterinarian in identification of cows with inflammatory diseases and the stages of disease can be better evaluated by monitoring more than one APP like serum amyloid A (SAA) and serum haptoglobin (Hp), so chronic conditions as well as acute ones should be evaluated and characterized by APP profiling (Karreman et al., 2000, Eckersall, 2004). Hp is an  $\alpha_2$ -globulin and it is one of the APPs, which increases in serum in acute inflammatory diseases. Measurement of APPs could be a useful tool for evaluation of health in calf herds (Ganheim et al., 2007). SAA is an apolipoprotein of high-density lipoprotein (Nakayama et al., 1993, Gruys et al., 1994; Husby et al., 1994). It is described to bind lipopolysaccharide, comparable to lipopolysaccharide binding protein (Schroedl et al., 2001). There are no published reports about the comparison of serum Hp, SAA and MAA concentrations in subclinical mastitis, theileriosis. anaplasmosis. pneumonia. metritis. and abomasal displacement. Therefore, the aim of this study was to evaluate these **APPs** as inflammatory indicators in various inflammatory diseases of cows.

# Materials and methods

Fifty-six Holstein cows with various inflammatory diseases from three dairy farms located in Tabriz suburb were included in this research. Eight clinically healthy adult cattle were also selected randomly as control. The diseased cows used in this study included subclinical mastitis. parasitic infections (theileriosis and anaplasmosis), pneumonia, metritis and abomasal displacement (n=8 each). The presence of disease was assessed on the basis of clinical examination and laboratory findings. Diseased cows were thoroughly examined and blood samples were collected from jugular vein of both groups for hematology, clinical biochemistry and other relevant analysis. To determine serum Hp, SAA was MAA serum separated and bv centrifugation at 750 g for 15min. Serum samples were stored at -20 °C until analysis. SAA and MAA concentrations were measured by using a solid phase sandwich enzyme-linked immunosorbent (ELISA) (Tridelta assay Development Pic, Co.Wicklow, Ireland) and Hp was measured on the basis of the preservation of the peroxidase activity of hemoglobin which is directly proportional to the amount of haptoglobin using a colorimetric commercial kit (Tridelta Development Plc, Co., Wicklow, Ireland). Statistical analysis was performed using SPSS software (Ver. 16, IBM Corporation, USA).

The data were checked for errors and compared with written reports; Outliers were rechecked during data processing to ensure that values were accurate. All variables were screened for normality by visual assessment of the distributions. All values were expressed as mean and standard error (mean $\pm$ SE). Differences between groups were analyzed using one way ANOVA and Duncan's multiple comparison tests and considered as statistically significant when the *p* value was less than 0.05.

# Results

The mean concentrations of SAA and MAA and their correlation in cows with inflammatory diseases and clinically healthy cows are presented in Table 1. Statistical evaluations showed that there was no significant correlation between SAA and MAA in each of inflammatory diseases (p<0.05). There were significant differences in mean concentrations of SAA and MAA between clinically healthy cows and diseased cows and also, between abomasal displacement and other infectious diseases (p < 0.05). Significant difference in Serum Hp concentrations was observed between clinically healthy cows and those with inflammatory diseases (p < 0.05). There were also significant differences in Hp concentrations between cattle suffering from subclinical mastitis. theileriosis and compared those with pneumonia to anaplasmosis, abomasal displacement and metritis (p < 0.05).

Groups				
Parameters	Number	SAA (µg/ml)	MAA (µg/ml)	<b>Correlation coefficient</b>
Subclinical mastitis	8	29± 2.1 <sup>a</sup>	$15 \pm 0.9^{a}$	-0.03
Theileriosis	8	$33 \pm 3.9^{a}$	$29 \pm 1.2^{b}$	-0.0033
Anaplasmosis	8	$30 \pm 4.3^{a}$	$17 \pm 2.6^{b}$	-0.003
Pneumonia	8	$36 \pm 2.6^{a}$	$28 \pm 3.8^{b}$	-0.0039
Metritis	8	$41 \pm 0.09^{a}$	$61 \pm 1.1^{c}$	0.008
Abomasal displacement	8	19± 4.9 <sup>b</sup>	$8\pm0.8^{ m d}$	-0.038
Clinically healthy cows	8	$6 \pm 0.5^{c}$	$0.5 \pm 0.08^{e}$	0.04*

 Table 1. SAA and MAA concentration (mean ± SE) in cows with inflammatory diseases and clinically healthy cows.

