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REVIEW ARTICLE

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Foodborne Bacteria in Iran: A 23-year Systematic Review of High-risk Foods

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ABSTRACT

Foodborne diseases are a significant global public health concern due to their high prevalence, mortality, and economic losses. The aim of this study was to conduct a systematic review of identified foodborne pathogens and outbreaks in Iran over the past 23 years to provide an overview of the risk assessment and prevention approaches in the country. Using appropriate keywords and searching major databases, such as ScienceDirect, Scopus, PubMed, Google Scholar, and the Iranian Scientific Knowledge Database, we initially identified 4,740 articles. Finally, 328 articles were selected for evaluation. Among these articles, publications on *Salmonella, Staphylococcus aureus*, and *Listeria* were the most numerous. Poultry meat was found to be the main source of major foodborne pathogens in Iran, including *Campylobacter* (46.21%), *Listeria monocytogenes* (38.45), *Salmonella* (24.83%), and *Yersinia enterocolitica* (16.81%). Given the high prevalence of foodborne bacteria in Iranian foods, it is crucial to implement effective control measures to reduce the risk and burden of foodborne diseases. In particular, poultry meat, which poses a high risk for the occurrence of foodborne diseases in Iran, should be subjected to further risk assessment and control measures throughout the food chain.

Keywords

Foodborne bacteria, Salmonella, Staphylococcus aureus, Listeria, food, prevalence

Abbreviations

EWHO: World Health Organization CDC: Center for Disease Control and Prevention RTE: Ready-To-Eat EFSA: European Food Safety Authority

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Introduction

Foodborne diseases typically result from the consumption of food or water contaminated with pathogens or their toxins [1]. These illnesses often present as acute health problems with diverse symptoms, such as gastrointestinal distress (e.g., diarrhea, vomiting, nausea, and abdominal cramps) or neurological symptoms (e.g., headache, paralysis, and paresthesia) [2, 3]. The bacterial pathogens most commonly associated with foodborne illness worldwide include *Listeria monocytogenes, Escherichia coli* O157:H7, *Staphylococcus aureus, Salmonella enterica, Bacillus cereus, Vibrio spp., Campylobacter jejuni,* and *Clostridium perfringens* [2, 4, 5]. The food products most frequently implicated in outbreaks include poultry, ground meat, seafood, dairy products, as well as fruits and vegetables [6].

The food industry faces significant challenges in ensuring the safety and nutritional quality of food products for consumers due to various sources of contamination, such as animals, soil, water, air, and food handlers during production and storage [7, 8]. However, the implementation of proper cold preservation methods (e.g., refrigeration and freezing) and appropriate thermal processing of foods can effectively prevent foodborne diseases [3].

In the contemporary era, regulatory frameworks and directives pertaining to food safety have been fortified and intensified. Nevertheless, foodborne diseases continue to represent a significant threat to global public health and an economic burden, particularly in developing countries [9]. In its inaugural estimation of the global burden of foodborne diseases in 2015, the WHO attributed 600 million cases of foodborne diseases, 420,000 deaths, and a loss of 33 million years of healthy life worldwide to unsafe food consumption [9].

In 2018, the United States documented 25,606 cases of foodborne infections, resulting in 5,893 hospitalizations and 120 deaths [10]. The burden of foodborne diseases is particularly significant in lowand middle-income countries. Identifying the source of contamination and transmission route is of paramount importance for preventing foodborne illnesses and implementing effective interventions in food safety. However, attributing an infection to specific food and identifying foodborne transmission is challenging and requires source attribution methodologies. Consequently, there is a dearth of studies identifying the sources of foodborne infections, particularly in developing countries [11].

In this study, we aimed to conduct a systematic review of the prevalence of foodborne pathogens in different types of foods in Iran. As a result, we can gain an overview of the role of food in the transmission of infections and emphasize the importance of food safety in controlling foodborne diseases and reducing their health and economic burden on society.

Materials and Methods

Search strategy

A comprehensive and systematic search was conducted in various databases, including ScienceDirect, Scopus, PubMed, Google Scholar, and local Iranian databases, namely the Iranian Scientific Information Database (www.sid.ir). The literature review was limited to studies published during 2000-2023. The keywords used for searching included "prevalence", "detection", and "identification" in conjunction with terms, such as "food", "Iran", "foodborne pathogen", "food infection", "food poisoning", "food illness", "food disease", "foodborne bacteria", "Campylobacter", "Listeria", "Salmonella", "Helicobacter pylori", "Vibrio", "Clostridium botulinum", "Clostridium difficile", "Clostridium perfringens", "Mycobacterium tuberculosis", "Coxiella burnetii", "Staphylococcus aureus", "Shigella", "Pseudomonas", "Bacillus cereus", "Brucella", and "Yersinia enterocolitica".

Eligibility criteria

This systematic review included articles that focused on the prevalence of foodborne pathogens in any type of food in Iran. Duplicate reports and articles without a clear sample size or other essential data were excluded.

Data extraction

Data collection included extracting information, such as the year of publication, types of foods tested for pathogen contamination, sample size, and number of positive samples contaminated with foodborne pathogens.

Results and Discussion

Results and Discussion

Figure 1 illustrates the study selection process presented in the PRISMA diagram. A systematic literature search using Scopus, ScienceDirect, Google Scholar, SID, Magiran, and cross-references yielded an initial total of 4740 articles. After removing duplicates, 1719 articles remained for title/abstract screening. Following this screening, 655 articles were selected for full-text review. Finally, 328 eligible studies were included in the systematic review.



Figure 1. Flow diagram showing the results of search

1) Salmonella spp. prevalence in food

Salmonella (S.) enterica enterica has more than 2300 serotypes, with *S. Enteritidis* and *S. Typhimurium* being the most commonly reported serotypes. Symptoms of salmonellosis include abdominal pain, vomiting, nausea, diarrhea, and fever [12]. Raw meat, particularly poultry, and egg products, are the main sources of foodborne salmonellosis. Other reported foods that transmit *Salmonella* to humans include fish, peanuts, unpasteurized juice, and milk. It is important to cook raw foods thoroughly to a safe minimum internal temperature to prevent foodborne salmonel-

losis, as *Salmonella* is heat-sensitive. However, processed foods, such as RTE meats and salads can become contaminated through cross-contamination during processing [12]. In Europe in 2020, 0.15% of RTE food samples and 2.4% of non-RTE food samples were positive for *Salmonella* [13].

Table 1 presents the prevalence of *Salmonella* in different foods in Iran based on our review. The highest levels of contamination were found in poultry meat (23.03%), followed by red meat (14.13%), dairy products (11.66%), RTE foods (11.34%), eggs (9.93%), vegetables (7.8%), fish and shrimp (5.93%), raw milk (3%), and water (2.25%) (Figure 2). In a study conducted in China in 2019, out of 1035 different food samples, a total of 147 samples (14.2%) were positive for Salmonella. In their study, the highest prevalence of Salmonella was found in fresh meat samples (28%), followed by RTE foods (9%), frozen foods (7.1%), and fresh produce (4.5%) [14]. Fresh meat is a common source of Salmonella contamination due to the nature of its production and processing [15]. During the slaughter and processing of animals, there is a high risk of cross-contamination with various bacteria, such as Salmonella [16]. In addition, fresh meat products consumed raw or undercooked increase the risk of foodborne illness [17]. The handling and storage of fresh meat products can also contribute to Salmonella contamination [18]. In contrast, RTE foods and frozen foods undergo processing and packaging that can reduce the risk of Salmonella contamination [19]. However, it is still possible for Salmonella to be introduced during the processing or packaging of these products [20]. Fresh produce, while less likely to be contaminated with Salmonella compared to fresh meat, can still pose a risk if not properly handled and washed before consumption [18].

2) Staphylococcus aureus prevalence in food

Although *Staphylococcus* (S.) *aureus* is the primary causative agent of hospital and community-acquired infections, it has also been associated with foodborne diseases. *S. aureus* can cause various gastrointestinal illnesses, which are characterized by nausea, vomiting, abdominal cramps, weakness, and diarrhea [21]. Table



Figure 2.

Prevalence of Salmonella spp. in different foods in Iran.

Table 1.

Summary of the studies reporting the prevalence of Salmonella spp. in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	Type of Salmonella spp.	Referenc- es	City
1	2023	Eggs	40	4	10	Salmonella Enteritidis	[72]	Qazvin
2	2023	poultry meat	440	39	9	Salmonella enteritidis and Salmonella typhimurium	[73]	Shahrekord
3	2022	Eggs	500	405	81	Salmonella spp	[74]	Lahijan
4	2022	retail raw meat	60	16	26	Salmonella spp.	[75]	Urmia
5	2022	poultry products	80	11	13.75	Salmonella spp.	[76]	Ardabil
6	2022	Eggs duck	130	21	16.6	Salmonella spp	[77]	Qazvin
7	2022	Chicken meat	150	4	2.7	Salmonella spp.	[78]	Zahedan
8	2022	cattle raw milk	100	2	2	Salmonella spp.	[79]	Mazandaran
9	2022	Red meat	300	35	11	Salmonella spp	[80]	Shahrekord
10	2021	Chicken meat	100	6	6	Salmonella spp.	[81]	Ardabil
11	2021	poultry and egg	3125	250	8	Salmonella spp.	[82]	six province of Iran
12	2021	chicken meat, beef	450	40	8	Salmonella enterica	[83]	Tehran
13	2019	Olivier salad	26	6	23	Salmonella spp.	[84]	Mashhad
14	2019	Raw chicken meat Egg yolk	60 30	29 0	48.3 0	Salmonella spp.	[85]	Karaj
15	2019	Eggs (shell & contents)	525	0	0	Salmonella spp.	[86]	Isfahan
16	2019	Traditional cheeses	100	0	0	Salmonella spp.	[87]	Mohabad
17	2018	Pastry	75	5	6.6	Salmonella spp.	[88]	Mashhad
18	2018	Chicken Meat	100	7	7	Salmonella spp.	[89]	Mashhad
		Industrial eggs (shell & con- tents)	60	0	0	_		
19	2018	Local eggs shell	60	0	0	Salmonella spp.	[90]	Zanjan
		Local egg con- tents	60	2	1.6			

Table 1 cont.