Means within a column with different superscript letters (a, b, c, d, e) significantly differ (p<0.05). \*Indicates p<0.05

#### Discussion

Numerous studies have described the serum concentration of APPs in animals with various diseases, but only a few are devoted to their levels in milk from cows with inflammatory disease. Therefore, the present study was undertaken to determine the concentration of SAA, MAA and serum hptoglobin and assess their usefulness for the detection of common inflammatory diseases in cows. Quantification of APP concentration in serum can provide valuable diagnostic information in the prognosis, detection, and monitoring of diseases in several animal species. The inclusion of APPs measurements in health monitoring programs on a herd basis in livestock has been suggested, not only for the identification of individual animals with disease, but also as a mean of identifying animals with subclinical disease. Measurement of APPs, especially Hp and SAA, as indicators of health in herds has gained an increasing interest among researchers (Murata et al., 2004, Ganheim et al., 2007). Hp and SAA have been found to increase in the serum of cattle affected by many different diseases al.,1994, (Alsemgeest et Horadagoda et al.,1994, Godson et al.,1996, Hirvonen et al.,1996, Smith et al.,1998, Heegard et al., 2000, Sheldon et al., 2001, Nielson et al., 2004, Murata et al., 2004, Grounland et al., 2005, Fe'lix et al., 2008). SAA could be a useful diagnostic marker of early postcomplications operative and also for evaluation of health in calves (Dabrowski et al.,2007, Ganheim et al., 2007). There is only one published report about the comparison of SAA and MAA concentrations in some infectious diseases (Nazifi et al., 2008a). In the present study the comparative measurement of SAA and MAA as well as serum Hp was conducted as inflammatory indicators in various inflammatory diseases of cattle in East Azarbaijan province located in northwest of Iran. The previous study has revealed that the concentration of SAA was higher in serum and milk of the cows with mastitis than in the cows with extramammary inflammatory conditions (Nielsen et al., 2004). However, in the present study the mean concentrations of SAA and

MAA were lower in serum and milk of those subclinical mastitis and abomasal with displacement than in the cows with other inflammatory diseases. The reason may be that the severity of these two diseases is lower than other inflammatory diseases. The mean concentration of serum Hp in our study was also at the minimum value in cows with abomasal displacement than the others. SAA concentrations below the detection limit were considered as a good indicator of healthy udder quarters (Gronlund et al., 2005). The results of the present investigation showed that there is no correlation between concentration of SAA and MAA in each of the inflammatory diseases under study. These results can be due to intramammary synthesis and secretion of SAA3 to colostrum and mastitis milk and has beneficial functions for the gut mucosa of the offspring (Eckersall et al., 2001, McDonald et al., 2001, Mack et al., 2003, Larson et al., 2003a,b, Larson et al., 2005).In diseased cattle, the lowest conce ntrations of Hp (0.075±0.018 mg/mL) were observed in cows with abomasal displacement (Table 2). In the present study, the concentration of SAA and Hp in cows infected with Theileria annulata was significantly higher than the clinically healthy cows (p < 0.05). An effect of free hemoglobin in serum samples towards reduction of measured concentration of Hp has been found (Eckersall et al., 1999, Petersen et al.,2001). These data are different from our results in cattle with anaplasmosis and theileriosis, in which we observed increased serum Hp concentrations. In one study reported by Glass et al. (2003), following experimental infection with Theileria annulatain in cattle, Hp appeared only in some of the animals, and generally at a low level. This shows that Hp should be used with caution as a marker of inflammation in Theileriosis. In the present study, the concentration of SAA and serum Hp in cows with pneumonia and metritis was significantly higher than clinically healthy cows (p < 0.05). Alsemgeest et al. (1994) introduced Hp as a valuable marker for differentiating healthy cows from pneumonic ones. Godson et al. (1996) evaluated APPs in cows with respiratory infections and introduced Hp as a diagnostic factor. Berry et al.(2004) found that Hp concentrations were higher in calves treated by respiratory disease for multiple times compared with those never treated, or treated on a single occasion. Increasing in Hp and SAA amounts has been reported after intratracheal inoculation with Manheimia hemolytica (Horadagoda et al., 1994, Cheryk et al., 1998). In diseased cows subjected to our study (n=48), Hp levels were significantly higher (0.132±0.069 mg/mL) than those in healthy ones  $(0.02\pm0.14 \text{ mg/mL}; p < 0.05)$ . Increased serum Hp concentration in cattle was found in various acute infections under field conditions (Skinner et al., 1991, Alsemgeest et al., 1994), mastitis (Hirvonen et al., 1999, Ohtsuka et al., 2001, Gronlund et al., 2003, Nielsen et al., 2004, Gronlund et al., 2005), and metritis (Smith et al., 1998). Our study showed that acute respiratory infections resulted in a significant increase of serum Hp levels. These results are in agreement with one previous study carried out by Nazifi et al. (2008b). Similar findings were demonstrated in cattle with experimental infection with Pasteurella haemolytica (Katoh & Nakagawa, 1999, Ganheim et al., 2003). In our study, subclinical mastitis, metritis, and abomasal displacement were also found to provoke significantly elevated Hp values.