lable I	cont.							
		Industrial eggs (shell & con- tents)	34	0	0			
20	2018	Local eggs shell	42	6	4.2	— Salmonella spp.	[91]	Sanandaj
20	2018	Local egg con- tents	42	2	4.7	— Sumoneuu spp.	[2]]	
		Bulk eggs shell	40	3	7.5	_		
		Bulk egg con- tents	40	0	0			
21	2018	Industrial eggs (shell & con- tents)	60	0	0	Salmonella spp.	[92]	Qazvin
22	2018	Shrimp	245	33	13.4	Salmonella spp.	[93]	Khuzestan
		Pizza	90	8	8.8			
23	2017	Frankfurter	90	22	24.4	S. arizonae	[94]	Ilam
		Sausages	90	19	21.1			
24	2017	Camel meat	150	0	0	S. Typhimurium	[95]	Kohgiluyeh & Boyerahmad/ Isfahan
25	2017	Hamburgers	100	2	2	Salmonella spp.	[96]	Kashan
	2016	beaf meat	190	7	3.6	S. Typhimurium	[07]	Cilur
26	2016	Poultry meat	190	15	7.8	S. Typhimurium	[97]	Gilan
27	2016	Poultry meat	183	52	28.4	S. Typhimurium	[98]	Zanjan
28	2016	Industrial Olovier salad	48	0	0	— Salmonella spp.	[99]	Isfahan
	2010	Traditional Olovier salad	54	11	20.4		[]	
29	2016	Traditional ice cream	90	62	68.8	Salmonella spp.	[100]	Zabol
30	2015	Pastry cream	120	0	0	Salmonella spp.	[101]	Arak
31	2015	Olivier salad	50	9	18	S. Ttypimurium	[102]	Shahrekord
22	2015	Poultry meat	625	35	5.6		[102]	
32	2015	Eggs	625	25	4	— S. Enteritidis	[103]	Tehran
33	2015	Eggs	50	5	10	S. Enteritidis	[104]	Shiraz
34	2015	Native eggs	64	0	0	Salmonella spp.	[105]	Yasuj
		Traditional	200	34	17		_	
35	2015	& Industrial – cheese		11	5.5	— Salmonella spp.	[106]	Shahrekord
		Eggshells	150	2	1.3	S. Enteritidis &		
36	2015	Egg contents	150	0	0	S. Typimurium	[107]	Tabriz

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Table 1	cont.							
37	2014	Meat Products (Burgers, Sau- sages, Kababs, Cutlets	42	0	0	Salmonella spp	[108]	Urmia
38	2014	Unpasteurized cream	100	2	2	S. Paratyphi B	[109]	Tehran
		Chicken	190	86	45	Salmonella spp.		
•		Beef meat	189	38	20.2	Salmonella spp.		-
39	2014	Beef meat	189	19	10	S. thompson	[110]	Tehran
		Chicken	190	65	34.2	S. thompson		
		Chicken meat	200	58	29			
40	2014	Liver	120	26	21.6	Salmonella spp.	[111]	Alborz
		Heart	120	17	14.1	_		
		Poultry meat	89	28	31.6			
41	2014	Beef meat	98	26	26.1	— Salmonella spp.	[112]	Shahrekord
42	2013	Olovier salad	50	9	18	S. Typhimurium	[112]	Shahrekord
10	2012	Tap water	144	5	3.4			
43	2013	304	304	5	1.1	— Salmonella spp	[113]	Isfahan
44	2013	Poultry slaugh- tered	250	7	2.8	S. infantis	[114]	Brijand
45	2013	Local eggs	210	14	66.6	Salmonella spp.	[115]	Kohgiluyeh & Boyerahmad
		Industrial egg contents	100	0	0			
		Industrial egg- shells	100	19	19			Talesh
		Local egg con- tents	100	0	0		[116]	
46	2013	Local eggshells	100	4	4	Salmonella spp.		
		Local chicken meat	100	21	21	_		
		Industrial chick- en meat	100	5	5	_		
		Red meat	150	5	3.3	_		
		Industrial						
47	2013	Olovieh salad	200	0	0	Salmonella spp.	[117]	Yazd
48	2012	Seafood	384	19	5	Salmonella spp.	[118]	Bushehr, Hormozgan, Khuzestan
		Chicken meat	150	14	9.3			
49	2012	Turkey meat	105	7	6.7	Salmonella spp.	[119]	Isfahan &
		Ostrich meat	45	1	2.2		-	Shahrekord
				7	11.6	Salmonella spp.		
50	2012	Beef meat	60	4	6.6	S. Typhimurium	[120]	Sanandaj
		Packed chicken meat	96	19	19.7	· · ·		
	2012					— Salmonella spp.	[121]	Mazandaran

Table 1 cont.

Table 1 cont.

able I	cont.							
		Salt water fish	70	2	2.9			
52	2012	Shrimp	70	3	4.3	Salmonella spp.	[122]	Ahvaz
		Shrimp burge	10	1	10			
53	2011	Raw cow's milk	350	14	4	Salmonella spp.	[123]	Shahrekord
54	2010	Egg	100	0	0	Salmonella spp.	[124]	Shahrekoro
55	2010	Chicken meat	190	86	45	- Salwayalla att	[125]	Tehran
55	2010	Beef meat	189	38	20	— Salmonella spp.	[125]	Tenran
		Turkey meat	144	14	9.7			
56	2010	Ostrich meat	65	3	4.6	Salmonella spp.	[126]	Isfahan
		Partridge meat	40	0	0			
	2000	Eggshells	250	4	1.6	S. Typhimurium	[107]	Mahhad
57	2009	Egg contents	250	0	0	Salmonella spp.	[127]	Mashhad
		Poultry car-	60	5	8.3	Salmonella spp.		
58	2009	- casses	60	1	1.6	S. Typhimurium	[128]	Mashhad
		Egg contents	120	0	0			
59	2009	Eggshells	120	68	56.6	Salmonella spp.	[129]	Zanjan
		Chicken meat	120	104	86.6	_		
		Chicken meat	67	32	47.7			
60	2009	Beef meat	66	19	28.7	— Salmonella spp.	[130]	Tehran
61	2008	Local egg con- tents	500	1	0.2	Salmonella spp.	[131]	Birjand
		Local eggshells	500	2	0.4			,
		Raw poultry	134	24	17.9			
		Cooked poultry	56	3	5.3	_		
		Turkey	3	1	33.3	_	[132]	
		Quail	5	2	40			
		Red meat	101	8	7.9			
		Cooked meat	118	2	1.6			
62	2008	Vegetables	38	3	7.8	Salmonella spp		Isfahan
		Fish	15	0	0	_		
		Yogurt	32	0	0	_		
		Olovieh salad	20	0	0	_		
		Hamburger	5	0	0	_		
		Mayonnaise souse	8	0	0			
63	2007	Poultry car- casses	132	92	69	Salmonella spp.	[133]	Tehran
64	2007	Traditional cheeses	200	0	0	Salmonella spp.	[134]	Jahrom
		Liver	145	12	8.1			
65	2006	Meat (before chiller)	145	28	18.4	Salmonella spp.	[135]	Yazd
55		Meat (after chiller)	145	50	34.4	_		

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Table 2.

Summary of the studies reporting the prevalence of *S. aureus* in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	City
1	2023	Poultry meat	94	16	17	Shahrekord
2	2022	Sausages and Bologna	100	31	31	Tehran
3	2022	Raw and ready-to-eat green leafy vege- tables	366	134	36.6	Tehran
4	2022	Retail raw meat	60	23	39	Urmia
5	2022	Raw Milk And Traditional Dairy	150	23	15.33	Alborz
6	2022	Ready To Eat Food	320	10	3.12	Tehran
7	2022	Raw Milk	380	42	11.05	Alborz
8	2022	Raw Cow Milk	90	35	38.88	Shahrekord
9	2021	Baghlava	112	3	2.67	Qazvin
10	2021	Raw milk	250	46	18.4	Mashhad
11	2021	Cheese, raw and pasteurized milk	100	10	10	-
12	2019	Meat Products	160	26	16.25	Shahrekord
13	2021	Milk and Cheese	200	23	11	Khuzestan
14	2021	Meat retail	90	31	34.5	Zanjan
15	2021	Ready-to-eat food	415	64	15.42	Tehran
16	2021	Chicken meat	24	6	25	Tehran
17	2019	Zoolbia & Bamieh	75	21	28	Mashhad
18	2019	Fowl meat	240	22	9.6	Tehran
19	2019	Traditional cheese	100	21	21	Maragheh
20	2018	Traditional cheese	100	45	45	Mahabad
		Raw foods with animal origin	84	20	23.8	-
21	2017	Cooked foods with animal origin	132	12	9	Isfahan
		Cooked foods without animal origin	269	15	5.7	
		Season salad	18	0	0	-
22	2017	Pasta salad	5	2	40	- Bandar abbas
		Lettuce	16	0	0	-
		Shirazi salad	7	0	0	

Table 2 cont.

able 2	cont.					
		Pizza	90	11	12.2	
23	2017	Frankfurter	90	25	27.7	Ilam
		Sausages	90	22	24.4	
24	2016	Cheese	120	18	41.6	Hamedan
25	2016	Meat	380	78	20.5	Gilan
		Shrimp	300	84	28	Persian Gulf,
26	2016	Fish	300	122	47	Caspian Sea
		Red meat	379	36	9.4	
27	2015	Dairy products	671	62	7.2	— Hamadan
		Raw milk	320	88	27.5	
28	2015	Dairy products	350	87	24.8	Chaharmahal va Bakhtiyari
				0,	21.0	Persian Gulf,
29	2015	Shrimp	300	74	24.6	Tehran
		Raw milk	1930	248	12.8	Tenran
30	2015		720	80		— Mazandaran
		Dairy products			11.1	
31	2015	Bovine milk	92	44	47	Maku
		Sheep milk	86	32	37	
32	2 2015 -	Industrial Olivier salad	30	15	50	- Shahrekord
		Traditional Olivier salad	20	8	40	
33	2015	Cheese	80	80	100	Marand
34	2015	Chicken nuggets	420	24	5.7	Isfahan
35	2015	Different food	606	12	1.9	Gilan
36	2014	Cream pastry	450	194	43.3	Gorgan
		Milk	100	9	9	
37	2014	Cheese	100	45	45	— Tabriz
		Traditional ice cream	30	2	6.7	
38	2014	Olovier salad	4	0	0	Yasuj
		Cream suit	30	9	30	_
39	2014	Raw milk	300	125	41.6	Ahwaz
40	2014	Dairy product	460	127	27.6	Marand
41	2014	Cheese	80	80	100	Tehran, Gilan
42	2014	Doogh	126	86	68	,Mazandaran

Prevalence of Foodborne Bacteria in Food in Iran

Iubic 2	cont.						
43	2014	Raw milk	120	49	40.8	Kurdistan	
44	2014	Meat products	150	19	12.6	Tonekabon	
45	2014	Traditional cheeses	100	16	16		East- Azer- baijan
46	2013	Dairy products	347	20	5.8		Isfahan, Chaharmaha va Bakhtyari, Khuzestan,
47	2013	Industrial Olivier salad	200	40	20	[117]	Yazd
48	2013	Milk	200	22	11	[175]	Fars
40	2012	Traditional white cheese	100	26	26	[1=4]	
49	2012	butter	150	24	16	- [176]	Tabriz
		Ground-meat kebab	72	72	100		
		Bakkhtiyari Kebab	72	72	100	-	
50	2012	Fish	72	72	100	- [177]	Shahrekord
		Salad	72	72	100	-	
51	2012	Seafood products	245	22	8.9	[122]	Different ma kets of Iran
52	2012	Packaged hamburger	256	64	25	[178]	Tehran
		Raw milk	100	50	50		
53	2012	Pasteurized milk	100	2	2	[176]	Urmia
		Ice cream	100	26	26	_	
54	2012	Raw milk	348	46	13.2	[179]	Shahrekoro
55	2010	Fruit juice	360	32	8.8	[180]	Shahrekor
56	2008	Different food	216	30	55.6	[181]	Tehran
57	2006	Fresh fish	67	15	22.3	[182]	Gilan, Caspian Sea

Table 2 cont.

2 presents the findings of studies conducted in Iran regarding the prevalence of this pathogen in different food categories, including seafood (38.51%), meat products (35.47%), dairy products (31.70%), red meat (25.85%), RTE foods (23.59%), raw milk (23.32%), and poultry meat (14.32%) (Figure 3). Seafood and fish are conducive to microbial growth due to their abundant protein and water content. *S. aureus* is not typically found in the natural microflora of fish, there-

fore, its presence can indicate poor personal hygiene, new contamination, or potential disease in the fish [22]. Improper conditions in the fishery, storage, and non-standard transportation provide conditions for pathogens to grow [23]. Furthermore, the hot climate in Iran can facilitate the growth and proliferation of *S. aureus* bacteria in food products, such as meat and dairy items, particularly if they are not stored and refrigerated correctly [24].



Figure 3.

Prevalence of S. aureus in different foods in Iran.

Table 3.
Summary of the studies reporting the prevalence of <i>Listeria</i> spp. in Iran.

	Year	Sample type	Sample size	Positive sam-	Prevalence	Type of	References	Area
		ownipie type	- I	ples (N)	(%)	Listeria spp.		
1	2023	Various Food	900	136	15.1	L. monocytogenes	[183]	Mazandaran and Goles- tan
2	2022	Retail raw meat	60	25	42	L. monocytogenes	[75]	Urmia
3	2022	Curd and cheese	150	14	9.33	L. monocytogenes	[184]	-
4	2021	Non-Pasteurized Milk	50	30	60	L. monocytogenes	[185]	Kerman
5	2021	Traditional cheeses	60	1	1.6	L. monocytogenes	[186]	Tehran
6	2021	Raw Milk	100	10	10	Listeria spp.	[187]	Tehran
7	2021	Seafood	350	40	11.42	Listeria spp.	[188]	Genaveh port
8	2020	Beef and chicken meat	90	45	50	L. monocytogenes	[189]	Zanjan
9	2019	Eggs	525	0	0	Listeria spp.	[86]	Isfahan
10	2019	Chicken meat retailers	811	257	30.5	Listeria spp.	[190]	Mashhad
11	2010	Traditional dairy	E 4 E	64	11.7	Listeria spp.	[101]	V J
11	2018	products	545	22	4.3	L. monocytogenes	- [191]	Yazd
12	2017	Food (sausage, milk, cheese, chicken and meat)	267	8	2.9	Listeria spp.	[192]	Urmia
13	2017	Fresh chicken carcass- es	200	80	40	Listeria spp.	[193]	Mashhad
14	2016	Dairy products	107	9	8.4	L. monocytogenes	[104]	TT 1
14	2016	Processed meat	210	11	5.2	Karaj &	- [194]	Tehran
15	2016	Seafood	237	7	2.9	L. monocytogenes	[195]	Tehran

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Tabl	e 3 cont.							
16	2016	Argyrosomus	240	30	12.5	Listeria spp.	[196]	Isfahan& Bandaran-
-		hololepidotus		5	16.6	Bandaranzali	[]	zali
17	2015	Koozeh cheeses	100	3	3	L. monocytogenes	[197]	Urmia
1.0	2015	Minaadhaaf	150	4	2.7	Listeria spp.	[109]	Ahvaz
18	2015	Minced beef	150	1	0.6	Mazandaran	[198]	Alivaz
19	2015	Raw fish	488	104	21.3	Listeria spp.	[199]	Mazandara
20	2015	Raw milk	60	0	0	L. monocytogene	[200]	Zanjan
	0015	Traditional dairy				.	[224]	
21	2015	products	292	21	19.7	Listeria spp.	[201]	Isfahan
22	2015	Raw milk	100	5	5	L. monocytogene	[202]	Kerman
23	2014	Ready to eat food (olovier salad, Yogurt stew, macaroni salad and meat salad)	235	20	8.5	Listeria spp.	[203]	Shahrekor
24	2014	Meat products	98	12	32.4	L. monocytogene	[204]	Qazvin
.4	2014	Milk products	84	25	29.7	Fars & Khuzestan	[204]	Qazviii
25	2014	Bulk milk	260	27	10.4	Listeria spp.	[205]	Fars & Kh
	2014	Duk mik	200	7	2.7	Bandar anzali	[205]	estan
	Smoked fish	80	7	8.8	Listeria spp.			
6	5 2014		00	2	2.5	L. monocytogene	[206]	Isfahan 8
		Salted Fish	40	6	15	Listeria spp.	[]	Bandar anz
				1	2.5	L. monocytogene		
7	2013	Meat and meat prod-	60	8	13.3	Listeria spp.	[207]	Khoramabad
-		ucts		2	6.6	L. monocytogene	[· ·]	& Tehrar
8	2013	Crayfish meat	40	3	7.5	L. monocytogene	[208]	Aras
9	2013	Raw cow milk	986	25	2.5	Listeria spp.	[209]	Isfahan
-				20	2	L. monocytogene	[]	
0	2013	Vegetables and ready	300	26	8.7	Listeria spp.	[210]	Tehran
0	2015	mayonnaise salads	500	21	7	L. monocytogene	[210]	
1	2013	Raw seafood products	331	16	4.8	L. monocytogene	[211]	Shahrekoi
1	2013	RTE seafoods	321	46	14.5	L. monocytogene	[211]	
2	2013	Raw milk	466	83	18.6	Listeria spp.	[212]	Tehran
		Dairy products	185	7	3.8	_		
3	2013	Meat products	187	51	27.2	Listeria spp.	[213]	Kermansh
		Ready-to-eat foods	158	8	5.1			
4	2013	Seafood	300	24	8	Listeria spp.	[214]	Isfahan &
				18	6	L. monocytogene		Shahrekord
5	2013	Quail products	150	10	6.6	Listeria spp.	[215]	Isfahan
		- 			0.6	L. monocytogene	[24.6]	
6	2013	Lamb	200	5	2.5	L.ivanovii	[216]	Shahreko
7	2012	Different types of raw meat	1107	27	12.7 2.4	Listeria spp.	[217]	Shahrekord Isfahan, Ahvaz ,Shi-
0	2012		402	104		Listavia	[210]	raz,Yazd,
38	2012	Poultry product	402	134	33.3	Listeria spp.	[218]	Shahrekor Isfahan &
39	2012	Seafood	264	20	7.6	Listeria spp.	[219]	Shahrekor

Table 3 cont.

Table	5 cont.							
40	2012	Various seafood prod- ucts	245	2	0.8	L. monocytogene	[122]	Different mar- kets of Iran
41	2011	Eggs	100	0	0	L. monocytogene	[124]	Shahrekord
42	2011	Fish	194	24	12.3	Listeria spp.	- [220]	Urmia
42	2011	F1811	194	5	2.5	L. monocytogene	[220]	Urmia
		Raw cow milk	45	5	1.1	Listeria spp.	_	
			45	2	4.4	L. monocytogene	_	
		Raw goat milk	20	1	3.1	Listeria spp.	_	
12			32	1	3.1	L. monocytogene	- [221]	Shiraz
43		Traditional cheese	41	10	24.4	Listeria spp.	- [221] -	Siiifaz
			41	4	9.7	L. monocytogene		
		Traditional ice-cream	60	8	11.7	Listeria spp.	_	
			00	2	3.3	L. monocytogene		
44		Raw milk	100	4	4	L. monocytogene	[222]	
45		Dairy products	360	6	1.6	L. monocytogene	[223]	
		Chilled ready to eat foods	41	3	7.3	L. monocytogene		
46		Meat, meat products	332	4	1.2	L. monocytogene	[224]	
		Milk and dairy prod- ucts	88	0	0	L. monocytogene	_	
47		Cattle carcasses	203	6	3	L. monocytogene	[225]	

Table 3 cont.

3) Listeria monocytogenes prevalence in food

Listeria (L.) *monocytogenes* represents a significant public health concern due to its ability to be transmitted from the environment to food, which can lead to foodborne listeriosis in humans [25]. In 2020, the EFSA reported a total of 1876 cases of listeriosis, with 97.1% of these cases necessitating hospitalization [26]. Moreover, the EFSA indicated an increase in the case fatality rate and hospitalization rate associated

with *L. monocytogenes* infections in 2020. Among all the reported zoonoses in Europe in 2020, listeriosis had the highest case fatality rate of 13% [26]. Those at the greatest risk of developing listeriosis include pregnant women, the elderly, newborns, and patients with compromised immune systems [27]. Moreover, a multitude of food items were identified as potential sources of listeriosis outbreaks during this period. Specifically, 4.8% of RTE meat products and 0.44% of



Figure 4. Prevalence of *Listeria* in different foods in Iran.

Table 4

Summary of the studies reporting the prevalence of C. burnetii in Iran..

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	References	Area
1	2022	Unpasteurized Camel Milk	100	6	6	[226]	Mashhad
2	2022	Kope cheese and cattle milk	800	103	12.87	[227]	West Azerbaijaı
3	2021	Raw Milk	162	23	14	[228]	Tehran, Hamadan, and Mazandaran
4	2021	Raw milk	100	27	27	[229]	Mazandaran
5	2020	Raw milk	204	21	10.2	[230]	Gilan
6	2020	Sheep and goats milk	420	51	12.1	[231]	West Azerbaija
7	2019	Milk	126	44	34.9	[232]	Qom
		Milk			16.9		
8	2019	Cattle milk	840	14	14.4	[233]	West Azerbaija
		Buffalo milk		-	19.3	-	,
		Cream & butter	200	6	3		
		Traditional bovine cream,	69	4	5.7	-	
)	2019	Traditional sheep butter,	20	1	5	- [234]	Shahrekord
		Traditional bovine butter	39	1	2.5	-	
		Raw milk		9			
.0	2018		500	3	1.8	[235]	Khorramabad
10	2010	Sheep milk	500	6	1.0	[255]	Kilorrainabad
		Goat milk					
11	2018	Bulk milk	100	3	3	[236]	Shiraz
12	2018	Non-pasteurized dairy products	238	20	8.4	[29]	Shiraz
1.2	2019	Raw milk	100	10	10	- [227]	T-1
13	2018	Traditional unpacked cheese	40	3	7.5	- [237]	Tehran
14	2016	Raw sheep milk	72	15	20.8	[238]	Khorramabad
				12	17.4		
5	2015	Bovine bulk milk	70	7	10	[239]	Jahrom
				7	10	-	
		Individual raw milk	60	7	11.6	_	
6	2015	Bovine	38	5	8.3	[200]	Zanjan
		Ovine	22	2	3.3	_	
17	2015	Cow milk	150	18	12	[240]	Tehran
18	2015	Goat milk	31	5	16.1	[241]	Kerman
19	2015	Cow milk	80	20	25	[242]	Ajabshir
		Cheese	28	2	7.1		
20	2014	Yoghurt	26	2	7.6	- [242]	Mashhad
20	2014	Sheep milk	23	8	34.7	[243]	wasiinad
		Cow milk	60	2	3.3		
21	2014	Bovine bulk tank	100	5	5	[244]	Mashhad
22	2014	milk	51	21	41.1	[245]	Khoramabad.
23	2013	Goat milk	100	14	14	[246]	Qom

Table	4 cont.						
24	2013	Bovine bulk milk	100	11	11	[247]	Jahrom
		Bovine milk	247	8	3.2		
25	2011	Ovine bulk milk	140	8	5.7	[240]	V
25	2011	Caprine bulk milk				- [248]	Kerman
		Camel bulk milk	110	5	4.5	-	
26	2010	Bulk milk	296	6	2	[249]	Fars, Ghom, Kerman, Yazd Khuzestan
		Cow milk	210	13	6.2		
27	2010	Sheep milk	110	0	0	[250]	Chaharmahal va Bakhtiari
		Goat milk	56	1	1.8	-	Durditiul1

Tabla	1	cont

milk and milk products were found to be contaminated with L. monocytogenes [26]. Table 3 and Figure 4 present the findings of studies conducted in Iran regarding the prevalence of *L. monocytogenes* in various food types. As illustrated in Figure 4, poultry meat exhibited the highest contamination rate of 38.45%, followed by meat products (14.94%), red meat (13.45%), raw milk (9.77%), dairy products (9.48%), seafood (8.75%), and RTE foods (7.32%) (Figure 4). A previous review study conducted in Iran until 2015 yielded comparable results regarding the contamination of food with Listeria. The highest prevalence of L. monocytogenes was approximately 9.2%, which was observed in RTE foods [25]. Therefore, RTE foods should be considered a potential hazard to consumers [25]. Similarly, other developing countries have also yielded comparable results. For example, a study conducted in Ethiopia revealed that 28.4% of raw milk and milk products were contaminated with Listeria spp., with 5.6% of these samples testing positive for L. monocytogenes [23].