In conclusion, the current study shows that Hp, SAA, and MAA measurements could be successfully applied in cattle as valuable indicators of inflammatory diseases. The studied parameter could be influenced by other disorders and significant changes in hemoglobin levels; so it should not be used independently but together with other blood laboratory indices with regard to better elucidation of the systemic status. However, APPs increase during the development of the disease and decrease in the recovery stage could be used for diagnosis of the disease in early stages.

Groups		
Parameters	Number	Serum Hp (mg/ml)
Subclinical mastitis	8	0.15± 0.09a
Theileriosis	8	$0.18 \pm 0.08a$
Anaplasmosis	8	$0.08 \pm 0.12b$
Pneumonia	8	$0.22 \pm 0.03a$
Metritis	8	$0.09 \pm 0.08 b$
Abomasal displacement	8	$0.075 \pm 0.018b$
Clinically healthy cows	8	$0.02 \pm 0.14c$

Table 2. Serum Hp concentrations (mean ± SE) in cows with inflammatory diseases and clinically healthy cows.

Means within a column with different superscript letters (a, b, c, d) significantly different(p<0.05).

#### Acknowledgements

This research was supported by a grant from University of Tabriz.

### References

- Akerstedt, M., Persson, W.K. andSternesjö, A. (2007) Haptoglobin and serum amyloid A in relation to the somatic cell count in quarter, cow composite and bulk tank milk samples. *Journal of Dairy Research* 74, 198-203.
- Alsemgeest, S.P., Kalsbeek, H.C., Wensing, T., Koeman, T., Van Ederen, A.M. and Gruys, E. (1994) Concentrations of serum amyloid-A (SAA) and haptoglobin (HP) as parameters of inflammatory disease in cattle. *Veterinary Questions* 16, 21-23.
- Berry, B.A., Confer, A.W., Krehbiel, C.R., Gill, D.R., Smith, R.A. and Montelongo, M. (2004) Effects of dietary energy and starch concentrations for newly received feedlot calves: II. Acute-phase protein response.

Journal of Animal Science 82, 845–850.

- Cheryk, L.A., McGrevy, K.E. and Gentry, P.A. (1998) Alterations in platelet function and acute phase proteins induced by Pasteurella heamolytica. *Canadian Journal* of *Veterinary Research* **62**, 1-8.
- Dabrowski, R.W. and Wand, K.K. (2007) Changes in CRP, SAA and haptoglobin produced in response to

ovariohysterectomy in healthy bitches and those with pyometra. *Theriogenology* **67**, 321-327.

- Eckrsall, P.D., Young, F.J., McComb, C., Hogarth, C.J., Safi, S., Weber, A., McDonald, T., Nolan, A.M. and Fitzpatrick J.L.(2001) Acute phase proteins in serum and milk from dairy cows with clinical mastitis. *Veterinary Record* 13, 35-41.
- Eckersall, P.D. (2004) The time is right for acute phase protein assays. *Veterinary Journal* 168, 3–5.
- Eckersall, P.D., Duthie, S., Safi, S., Moffat,
  D., Horagoda, N.U., Doyle, S., Parton,
  R., Bennett, D. and Fitzpatrick, J.L. (1999) An automated biochemical assay for haptoglobin: Prevention of interference from albumin. *Comparative Haematology International* 9, 117-124.
- Fe'lix, H.D., Gonza'lez, F.T., Silvia, M.S., Asta, T., Laura, S.V., Jose'J. and Ceron J.J.(2008) Acute phase protein responses in goats. *The Journal of Veterinary Diagnostic Investigation* 20, 45-48.
- Ganheim, C., Hulten, C., Carlsson, U., Kindahl, H., Niskanen, R. and Waller, K.P. (2003) The acute phase response in calves experimentally infected with bovine viral diarrhea virus and/or Mannheimiahaemolytica. *Journal of Veterinary Medicine* Series B, **50**, 183-190.
- Ganheim, C., Alenius, S. and Waller, K.P. (2007) Acute phase proteins as

Iranian Journal of Veterinary Science and Technology, Vol. 3, No. 2

indicators of calf herd health. *Veterinary Journal* **173**, 645-651.