4) Coxiella burnetii prevalence in food

Coxiella burnetii is a zoonotic pathogen that causes Q fever in humans and coxiellosis in livestock. Cattle, goats, and sheep serve as the primary reservoirs for the pathogen, facilitating its transmission to humans [28]. The primary routes of human infection are through the inhalation of contaminated aerosols or the consumption of unpasteurized milk and dairy products [29]. In Europe, 523 cases of Q fever were identified in 2020, resulting in a case fatality rate of 2.1% [30]. Table 4 presents the results of studies conducted in Iran concerning the prevalence of C. burnetii in different food items. As illustrated in Figure 5, the foods with the highest contamination rates were raw milk (12.36%) and dairy products (6.40%). C. burnetii is a bacterium that causes Q fever, a zoonotic disease that can be transmitted from animals to humans. In numerous rural regions of Iran, milk is still produced and processed using traditional methods that fail to meet the requisite modern hygiene standards [31]. The absence of adequate hygiene protocols in milk



Figure 5. Prevalence of *C. burnetii* in different foods in Iran.

Prevalence of Foodborne Bacteria in Food in Iran

production and processing facilities may result in the contamination of milk with *C. burnetii*. Moreover, the proximity of animals to humans in the rural areas of Iran contributes to the high levels of contamination of raw milk and dairy products with *C. burnetii* [32]. Animals, such as cows and goats, can carry the bacterium and shed it in their milk, which can then be transmitted to humans through consuming contaminated dairy products [33].

A study conducted in Italy in 2017 reported that 15% of milk samples were contaminated with *C. burnetii*, with a higher prevalence of contamination in bovine milk (41%) compared to sheep milk (12%) [34]. In Brazil, in 2020, 9.43% of cheese samples (out of 53 samples) were positive for *C. burnetii* DNA [35]. Another research in the United States reported that 94% of bulk milk samples from dairy herds were contaminated with *C. burnetii* [36]. Our review indicates that the data from Iran align with the reports from other countries. However, it should be noted that the prevalence of *C. burnetii* contamination varies depending on the type of dairy products, including specific variations within milk.

5) Bacillus cereus prevalence in food

Bacillus cereus spores are a well-documented contaminant of food that can survive high temperatures during cooking and pasteurization [37]. This bacterium is associated with two distinct types of gastrointestinal diseases: the emetic (vomiting) syndrome and the diarrheal syndrome [38]. In Europe, 835 cases of foodborne illness caused by *B. cereus* were reported in 2020, with a hospitalization rate of 1.2% and a mortality rate of 0.1% [30]. The diarrheal syndrome is typically attributed to the consumption of contaminated foods, including raw and cooked beef, meat products, fish, poultry, soups, sauces, stews, milk, and vegetables. In contrast, the emetic syndrome is associated with the consumption of a toxic dose of the pre-formed emetic (cereulide) toxin produced by B. cereus in starchy foods, such as rice, pasta, noodles, potatoes, bread, pastries, and sesame products [39]. Table 5 presents the results of studies conducted in Iran regarding the prevalence of B. cereus in different food items. As illustrated in Figure 6, the highest prevalence of B. cereus contamination was observed in rice (100%), followed by raw milk (48.8%), poultry meat (42.17%), spices (42%), infant food (32.62%), dried vegetables (31.42%), meat products (11.16%), red meat (9.33%), and dairy products (8.9%) (Figure 6). In Australia, B. cereus contamination was identified in a variety of food samples, including uncooked pizza bases (1.58%), cooked pizzas (4.57%), processed meats (0.28%), cooked meat pies (4.45%), cooked sausage rolls (3.26%), and raw diced chicken (5.45%) out of 1,263 retail food samples [40]. In China, B. cereus contamination was observed in 50% of rice and noodle samples, 34% of cooked meat samples, and 22% of cold vegetable dishes [41]. In Poland, the highest prevalence of B. cereus contamination was found in herbs and spices, with a rate of 63.3%. Moreover, other food items, including breakfast cereals, pasta, rice, pasteurized milk, infant formulas, as well as fresh and ripening cheeses, were also found to be contaminated with B. cereus [37].



Figure 6.

Prevalence of *B. cereus* in different foods in Iran.

Table 5.

Summary of the studies reporting the prevalence of B. cereus in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	References	Area
1	2023	Various Spices	200	84	42	[251]	Isfahan
		Individual meat	200	29	14.5		
		Raw lamb					
2	2020	Raw beef	60	7	3.5	[252]	Zanjan
		Cooked beef	60	10	5		
		Cooked beer	80	12	6		
3	2020	Traditional dairy products	150	16	10.6	[253]	Tabriz
4	2019	Different rice	10	10	100	[254]	Zanjan
5	2018	Cow's raw milk	120	13	10.8	[255]	Tabriz
6	2018	Dried vegetable	140	44	31.4	[256]	Tehran
7	2018	Cream	62	0	0	[257]	Zanjan
8	2017	Different types of beef burgers	80	18	22.5	[258]	Tehran
9	2017	Milk-based infant food	300	9	3	[259]	
10	2017	Powdered infant formula milk	125	84	67.2	[260]	
11	2017	Cow milk	42	41	97.6	[261]	
12	2016	Cheese	200	10	5	[262]	
13	2016	Beef meat and poultry	380	44	11.8	[97]	
14	2016	Dairy products	230	46		[263]	
		Poultry meat foods Frozen	104	80	76.5		
15	2015	Semi cooked	39	9	24.2	[264]	Tehran
		Refrigerated	46	26	[97]		
16	2014	Rice	408	408	100	[265]	Urmia
17	2013	Infant foods	200	84	42	[266]	Isfahan
18	2012	Kefir type drinks	32	9	28	[267]	
10	2012	Pasteurized milk	32	12	Tehran	[207]	
19	2007	Infant formula	60	11	18.3	[268]	Tehran

6) Yersinia enterocolitica prevalence in food

In Europe, 236 cases of foodborne yersiniosis were reported in 2020, with 4.7% of cases necessitating hospitalization [30]. *Yersinia enterocolitica* contamination has been documented in a variety of foods in Europe, including red meat (beef, pork, and lamb), poultry, seafood, eggs, milk and milk products, bean sprouts, vegetables, tofu, and stewed mushrooms [42]. Table 6 presents the results of studies conducted in Iran regarding the prevalence of *Y. enterocolitica* in different food items. As illustrated in Figure 7, poultry meat exhibited the highest contamination rate of 16.81% in Iran. This was followed by raw milk (11.93%), red meat (11.63%), and dairy products (10%) (Figure 7). In Europe, 5.2% of RTE meat was found to be positive for *Yersinia* in 2020, which is a relatively high and concerning rate [30]. A study conducted in Argentina in 2019 reported chicken (12.4%) and bovine-originated foods (10.2%) as the most contaminated foods with *Y. enterocolitica* [43], which aligns with the findings in Iran. However, the latter study reported a lower prevalence of contamination in dairy products (0.7%) compared to the findings in Iran [43]. The elevated contamination rates of *Y. enterocolitica* in poultry meat observed in Iran and Argentina can be attributed to several factors, including the hygiene practices employed during the processing, transportation, and

Table 6.

Summary of the studies reporting the prevalence of *Y. enterocolitica* in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	References	Area
1	2022	Raw Milk	360	3	0.83	[269]	Tehran
2	2021	red meat	200	26	13	[270]	Shiraz
3	2021	Bovine Raw Milk	100	33	33	[271]	Mashhad
4	2021	Traditional Cheeses	200	38	19	[272]	Khorasan Razavi and Golestan
5	2020	Raw milk	360	3	0.8	(Soltan Dallal, 2020)	Tehran
6	2019	Cheeses	200	38	19	- [273]	Khorasan Razavi
0	2019	Raw milk	100	33	33	[2/3]	and Golestan
7	2018	Chicken meat	100	25	25	[274]	Mashhad
8	2018	Raw milk (sheep & goats)	100	9	9	[275]	Shahrekod
9	2018	Turkey meat	300	55	18.3	[276]	Shahrekord
		Meat	450	56	12.4		
10	2015	Chicken meat	226	35	15.4	[277]	Tehran
		Beef meat	224	21	9.3	_	
11	2015	Raw milk	446	19	4.3	[278]	Varamin
12	2014	Dairy products	552	28	5	[279]	Isfahan
13	2014	Raw chicken meat	300	65	21.6	[280]	Shahrekod
14	2014	Unpasteurized cream	100	3	3	[281]	Tehran
15	2013	Chicken meat	720	132	18.3	[282]	Shahrekod
16	2012	Bulk raw milk	354	8	2.6	[202]	Eastern Azer-
10	2012	Cheeses	200	8	4	- [283]	baijan
17	2012	Chicken meat	200	18	9	[121]	Mazandaran
18	2012	Broiler meat	120	19	15.8	[284]	Tabriz
19	2011	Beef and chicken meat	379	48	12.6	[285]	Tehran



Figure 7. Prevalence of *Y. enterocolitic*a in different foods in Iran.

storage of these products [44]. Poultry meat has been identified as a significant source of *Y. enterocolitica* contamination due to the presence of the bacterium in the intestinal tracts of birds [45]. Inappropriate handling and processing of poultry can result in the cross-contamination of the meat with *Y. enterocolitica*. In addition, raw milk, red meat, and dairy products can serve as reservoirs for *Y. enterocolitica* if not properly pasteurized or handled [46].

7) Campylobacter prevalence in food

Campylobacter spp. has been identified as the leading cause of foodborne gastroenteritis in Europe since 2005 [30]. In addition to acute gastroenteritis, *Campylobacter*

infections can also result in chronic manifestations in humans [47]. Among the various species within the genus Campylobacter, *C. jejuni* and *C. coli* are the most commonly reported causes of Campylobacteriosis in humans [48]. Table 7 presents the results of studies conducted in Iran regarding the prevalence of *Campylobacter* in different food items. As illustrated in Figure 8, the most prevalent occurrence of *Campylobacter* contamination in Iran was observed in poultry meat (46.21%), followed by red meat (40%) and eggs (28.06%). The contamination of dairy products and raw milk was observed in 2.36% and 2.5% of samples, respectively (Figure 8). A study conducted in

Table 7.

Summary of the studies reporting the prevalence of *Campylobacter* spp. in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	Campylobacter spp.	References	Area
1	2023	chicken meat	100	81	81	Campylobacter spp.	[286]	Hamedan
2	2023	chicken meat	255	64	25.09	Campylobacter spp	[287]	Shahrekord
3	2023	Mushrooms	740	74	10	Campylobacter spp	[288]	-
4	2022	poultry meat	380	24	6.25	Campylobacter spp	[289]	Shahrekord
5	2022	poultry meat	100	35	35	Campylobacter spp	[290]	Tehran
6	2022	raw meat	200	27	13.5	Campylobacter jejuni	[291]	-
7	2022	cattle raw milk	100	7	7	Campylobacter jejuni	[292]	Mazandaran
8	2021	Poultry Carcasses	370	203	54.8	Campylobacter spp	[293]	south of Iran
9	2021	Camels meat	40	5	12.5	Campylobacter spp.	[294]	Chaharmahal and Bakhtiari
10	2019	poultry meat	328	217	66.7	Campylobacter spp.	[295]	Jahrom
		Industrial chicken		1	0.6			
		meat		0	0	Campylobacter spp.		
		Traditional chicken		8	16	C. jejuni	-	
		meat		3	37.5	Campylobacter spp	-	
		Fresh packed		2	-	C. jejuni	_	
11	2019	chicken meat		0	0	Campylobacter spp	[295]	Ahvaz
				7	14			
		Beef meat		7	100	Campylobacter spp.		
		Mutton meat		12	24	C. jejuni	-	
				12	100	С. јејит	_	
		Water buffalo meat	50	4	8	Campylobacter spp.		
				4	100	C. jejuni		
12	2019	Packed chicken meat		26	28.9	Campylobacter spp.	[296]	Shiraz

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Tabl	e 7 cont.							
		Red meat	90	21	23	Campylobacter spp.		
				13	61.9			
13	2016	Chicken-meat	120	33	27.5	C. jejuni	[297]	Zanjan
				22	66.6	Campylobacter spp.		
		Eggshells	120	38	31.6	C. jejuni		
				20	52.6	Campylobacter spp.		
14	2015	Chicken wing	96	37	38.5	Campylobacter spp.	[298]	Urmia
15	2015	Meat	360	227	63.1	Campylobacter spp.	[299]	Mashhad
				200	88.1	C. jejuni		
16	2015	Raw ovine milk	38	0	0	C. jejuni	[300]	Zanjan
		Raw bovine milk	22	0	0	C. jejuni	[000]	
17	2014	Chicken	250	110	44	Campylobacter spp.	[300]	Tehran
			200	87	79		[000]	
		Pasteurized milk	30	0	0			
		Camel milk	37	0	0	Campylobacter spp.		
		Commercial dairy	290	0	0			
		Raw cow milk	80	5	6.2	Campylobacter spp.		Isfahan &
18	2013	Raw sheep milk	60	1	1.6	Campylobacter spp.	[301]	Chaharmahal v
		Raw goat milk	60	2	3.3	Campylobacter spp.		Dalahtarani
		Traditional cheese	60	3	5	Campylobacter spp.		Bakhtyari
		Traditional ice- cream	35	1	2.8	Campylobacter spp.		
		Traditional butter	25	1	4	Campylobacter spp.		
		Packed chicken meat	96	22	22.9	Campylobacter spp.	[]	
19	2012	Unpacked chicken meat	104	31	28.8	Campylobacter spp.	[121]	Mazandaran
20	2011	Eggs	100	0	0	C. jejuni	[124]	Shahrekord
21	2011	Raw bovine milk	120	3	2.5	Campylobacter spp.	[302]	Isfahan
		Chielson	200	94	47	Campylobacter spp.		
		Chicken	200	91	96.8	C. jejuni		
			40	49	49	Campylobacter spp.		
		Turkey	49	41	83.7	C. jejuni	•	
~~		0 1	22	37	43	Campylobacter spp.	[202]	
22	2011	Quail	33	33	89.2	C. jejuni	[303]	Shahrekord
				6	35.3	Campylobacter spp.		
		Partridge	6	6	100	C. jejuni		
				1	4.8	Campylobacter spp.		
		Ostrich	21	1	100	C. jejuni		
	1			37	61.7	Campylobacter spp.		
		Chicken meat	60	35	94.6	C. jejuni		
				18	83.3	Campylobacter spp.		
		Turkey meat	50	15	36	C. jejuni		
23	2010			3	6	Campylobacter spp.	[304]	Ahvaz
		Sheep meat	50	1	33.3	C. jejuni		
				17	4.4	Campylobacter spp.		
		Goat meat	45	17	100	C. jejuni		
				1/	100	C. jejuni		

Table 7 cont.

	~ 1		1	0.9	Campylobacter spp.		
	Raw camel meat	107	0	0	C. jejuni	•	
		100	5	2.4	Campylobacter spp.	•	
24 2010	Beef meat	190	3	60	C. jejuni	[305]	Isfahan & Yazd
24 2010	Lamb meat	225	27	12	Campylobacter spp.	[305]	
		225	23	92	C. jejuni		
	Costmost	190	17	9.4	Campylobacter spp.		
	Goat meat	180	16	94.1	C. jejuni	-	
	Raw chicken meat	280	157	56.1	Campylobacter spp.		
	Raw chicken meat	280	140	89.2	C. jejuni		
	Queilment	248	68	27.4	Campylobacter spp.		
25 2008	Quail meat	248	53	77.9	C. jejuni	[206]	Isfahan
2008		212	145	68.4	Campylobacter spp.	[306]	Islanan
	Turkey meat	212	92	63.4	C. jejuni	-	
	Ostrich meat	60	7	11.7	Campylobacter spp.	-	
	Ostricii ineat	00	3	42.9	C. jejuni	-	

Table 7 cont.

the United States in 2020 reported that while various broiler products carry the risk of *Campylobacter spp*. contamination, the highest prevalence of contamination was observed in chicken carcasses [49]. Similarly, in the European Union, *C. jejuni* has been identified as the most prevalent species (51%) in broiler meat, followed by *C. coli* (35.5%) [47]. Consequently, poultry meat represents the greatest risk of *Campylobacter* transmission to humans worldwide. The consistent reporting of the highest prevalence of *Campylobacter* contamination in poultry meat in multiple studies, including those conducted in Iran, the United States, and the European Union, underscores the importance of addressing this issue [47, 49]. This finding highlights the necessity of implementing rigorous food safety measures and regulations in the poultry industry to prevent the transmission of *Campylobacter* to consumers.

8) Helicobacter pylori prevalence in food

Helicobacter pylori is associated with several digestive diseases, including peptic ulcer, mucosa-associated lymphoid tissue lymphoma, gastritis, and an increased risk of gastric cancer [50]. It is estimated that approximately 50% of the global population is infected with *H. pylori* [51]. The prevalence of *H. pylori*



Figure 8. Prevalence of *Campylobacter* in different foods in Iran.

infection is observed to be higher in developing countries, with rates ranging from 70% to 90%, compared to developed countries, where rates are reported to be 25%-50%. Iran is considered a high-risk region for *H*. *pylori* infection due to the high prevalence (60%-90%) among its population [52]. H. pylori can be found in a variety of animal-derived foods, vegetables, and water sources, which contribute to its transmission [50]. Table 8 presents the findings of studies conducted in Iran regarding the prevalence of H. pylori

Table 8.

Summary of the studies reporting the prevalence of *H. pylori* in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	References	Area
1	2023	Raw Poultry Meat	320	20	6.25	[307]	Shahrekord
2	2020	Red meat	600	52	8.6	[308]	Tehran
3	2018	Traditional dairy products	800	31	3.8	[309]	Isfahan
4	2017	Red meat	220	11	5	[310]	Isfahan
		Meat	150	11	7.3		
5	2017	Milk	150	24	16	[311]	Alborz
		Vegetable	40	5	12.5		
		Cream-candy	50	9	18		
		Traditional bread	50	3	6		
		Sausage	50	0	0		
		Salami	50	0	0		
		Hamburger	50	1	2		
6	2016	Soup	50	11	22	[312]	Isfahan
		Restaurant salad	50	15	30		
		Falafel	50	3	6		
		Olivier salad	50	18	36		
		Chicken nugget	50	0	0		
		Fruit salad	50	14	28		
_		Milk	420	92	21.9	[040]	
7	2016	Meat	400	105	26.2	[313]	Shahrekord
8	2016	Meat products	150	11	7.3	[314]	Isfahan
			120	5	4.2		Isfahan
0	2016	Dainlying water (total)	110	2	1.8	[215]	Shiraz
9	2016	Drinking water (total)	100	3	3	[315]	Yazd Shahrekord
		Ready to eat fish	70	2	2.8		
		Ham	60	9	15		
		Chicken sandwich	60	5	8.3		
10	2016	Vegetable sandwich	40	2		[017]	Cl. :
10	2016	Meat sandwich	40	18	5	[316]	Shiraz
		Minced meat	50	10	45		
		Minced meat	50	16	20		
					32		
11	2015	Raw milk	210	28	13.3	[317]	Tehran
		Bovine milk	120	20	16.6		
10	2017	Traditional cheese	80	10	[318]		
12	2015	Traditional cream	40	3	7.5	[318]	
		Total	240	33	[319]		

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13	2014	Vegetable and salad	460	44	9.5	[319]	Shahrekord
14	2014	Vegetable and salad	430	59	13.7	[320]	Isfahan
15	2013	Water	200	14	7.2	[51]	Isfahan
16	2012	Milk	447	56	12.5	[321]	Isfahan, Fars, Chaharmahal & Bakhtiari, Khuz- estan

Table 8 cont.

in various food items. As illustrated in Figure 9, the highest prevalence of H. pylori in food samples in Iran was observed in RTE foods (25.5%) and vegetables (22.14%), followed by raw milk (16.06%), red meat (15.82%), dairy products (7.93%), meat products (6.26%), and water (3.8%) (Figure 9). In other countries, studies have also identified the presence of H. pylori in a variety of food sources. In Japan, the ureA gene of H. pylori was found in 72.2% of raw milk samples and 55% of pasteurized milk samples [53]. In Italy, the glmM gene of H. pylori was identified in 34.7% of raw milk samples [54]. In the United States, H. pylori was detected in 44% of RTE raw tuna meat and 36% of raw chickens using a multiplex PCR assay [55]. These findings underscore the potential presence of H. pylori in various food sources and the significance of food as a potential route of transmission.

9) Clostridium prevalence in food

Clostridium botulinum

Clostridium botulinum is a gram-positive, anaerobic bacterium that is capable of producing spores. It is known to cause botulism, a severe illness characterized by the production of a potent neurotoxin. Table 9 presents the findings of research conducted in Iran on the prevalence of *C. botulinum* in various food items. As illustrated in Figure 10, the most prevalent contamination of *C. botulinum* in Iran was observed in seafood (12.56%), followed by red meat (12.23%), dairy products (9.02%), and honey (2%) (Figure 10). Honey is recognized as a reservoir for *C. botulinum* spores, particularly types B and A, and has been implicated in cases of neonatal botulism [30]. Studies conducted in various countries, including Turkey, Brazil, Denmark, Sweden, and Norway, have demonstrated the presence of *C. botulinum* spores in honey samples, with prevalence rates ranging from 2% to 26% [30]. In Iran, the prevalence of *C. botulinum* contamination in honey samples was reported to be 2% (Figure 10), indicating a relatively lower level of contamination compared to some other regions.