- Glass, E.J., Craigmile, S.C., Springbett, A., Preston, P.M., Kirvar, E., Wilkie, G.M., Eckersall, P.D., Hall, F.R. and Brown, C.G. (2003) The protozoan parasite, Theileriaannulata, induces a distinct acute phase protein response in cattle that is associated with pathology. *International Journal for Parasitology* 33, 1409-1418.
- Godson, D.L., Campos, M., Attah-Poku, S.K., Redmond, M.J., Cordeiro, D.M., Sethi, M.S., Harland, R.J. and Babiuk, L.A. (1996) Serum haptoglobin as an indicator of the acute phase response in bovine respiratory disease. *Veterinary Immunology and Immunopathology* 51, 277-292.
- Grönlund, U., Hulten, C., Eckersall, P.D., Hogarth, C.J. and Persson, W.K. (2003) Haptoglobin and serum amyloid A in milk and serum during acute and chronic experimentally induced Staphylococcus aureus mastitis. *Journal of Dairy Research* **70**, 379-386.
- Grönlund, U., Hallen, S.C. and Persson, W.K.
  (2005) Haptoglobin and serum amyloid
  A in milk from cows with chronic subclinical mastitis. *Veterinary Research* 36, 191-198.
- Gruys, E., Obwolo, M.J. and Toussaint, M.J.M. (1994) Diagnostic significance of the major acute phase proteins in veterinary clinical chemistry: a review. *Veterinary Bulletin* **64**, 1009-1018.
- Hiss, S., Mueller, U., Neu-Zhren, A. andSauerwein, H. (2007) Haptoglobin and lactate dehydrogenase measurements in milk for the identification of subclinically diseased udder quarters. *Veterinary Medicine* **52**, 245-252.
- Husby, G., Marhaug, G., Dowton, B., Sletten, K. and Sipe, J.D. (1994) Serum amyloid A (SAA): biochemistry,

genetics and the pathogenesis of AA amyloidosis. *Amyloid* **1**, 119-137.

- Heegaard, P.M.H., Godson, D.L., Toussaint, M.J.M., Tjørnehøj, K., Larsen, L.E., Viuff, B. and Rønsholt, L. (2000) The acute phase reponse of haptoglobin and serum amyloid A (SAA) in cattle undergoing experimental infection with bovine respiratory syncytial virus. *Veterinary Immunology and Immunopathology* 77, 151-159.
- Hirvonen, J., Pyärölä, S. and Jousimies-Somer,
  H. (1996) Acute phase response in heifers with experimentally induced mastitis. *Journal of Dairy Research* 63, 351-360.
- Hirvonen, J., Eklund, K., Teppo, A.M., Huszenica, G., Kulcsar, M., Saloniemi, H. and Pyorala, S. (1999) Acute phase response in dairy cows with experimentally induced E. coli mastitis. *Acta Veterinaria Scandinavica* 40, 35-46.
- Horadagoda, A., Eckersall, P.D., Hodgson, J.C., Gibbs, H.A. and Moon, G.M. (1994) Immediate responses in serum TNF- $\alpha$  and acute phase protein concentrations to infection with Pasteurellahemolytica A1 in calves. *Research in Veterinary Medicine* xx, 129-132.
- Ihnát, O. (2007) Interrelationship between somatic cell count and acute phase proteins in serum and milk of dairy cows. *Journal Acta Veterinaria Brno* **76**, 51-57.
- Karreman, H.J., Wentink, G.H. and Wensing, T. (2000) Using serum amyloid A to screen dairy cows for subclinical inflammation. *Veterinary Questions* 22, 175-178.
- Katoh, N. and Nakagawa, H. (1999) Detection of haptoglobin in the high-density lipoprotein fractions from sera of calves with experimental pneumonia and cows with naturally occurring fatty liver. *Journal of Veterinary Medical*

Iranian Journal of Veterinary Science and Technology, Vol. 3, No. 2

Science 61, 119-124.