While *C. botulinum* spores may be present in certain foods, the risk of botulism is contingent upon the conditions that facilitate the germination of spores and toxin production, such as inadequate food processing, storage, or handling. Proper food safety practices, including adequate cooking, storage at appropriate temperatures, and hygienic handling, can help prevent the growth and tox-in production of *C. botulinum* in food.

Clostridium perfringens

C. perfringens is a significant contributor to foodborne gastrointestinal illnesses in both humans and animals. The spores of *C. perfringens* exhibit remarkable resilience to external influences. In Europe in 2020, there



Figure 9.	
Prevalence of <i>H. pylori</i> in different foods in Iran.	



Figure 10. Prevalence of *Clostridium* in different foods in Iran.

were 682 reported cases of food poisoning caused by *C. perfringens* toxins, with a hospitalization rate of 1.5%. Conversely, there were fewer cases (n = 34) of food poisoning due to *C. botulinum* toxins, yet the hospitalization rate for botulism cases was 100%. It is noteworthy that no fatalities were reported in these cases. Early diagnosis, hospitalization, and treatment are essential for reducing the severity of botulism [30]. Table 9 presents the findings of studies conducted in Iran regarding the prevalence of *C. perfringens* in various food items. *C. perfringens* type A is the most prevalent cause of food poisoning associated with this bacterium. The available data indicate that

C. perfringens was most commonly isolated from red meat in Iran. It is of paramount importance to ensure that meat is cooked and handled properly to minimize the risk of contamination with *C. perfringens* and subsequent foodborne illnesses. In Europe in 2019, two outbreaks were associated with pig meat and products, one caused by toxins produced by *C. perfringens* and the other by *C. botulinum*. Conversely, vegetables, juices, and other related products were linked to a greater number of outbreaks, with two outbreaks reported for each category during the same period [30]. Nevertheless, only one study has been conducted in Iran regarding the presence of *C. perfringens* in vege-

Table 9.

Summary of the studies reporting the prevalence of <i>Clostridium spp.</i> in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	Clostridium spp.	References	Area
1	2023	Meat native birds	300	35	11.6	C. perfringens	[322]	Shahrekord
2	2023	Meat Nuggets	600	7	1.17	C. perfringens	[323]	Isfahan
3	2023	Ground Beef	133 94	24 3	18.04 3.22	C. perfringens	[324]	Qazvin
4	2022	Raw Meat	240	7	2.91	C. perfringens	[325]	Mazandaran
5	2022	Raw Beef Meats	133	18	13.53	C. perfringens	[326]	Qazvin
6	2022	raw and ready-to- eat green leafy veg- etables	366	66	18	C. perfringens	[139]	Tehran
7	2022	Olivier Salad	26	0	0	C. perfringens	[327]	Mashhad
8	2021	Cattle and sheep carcasses	200	61	30.5	C. perfringens	[328]	Shiraz

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9	2021	Broiler chickens	122	95	77.8	C. perfringens	[329]	Kerman
10	2019	Broiler chickens	400	169	42.2	C. perfringens	[330]	Chaharmahal 8 Bakhtiari
11	2019	Honey	130	0	0	C. perfringens	[331]	
		Traditional curds	50	12	25			
10		Commercial curds	50	5	10		[]	
12	2017	Beef meat	20	1	6	C. perfringens	[332]	Shahrekord
		Lamb meat	23	3	13			
13	2015	Broiler meat	200	31	15.5	C. perfringens	[333]	Mashhad
14	2015	Minced meat	200	25	12.5	C. perfringens	[334]	Mashhad
15	2013	Honey	100	2	2	C. perfringens	[335]	Shiraz
		Fish	80	4	5	C. perfringens	[336]	Shiraz
16	2012	Honey	50	2	4			
16	2013	Kashk	80	2	2.5			
		Dough	80	1	1.2			
		Dairy products	57	12	21		[337]	Gilan, Tehran,
17	2013	Fish	68	18	26.4	C. perfringens		Golestan,
		Meat	14	1	7.1			Hamedan
		Cheese	57	2	3.5		[338]	
18	2010	Kashk	11	0	0	C. perfringens		Gilan
		Salted fish	63	4	6.3			

Table 9 cont.

tables and juices, and other related products. Further research and surveillance are necessary to gain a more comprehensive understanding of the prevalence and sources of *C. perfringens* in various food items in Iran.

10) Brucella prevalence in food

Brucella spp. are the causative agents of brucellosis [56], an infectious disease of humans that presents with chronic and recurring febrile symptoms that can be life-threatening [57]. The primary etiological agent of the disease is B. melitensis, although other species, including B. abortus, B. canis, and B. suis, can also result in human brucellosis [58]. The infection can be transmitted to humans from various animals, including buffalo, cattle, yak, elk, camel, domestic pig, and rodents [58]. Globally, approximately 500,000 cases of human brucellosis are reported annually, with animals and animal-derived foods serving as the primary sources of infection [57]. A global systematic review conducted in 2020 revealed that the Southeast Asia region exhibited the highest prevalence of Brucella spp. at 25.55% [57]. The consumption of unpasteurized dairy products plays a significant role in the transmission of Brucella spp. to humans [57]. Table 10 presents the results of studies conducted in Iran on the prevalence of Brucella spp. in food. As illustrated in Figure 11, the primary sources of reported contamination with *Brucella spp.* are dairy products (34.28%) and raw milk (16.64%). Dairy products, particularly unpasteurized or inadequately pasteurized ones, can serve as reservoirs for *Brucella* contamination [57]. This can occur due to infected dairy animals shedding the bacteria in their milk. Raw milk, in particular, has been identified as a common source of *Brucella* infection in various parts of the world, including Iran. Improper handling and processing of raw milk can contribute to the transmission of *Brucella spp.* to humans [59].

In Iran, where dairy products hold cultural and dietary significance, ensuring the safety of these products from *Brucella* contamination is crucial for public health [60]. Implementing stringent control measures in dairy production, processing, and distribution can help mitigate the risk of *Brucella* transmission through dairy products and raw milk [57, 59, 60].

11) Vibrio prevalence in food

Vibrio spp. are halophilic marine bacteria. Some species, including *V. cholerae, V. parahaemolyticus,* and *V. vulnificus,* have the potential to cause gastroenteritis or septicemia in humans. The primary mode of transmission for this foodborne illness is the ingestion of raw, undercooked, or mishandled seafood contaminated by bacteria [61]. Table 11 presents the re-



Figure 11. Prevalence of *Brucella* in different foods in Iran.

Table 10.Summary of the studies reporting the prevalence of *Brucella spp.* in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	Brucella spp.	References	Area
1	2022	Unpasteurized Milk and Dairy	291	12	4.1	Brucella spp.	[339]	Hamadan
2	2021	Bovine Milk	240	16	6.66	Brucella spp.	[340]	Kurdistan
		Dairy products	227	9	4			
3	2020	Non boiling milk	43	1	2.3	Brucella spp.	[341]	Hamadan
		Fresh cheese Cream	21	2	[341]			
			96	3	3			Isfahan,
4	2019	19 Raw camel milk	51	2	[342]	Brucella spp.	[342]	
			45	1				Semnan
	2018	Dairy Products	208	60	28.8	Brucella spp.		Tehran
		Goat raw milk	33	15	45.5		[50]	
		Non-pasteurized cheese	23	9	39.1			
5		Sheep raw milk	33	9	27.3		[58]	
		Cow raw milk	57	15	26.3			
		Pasteurized cheese	28	7	25			
		Pasteurized milk	34	5	14.7			
				11	78.6	Brucella spp.	[343]	Tehran
6	2017	Dairy Products	14	8	72.7	B. melitensis [343] B. abortus		
				3	27.3			
7	2017	Sheep raw milk Goat raw milk	530	41	8.1	Brucella spp.	[344]	Kerman
		Raw milk	700	9	1.28			
8	2017	Sheep's raw milk	300	3	1	Brucella spp.	[345]	Kerman
		Goats raw milk	400	6	1.5			
9		Unpasteurized milk	132	4	3	Brucella spp.	[346]	Isfahan
		Dairy products	65	1	1.5			
10	2016	Cow's raw milk	48	4	8.3	Brucella spp.	[347]	Kerman

Tabi	e 10 cont.							
	2016	Milk	225	20	8.9		[348]	Shahrekord & Isfahan
11		Sheep milk	125	12	9.6	Brucella spp.		
		Goat milk	100	18	18			∝ Islanan
12	2016	Raw goat milk	470	51	10.8	Brucella spp.	[344]	Southeast region of Iran
12	2016	Raw sheep milk	330	18	5.4			
13	2015	Raw milk	60	32	53.3	Brucella spp.	[200]	Zanjan
	2014	Raw cow milk	57	19	33	Brucella spp.	[349]	Tehran
		Pasteurized cow milk	34	10	29			
14		Pasteurized cheese	28	8	28			
		Traditional cheese	23	14	60			
		Raw goat milk	33	21	63			
		Raw sheep milk	33	19	57			
15	2012	Cattle milk	1117	18	1.6	Duvalla	[350]	Urmia
15	2013	Sheep milk	598	99	16.5	Brucella spp.		

Table 10 cont.

sults of studies conducted in Iran on the prevalence of Vibrio spp. in different types of food. *Vibrio spp*. were predominantly detected in seafood, including lobster, fish products, crayfish, fish, and shrimp, as well as drinking water. As illustrated in Figure 12, the prevalence of Vibrio spp. was highest in seafood, with fish exhibiting the greatest incidence (49.33%), followed by lobster (21.53%), crayfish (8.63%), shrimp (8.12%), fish products (7.8%), and drinking water (1.3%) (Figure 12). The findings from Iran are in alignment with those from other countries. For instance, a comprehensive systematic review conducted in 2016 revealed that V. parahaemolyticus contamination was observed in 63.4% of oysters, 52.9% of clams, 51% of fish, and 48.3% of shrimps [62]. A similar study in China in

2020 reported that 15.34% of shrimp samples, 14.17% of fish samples, and 3.67% of RTE food were contaminated with *V. parahaemolyticus* [63]. However, there are no reports available from Iran regarding the prevalence of *V. parahaemolyticus* in RTE foods.

12) Shigella prevalence in food

The Shigella genus encompasses four known species: S. dysenteriae, S. boydii, S. flexneri, and S. sonnei, which have also been classified as subgroups A to D, respectively [64]. While S. flexneri has traditionally been reported as the main cause of shigellosis in developing countries, recent studies have shown that S. sonnei has become the predominant species of Shigella in Iran [64]. According to the WHO, Shigella



Figure 12. Prevalence of *Vibrio* in different foods in Iran.

Table 11.

Summary of the studies reporting the prevalence of Vibrio spp. in Iran.