- Kostro, K., Gliński, Z., Wójcicka, L.K. andKrakowski, K. (2001) Acute-phase proteins as indicators of diseases in animals. *Medycyna Weterinaryna* 57, 539-542.
- Larson, M.A., Weber, A., Weber, A.T. and McDonald, T.L. (2005) Differential expression and secretion of bovine serum amyloid A3 (SAA3) by mammary epithelial cells stimulated with prolactin or lipopolysaccharide. *Veterinary Immunology and Immunopathology* **107**, 255-264.
- Larson, M.A., Wei, S.H., Weber, A., Mack, D.R. and McDonald, T.L. (2003a) Human serum amyloid A3 peptide enhances intestinal MUC3 expression and inhibits EPEC adherence. *Biochemical and Biophysical Research Communications* **300**, 531-540.
- Larson, M.A., Wei, S.H., Weber, A. and McDonald, T.L. (2003b) Induction of human mammaryassociated serum amyloid A3 expression by prolactin or lipopolysaccharide. *Biochemical and Biophysical Research Communications* 301, 1030-1037.
- Mack, D.R., McDonald, T.L., Larson, M.A., Wei, S.H. and Weber, A. (2003) The conserved TFLK motif of mammaryassociated serum amyloid A3 is responsible for up- regulation of intestinal MUC3mucin expression in vitro. *Pediatric Research* **53**, 137-142.
- McDonald, T.L., Larson, M.A., Mack, D.R. and Weber, A. (2001) Elevated extrahepatic expression and secretion of mammaryassociated serum amyloid A3 (M-SAA3) into colostrum. *Veterinary Immunology and Immunopathology* **83**, 203-211.
- Murata, H., Shimada, N. and Yoshioka, M. (2004) Current research on acute phase proteins in veterinary diagnosis: an overview. *Veterinary Journal* **168**, 28-40.

- Nakayama, T., Sonoda, S., Urano, T., Yamada, T. and Okada, M. (1993) Monitoring both serum protein A and C-reactive protein as inflammatory markers in infectious diseases. *Clinical Chemistry* 39, 293-297.
- Nazifi, S., Khoshvaghti, A. and Gheisari, H.R. (2008a) Evaluation of serum and milk amyloid A in some inflammatory diseases of cattle. *Iranian Journal* of *Veterinary Research* **9**, 222-226.
- Nazifi, S., Rezakhani, A., Koohimoghada, M., Ansari, L.M. and Esmailnezhad, Z. (2008b) Evaluation of serum haptoglobin in clinically healthy cattle and cattle with inflammatory diseases in shiraz, a tropical area in southern Iran. *Bulgarian Journal of Veterinary Medicine* **11**, 95-101.
- Nielsen, B.H., Jacobsen, S., Andersen, P.H., Niewold, T.A. and Heegaard, P.M. (2004) Acute phase protein concentrations in serum and milk from healthy cows, cows with clinical mastitis and cows with extramammary inflammatory conditions. *Veterinary Record* 154, 361-365.
- Niewold, T.A., Toussaint, M.J.M. and Gruys, E. (2003) Monitoring health by acute phase proteins. Proceedings, 4th European Colloquium on Acute Phase Proteins. Segovia, Spain.
- Ohtsuka, H., Kudo, K., Mori, K., Nagai, F., Hatsugaya, A., Tajima, M., Tamura, K., Hoshi, F., Koiwa, M. and Kawmura, S. (2001) Acute phase naturally response in occurring coliform mastitis. Journal of Veterinary Medical Science 63, 675-678.
- O'Mahony, M.C., Healy, A.M., Harte, D., Walshe, K.G., Torgerson, P.R. and Doherty M.L. (2006) Milk amyloid A: Correlation with cellular indices of mammary inflammation in cows with normal and raised serum amyloid A. *American Research in Veterinary*

Iranian Journal of Veterinary Science and Technology, Vol. 3, No. 2

Science 80, 155-161.

- Petersen, H.H., Nielsen, J.P., Jensen, A.L. and Heegaard, P.M.H. (2001) Evaluation of an enzyme linked immunosorbent assay (ELISA) for determination of porcine haptoglobin. *Journal of Veterinary Medicine Series A*, **48**, 513-523.
- Petersen, H.H., Nielsen, J.P. and Heegaard, P.M. (2004) Application of acute phase protein measurements in veterinary clinical chemistry. *Veterinary Research* 35, 163-187.
- Schroedl, W., Fuerll, B., Reinhold, P., Krueger, M. and Schuett, C. (2001) A novel acute phase marker in cattle: lipopolysaccharide binding protein (LBp). *Journal* of *Endotoxin Research* 7, 49-52.
- Sheldon, I.M., Noakes, D.E., Rycroft, A. and

Dobson. H. (2001) Acute phase protein responses to uterine bacterial contamination in cattle after calving. *Veterinary Record* **148**, 172-175.

- Skinner, J.G., Brown, R.A.L. and Roberts, L. (1991) Bovine haptoglobin response in clinically defined field conditions. *Veterinary Record* 128, 147-149.
- Smith, B.I., Donovan, G.A., Risco, C., Young, C. and Stanker, L.H. (1998) Serum haptoglobin concentrations in Holstein dairy cattle with toxic puerperal metritis. *Veterinary Record* 142, 83-85.
- Whicher, J.T. and Westacott, C.I. (1992) The acute phase response,pp. 243-269 InWhicher and Evans (Ed.): Biochemistry of inflammation, Kluwer Academic Publishers, London.

Iranian Journal of Veterinary Science and Technology Vol. 3, No. 2, 2011, 57-65

# بررسی ارتباط بین پروتئینهای فاز حاد انتخابی (آمیلوئید سرمی و شیری A و هاپتوگلوبین) در بیماریهای التهابی گاوهای شیری

 $^{*}$ عزت اله فتحى $^{*}$ ، راحله فرحزادى ج

<sup>ا</sup> گروه علوم درمانگاهی دانشکده دامپزشکی دانشگاه تبریز، تبریز، ایران ۲ دستیار تخصصی گروه بیوشیمی بالینی دانشکده پزشکی دانشگاه تربیت مدرس تهران، تهران، ایران

پذیرش نهایی:۱۳۹۱/۱۱/۲۴

#### چکیدہ

هدف از مطالعه حاضر بررسی غلظت سرمی پروتئینهای فاز حاد انتخابی (آمیلوئید سرمی و شیری A و هاپتوگلوبین) و نیز تغییرات بوجود آمده در پارامترهای خونشناختی برخی از بیماریهای التهابی در گاوهای شیری نژاد هولشتاین است. ۵۶ راس گاو شیری هولشتاین که مبتلا به بیماریهای التهابی مختلف از جمله ورم پستان تحت بالینی، عفونت های انگلی خون (تیلریوز و آناپلاسموز)، متریت، پنومونی و جابجایی شیردان بودند در این مطالعه انتخاب شدند (از هر بیماری ۸ راس گاو). تعداد ۸ راس گاو نیز به عنوان گروه کنترل در نظر گرفته شدند. سپس اقدام به خونگیری از ورید گردنی گاوهای مبتلا و سالم گردید. نمونههای شیر اخذ شده از چهار کارتیه پستانی هر دو گروه بهطور جداگانه با هم مخلوط شدند و پس از آن اقدام به انجام آزمایش روی نمونه واحد شیری گردید. نتایج نشان داد که در رابطه با غلظت سرمی و شیری آمیلوئید A بین دو گروه گاوهای مبتلا و سالم و نیز بین جابجایی شیردان و سایر بیماری های عفونی اختلاف معنی داری وجود دارد (20.05). همچنین افزایش قابل ملاحظهای در غلظت سرمی هاپتوگلوبین در گاوهای مبتلا به ورم پستان تحت بالینی، تیلریوز و پنومونی در مقایسه با گاوهای مبتلا و سالم و نیز بین جابجایی شیردان و سایر بیماری های عفونی اختلاف معنی داری اندازه گیری پروتئینهای فاز حاد انتخابی (آمیلوئید سرمی و شیری می می می می می و هرین و سایر بیماری های عفونی اختلاف معنی داری می می و پنومونی در مقایسه با گاوهای مبتلا و آلیم و شیری A و هاپتوگلوبین در گاوهای مبتلا به ورم پستان تحت بالینی، تیلریوز و پنومونی در مقایسه با گاوهای مبتلا به آناپلاسموز، جابجایی شیردان و متریت مشاهده شد (20.05). نتایج مطالعه حاضر کاربرد اندازه گیری پروتئینهای فاز حاد انتخابی (آمیلوئید سرمی و شیری A و هاپتوگلوبین) به عنوان شاخص های از شدان

واژگان کلیدی: پروتئین فاز حاد، سرم آمیلوئید A، هاپتوگلوبین، بیماریهای التهابی

در بافت مقاله: ۱ ۱/۰۹/۰۱

IJVST