			-					
	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	Type of Vibrio spp.	References	Area
1	2021	Fish	64	61	95	Vibrio species	[351]	Isfahan
2	2020	Frozen fish	200	0	0	V. parahaemolyticus	[352]	Mazandaran
3		Shrimp	70	12	17.1	V. parahaemolyticus	[353]	Zanjan
		Cold Smoked Salt-	• • • •				[0=4]	
4	2018	ed Fishes	200	46	23	Vibrio spp.	[354]	Mazandaran
5	2016	Fish	58	18	31	- V. parahaemolyticus	[255]	Persian Gulf
5	2016	Shrimps	55	7	12.7	• v. paranaemolylicus	[355]	Persian Guii
		Fresh shrimps	30	2	6.6	-		Genaveh
6	2015	Salted shrimps	30	2	6.6	V. parahaemolyticus	[356]	seaport
-	2014		26		10.4	17.1	[]	South coast of
7	2014	Shrimps	36	7	19.4	Vibrio spp.	[357]	Iran
		Fish	100	22	22		[358]	Bushehr, Persian
8	2014	Lobster	60	13	21.6	V. parahaemolyticus		
		Crab caught	40	7	17.5	_		Gulf
		Crayfish		11	11.3	V. vulnificus	_	Aras
9	2014		97	7	7.2	V. harveyi	- [357]	
	2011		27	2	2	V. alginolyticus	_	
				1	1	V. mimicus		
		Tap-water	144	3	2	_		
10	2013	Bottled mineral water	304	3	0.6	V. cholerae	[360]	Isfahan
		Fresh shrimp	70	5	7.1	-		
11	2012	Salted fishes	70	2	2.9	- V. parahaemolyticus	[122]	
	2012	Fish nugget	10	0	0	•	[122]	
		Shrimp burger	10	0	0			
		Lobsters	100	40	40	Vibrio spp.	_	
12	2012		100	3	3	V. parahaemolyticus	[361]	Persian Gulf
		Crab	32	4	12.5	Vibrio spp.		
			32	1	3.1	V. parahaemolyticus		
13	2010	Fresh shrimp	300	29	9.6	V. parahaemolyticus	[361]	Bohsher. Hor- mozgan,Khoozc stan
14	2004	Fresh shrimp	770	16	2.1	Vibrio spp.	[361]	Bohsher. Hor- mozgan, Khooz- estan

spp. cause approximately 165 million cases of *bacillary* dysentery and 1 million deaths worldwide each year [64]. In general, *Shigella spp.* are among the most prevalent causes of acute diarrhea in Iran, with a particularly high incidence among children and young adults. A diverse array of foods, encompassing meat, dairy products, and vegetables, have been identified as potential sources of shigellosis outbreaks worldwide [64]. Table 12 presents the results of studies conducted in Iran on the prevalence of *Shigella spp.* in different types of food. As illustrated in Figure 13, contamina-

tion with *Shigella spp*. is most commonly reported in RTE foods (1.72%) and vegetables (1.05%), followed by red meat (0.4%). In contrast to the data from Iran, a high prevalence of *Shigella spp*. contamination has been reported in vegetables (25.25%) in India [65], and in beef, chicken, and dairy products in Egypt [66]. According to our review, poultry meat should be considered a high-risk food with the potential to spread foodborne zoonoses in Iran. In general, poultry meat is more susceptible to contamination during processing and handling due to its higher water con-

Table 12.

Summary of the studies reporting the prevalence of *Shigella spp*. in Iran.

	Year	Sample type	Sample size	Positive sam- ples (N)	Prevalence (%)	Type of Shigella spp.	References	Area
1	2022	raw milk, ground meat, and raw vegetable	580	13	2.24	Shigella sonnei	[364]	Tehran and Qazvin
2	2021	Vegetable salad, ground meat, and raw cow's milk	405	18	4.44	Shigella spp.	[365]	Qazvin
3	2021	meat, vegetable sal- ad and raw milk	165	8	4.84	Shigella spp.	[366]	Qazvin
				2	0.8	S. sonnei	_	
	2019	Ready to eat food	250	0	0	S. flexneri	_	
			230	0	0	S. dysenteriae	_	
				0	0	S. boydii	 [64]	Isfahan, Fars, Hormozgan, Kohkiloyeh va Boyer Ahmad
		Fresh meat	150	1	0.7	S. sonnei		
				2	1.3	S. flexneri		
			150	0	0	S. dysenteriae		
1				0	0	S. boydii		
t		Frozen meat	150	0	0	Shigella spp.		
		Cow milk	100	0	0	Shigella spp.		
		Domestic cheese	100	0	0	Shigella spp.		
		Vegetables		8	1.2	S. sonnei	_	
			650	6	0.9	S. flexneri	-	
			030	0	0	S. dysenteriae		
				0	0	S. boydii		
		1400		19	1.4	Shigella spp.		
5	2018	Ready-to-Eat Salad	90	7	7.8	Shigella spp.	[367]	Tehran
5	2018	Food (vegetables, chicken, minced meat, fish)	100	6	6	Shigella spp.	[368]	Shiraz
7	2014	Camel milk	18	0	0	Shigella spp.	[369]	Golestan



raw state, thereby increasing the probability of contamination if the requisite hygiene standards are not observed during slaughter, processing, and storage. In contrast, red meat and seafood have lower contamination rates compared to poultry meat, likely due to differences in processing and handling practices [68]. These findings underscore the necessity of developing strategies to reduce the contamination levels of poultry meat to effectively control and prevent foodborne illnesses in Iran.

tent and pH levels, which provide an optimal environment for the proliferation of bacteria [67]. Moreover, poultry meat is frequently sold and consumed in its

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The risk of food contamination, particularly in meat products, is significant. However, to effectively underscore the importance of foodborne diseases, it is imperative to document the consequences of infection

Prevalence of Shigella in different foods in Iran.

with these pathogens and generalize this information to the population in Iran. Currently, foodborne diseases in Iran are not generally reported, leading to a likely gross underestimation of their burden. This underestimation is attributable to the fact that many foodborne illnesses do not exhibit sufficient severity, duration, or specific diagnostic criteria for accurate identification and intervention. Similar circumstances exist in developed countries, such as the United States. For instance, the CDC estimates that foodborne pathogens cause approximately 48 million illnesses, 128,000 hospitalizations, and 3,000 deaths annually in the US [70].

Therefore, it is crucial to emphasize the necessity of establishing robust monitoring systems in Iran. Such a surveillance network would require the collaboration of multidisciplinary teams comprising medical doctors, veterinarians, microbiologists, public health specialists, and other relevant experts, in alignment with the One Health concept. By adopting a methodology similar to that employed by the CDC's Foodborne Diseases Active Surveillance Network (Food-Net), which monitors the incidence of nine foodborne pathogens in ten US cities, representing approximately 15% of the American population [71], Iran can enhance the awareness of foodborne disease events and trends. These practices enable the implementation of effective intervention and prevention strategies.

Authors' Contributions

MH suggested the topic and supervised the conduction of the systematic review. SA wrote the first draft of the manuscript. FA and SA performed the literature review. GS was the major contributor in writing the manuscript. AA gave advice for conducting and writing the manuscript. All authors read and approved the final manuscript.

Competing Interests

The authors declare that there is no conflict of interest.

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It is not applicable

References

- Touranlou FA, Noori SMA, Salari A, Afshari A, Hashemi M. Application of kefir for reduction of contaminants in the food industry: A systematic review. International Dairy Journal. 2023:105748. Doi:10.1016/j.idairyj.2023.105748.
- Law JW-F, Ab Mutalib N-S, Chan K-G, Lee L-H. Rapid methods for the detection of foodborne bacterial pathogens: principles, applications, advantages and limitations. Frontiers in microbiology. 2015;5:122692. Doi:10.3389/fmicb.2014.00770.

- 3. Imre K, Herman V, Morar A. Scientific achievements in the study of the occurrence and antimicrobial susceptibility profile of major foodborne pathogenic bacteria in foods and food processing environments in Romania: Review of the last decade. BioMed Research International. 2020;2020. Doi:10.1108/NFS-01-2023-0023.
- Shakeri G, Jamshidi A, Ghazvini K. Isolation of broad-hostrange bacteriophages against food-and patient-derived Shiga toxin-producing Escherichia coli. Iranian Journal of Veterinary Science and Technology. 2020;12(1):11-20. Doi:10.22067/veterinary.v12i1.85298.
- Moghimani M, Noori SMA, Afshari A, Hashemi M. D tryptophan, an eco friendly natural, safe, and healthy compound with antimicrobial activity against food borne pathogens: A systematic review. Food Science & Nutrition. 2024;12(5):3068-79. Doi:10.1002/fsn3.3987.
- Gourama H. Foodborne pathogens. Food safety engineering: Springer; 2020. p. 25-49. Doi:10.1007/978-3-030-42660-6_2.
- Tîrziu E, Bărbălan G, Morar A, Herman V, Cristina RT, Imre K. Occurrence and antimicrobial susceptibility profile of Salmonella spp. in raw and ready-to-eat foods and Campylobacter spp. in retail raw chicken meat in Transylvania, Romania. Foodborne Pathogens and Disease. 2020;17(8):479-84. Doi:10.1089/fpd.2019.2738.
- Hashemi M, Erfani A, Dadgar Moghadam M, Dousti Nouri M, Asadi Touranlou F, Azadi N, et al. Assessment of knowledge, Attitudes, and Practices of Food Service Staff Regarding Food Safety at Mashhad University Hospitals. Journal of Nutrition, Fasting and Health. 2024. Doi: 10.22038/ JNFH.2024.78388.1505.
- WHO. WHO estimates of the global burden of foodborne diseases: foodborne disease burden epidemiology reference group 2007-2015: World Health Organization; 2015.
- Tack DM, Marder EP, Griffin PM, Cieslak PR, Dunn J, Hurd S, et al. Preliminary incidence and trends of infections with pathogens transmitted commonly through food— Foodborne Diseases Active Surveillance Network, 10 US Sites, 2015–2018. Morbidity and Mortality Weekly Report. 2019;68(16):369. Doi:10.15585/mmwr.mm6816a2.
- 11. Pires SM, Evers EG, Van Pelt W, Ayers T, Scallan E, Angulo FJ, et al. Attributing the human disease burden of foodborne infections to specific sources. Foodborne pathogens and disease. 2009;6(4):417-24. Doi:10.1089/fpd.2008.0208.
- Mukhopadhyay S, Ramaswamy R. Application of emerging technologies to control Salmonella in foods: A review. Food Research International. 2012;45(2):666-77. Doi:10.1016/j. foodres.2011.05.016.
- 13. WHO. Estimating the burden of foodborne diseases: a practical handbook for countries: a guide for planning, implementing and reporting country-level burden of foodborne disease.

2021.

- Yang J, Zhang Z, Zhou X, Cui Y, Shi C, Shi X. Prevalence and characterization of antimicrobial resistance in Salmonella enterica isolates from retail foods in Shanghai, China. Foodborne Pathogens and Disease. 2020;17(1):35-43. Doi:10.1089/ fpd.2019.2671.
- Nair DV, Kollanoor Johny A. Salmonella in poultry meat production. Food safety in poultry meat production. 2019:1-24. Doi:10.1007/978-3-030-05011-5_1.
- Shafini A, Son R, Mahyudin N, Rukayadi Y, Zainazor TT. Prevalence of Salmonella spp. in chicken and beef from retail outlets in Malaysia. International Food Research Journal. 2017;24(1):437.
- Carrasco E, Morales-Rueda A, García-Gimeno RM. Cross-contamination and recontamination by Salmonella in foods: A review. Food Research International. 2012;45(2):545-56. Doi:10.1016/j.foodres.2011.11.004.
- Ehuwa O, Jaiswal AK, Jaiswal S. Salmonella, food safety and food handling practices. Foods. 2021;10(5):907. Doi:10.3390/ foods10050907.
- Ehuwa O, Jaiswal A, Jaiswal S. Salmonella, Food Safety and Food Handling Practices. Foods 2021, 10, 907. Foodborne Pathogens and Food Safety. 2021:175. Doi:10.3390/ foods10050907.
- Kowalska B. Fresh vegetables and fruit as a source of Salmonella bacteria. Annals of Agricultural and Environmental Medicine. 2023;30(1). Doi: 10.26444/aaem/156765.
- Kadariya J, Smith TC, Thapaliya D. Staphylococcus aureus and staphylococcal food-borne disease: an ongoing challenge in public health. BioMed research international. 2014;2014. Doi:10.1155/2014/827965.
- 22. Naas HT, Edarhoby RA, Garbaj AM, Azwai SM, Abolghait SK, Gammoudi FT, et al. Occurrence, characterization, and antibiogram of Staphylococcus aureus in meat, meat products, and some seafood from Libyan retail markets. Veterinary World. 2019;12(6):925. Doi: 10.14202/vetworld.2019.925-931.
- Dallal MMS, Foroushani AR, Sharifi-Yazdi S, Sharifi-Yazdi MK, Arfatahery N. Prevalence of Staphylococcus aureus in Shrimps in Tehran during 2013. Journal of Medical Bacteriology. 2015;4(5-6):42-6.
- 24. Pourbabaee M, Hadadi MR, Hooshyar H, Pourbabaee P, Nazari-Alam A. Prevalence of Staphylococcus aureus in Raw Hamburgers from Kashan in 2017. International Archives of Health Sciences. 2020;7(1):47-50. Doi: 10.4103/iahs. iahs_39_19.
- 25. Hamidiyan N, Salehi-Abargouei A, Rezaei Z, Dehghani-Taf-

ti R, Akrami-Mohajeri F. The prevalence of Listeria spp. food contamination in Iran: A systematic review and meta-analysis. Food Research International. 2018;107:437-50. Doi:10.1016/j.foodres.2018.02.038.

- Authority EFS, Prevention ECfD, Control. The European Union one health 2020 zoonoses report. EFSA Journal. 2021;19(12):e06971. Doi:10.2903/j.efsa.2021.6971.
- 27. Khademi F, Sahebkar A. A systematic review and meta-analysis on the prevalence of antibiotic-resistant Listeria species in food, animal and human specimens in Iran. Journal of food science and technology. 2019:1-17. Doi:10.1007/s13197-019-04040-w.
- Dalton HR, Dreier J, Rink G, Hecker A, Janetzko K, Juhl D, et al. Coxiella burnetii-pathogenic agent of Q (query) fever. Transfusion medicine and hemotherapy. 2014;41(1):60-72. Doi:10.1159/000357107.
- 29. Abdali F, Hosseinzadeh S, Berizi E, Shams S. Prevalence of Coxiella burnetii in unpasteurized dairy products using nested PCR assay. Iranian Journal of Microbiology. 2018;10(4):220.
- EFSA. The European Union one health 2018 zoonoses report. EFSa Journal. 2019;17(12):e05926. Doi:10.2903/j. efsa.2019.5926.
- Rafiee S, Khoshnevisan B, Mohammadi I, Aghbashlo M, Clark S. Sustainability evaluation of pasteurized milk production with a life cycle assessment approach: an Iranian case study. Science of the total environment. 2016;562:614-27. Doi:10.1016/j.scitotenv.2016.04.070.
- 32. Moradnejad P, Esmaeili S, Maleki M, Sadeghpour A, Kamali M, Rohani M, et al. Q fever endocarditis in Iran. Scientific reports. 2019;9(1):15276. Doi:10.1038/s41598-019-51600-3.
- 33. Van den Brom R, Van Engelen E, Roest H, Van der Hoek W, Vellema P. Coxiella burnetii infections in sheep or goats: an opinionated review. Veterinary microbiology. 2015;181(1-2):119-29. Doi:10.1016/j.vetmic.2015.07.011.
- 34. Guidi F, Petruzzelli A, Ciarrocchi F, Duranti A, Valiani A, Amagliani G, et al. Prevalence of Coxiella burnetii in cows' and ewes' bulk tank milk samples from selected dairy farms of Central Italy. Italian Journal of Animal Science. 2017;16(4):673-6. Doi:10.1080/1828051X.2017.1321474.
- 35. Rozental T, Faria LSD, Forneas D, Guterres A, Ribeiro JB, Araújo FR, et al. First molecular detection of Coxiella burnetii in Brazilian artisanal cheese: a neglected food safety hazard in ready-to-eat raw-milk product. Brazilian Journal of Infectious Diseases. 2020;24(3):208-12. Doi:10.1016/j. bjid.2020.05.003.
- Kim SG, Kim EH, Lafferty CJ, Dubovi E. Coxiella burnetii in bulk tank milk samples, United States. Emerging infectious diseases. 2005;11(4):619. Doi: 10.3201/eid1104.041036.

- Berthold-Pluta A, Pluta A, Garbowska M, Stefańska I. Prevalence and toxicity characterization of Bacillus cereus in food products from Poland. Foods. 2019;8(7):269. Doi:10.3390/ foods8070269.
- Tewari A, Abdullah S. Bacillus cereus food poisoning: international and Indian perspective. Journal of food science and technology. 2015;52(5):2500-11. Doi:10.1007/s13197-014-1344-4.
- Webb MD, Barker GC, Goodburn KE, Peck MW. Risk presented to minimally processed chilled foods by psychrotrophic Bacillus cereus. Trends in food science & technology. 2019;93:94-105. Doi:10.1016/j.tifs.2019.08.024.
- 40. Eglezos S, Huang B, Dykes GA, Fegan N. The prevalence and concentration of Bacillus cereus in retail food products in Brisbane, Australia. Foodborne pathogens and disease. 2010;7(7):867-70. Doi:10.1089/fpd.2009.0469.
- 41. Yu S, Yu P, Wang J, Li C, Guo H, Liu C, et al. A study on prevalence and characterization of Bacillus cereus in ready-to-eat foods in China. Frontiers in microbiology. 2020;10:3043. Doi:10.3389/fmicb.2019.03043.
- 42. Gupta V, Gulati P, Bhagat N, Dhar M, Virdi J. Detection of Yersinia enterocolitica in food: an overview. European Journal of Clinical Microbiology & Infectious Diseases. 2015;34(4):641-50. Doi:10.1007/s10096-014-2276-7.
- Lucero-Estrada C, Favier GI, Escudero ME. An overview of Yersinia enterocolitica and related species in samples of different origin from San Luis, Argentina. Food microbiology. 2020;86:103345. Doi:10.1016/j.fm.2019.103345.
- 44. Mahdavi S, Farshchian MR, Amini K, Abbasi M, Rad MG, Ebadi AR. Survey of Yersinia enterocolitica contamination in distributed broiler meats in Tabriz City, Iran. African J Microbiol Res. 2012;6(12):3019-23. Doi: 10.5897/AJMR12.094.
- Zadernowska A, Chajęcka-Wierzchowska W, Łaniewska-Trokenheim Ł. Yersinia enterocolitica: a dangerous, but often ignored, foodborne pathogen. Food Reviews International. 2014;30(1):53-70. Doi:10.1080/87559129.2013.85377 5.
- Chlebicz A, Śliżewska K. Campylobacteriosis, salmonellosis, yersiniosis, and listeriosis as zoonotic foodborne diseases: a review. International journal of environmental research and public health. 2018;15(5):863. Doi:10.3390/ijerph15050863
- Soro AB, Whyte P, Bolton DJ, Tiwari BK. Strategies and novel technologies to control Campylobacter in the poultry chain: A review. Comprehensive Reviews in Food Science and Food Safety. 2020. Doi:10.1111/1541-4337.12544.
- Rahimi E, Ameri M, Alimoradi M, Chakeri A, Bahrami AR. Prevalence and antimicrobial resistance of Campylobacter jejuni and Campylobacter coli isolated from raw camel, beef,

and water buffalo meat in Iran. Comparative Clinical Pathology. 2013;22(3):467-73. Doi:10.1007/s00580-012-1434-5.

- 49. Thames H, Sukumaran A. A Review of Salmonella and Campylobacter in Broiler Meat: Emerging Challenges and Food Safety Measures. Foods. 2020;9(6):776. Doi:10.3390/ foods9060776.
- Quaglia NC, Dambrosio A. Helicobacter pylori: A foodborne pathogen? World journal of gastroenterology. 2018;24(31):3472. Doi: 10.3748/wjg.v24.i31.3472.
- Bahrami AR, Rahimi E, Ghasemian Safaei H. Detection of Helicobacter pylori in city water, dental units' water, and bottled mineral water in Isfahan, Iran. The Scientific World Journal. 2013;2013. Doi:10.1155/2013/280510.
- 52. Souod N, Kargar M, Doosti A, Ranjbar R, Sarshar M. Genetic analysis of cagA and vacA genes in Helicobacter pylori isolates and their relationship with gastroduodenal diseases in the west of Iran. Iranian Red Crescent Medical Journal. 2013;15(5):371. Doi: 10.5812/ircmj.3732.
- Fujimura S, Kawamura T, Kato S, Tateno H, Watanabe A. Detection of Helicobacter pylori in cow's milk. Letters in Applied Microbiology. 2002;35(6):504-7. Doi:10.1046/j.1472-765X.2002.01229.x.
- 54. Quaglia N, Dambrosio A, Normanno G, Parisi A, Patrono R, Ranieri G, et al. High occurrence of Helicobacter pylori in raw goat, sheep and cow milk inferred by glmM gene: a risk of food-borne infection? International journal of food microbiology. 2008;124(1):43-7. Doi:10.1016/j.ijfoodmicro.2008.02.011.
- Meng X, Zhang H, Law J, Tsang R, Tsang T. Detection of Helicobacter pylori from food sources by a novel multiplex PCR assay. Journal of food safety. 2008;28(4):609-19. Doi:10.1111/ j.1745-4565.2008.00135.x.
- Mirnejad R, Jazi FM, Mostafaei S, Sedighi M. Epidemiology of brucellosis in Iran: A comprehensive systematic review and meta-analysis study. Microbial pathogenesis. 2017;109:239-47. Doi:10.1016/j.micpath.2017.06.005.
- 57. Dadar M, Fakhri Y, Shahali Y, Khaneghah AM. Contamination of milk and dairy products by Brucella species: A global systematic review and meta-analysis. Food Research International. 2020;128:108775. Doi:10.1016/j.foodres.2019.108775.
- 58. Moslemi E, Soltandalal MM, Beheshtizadeh MR, Taghavi A, Kheiri Manjili H, Mahmoudi Lamouki R, et al. Detection of Brucella spp. in dairy products by real-time PCR. Archives of Clinical Infectious Diseases. 2018;13(1). Doi:10.5812/archcid.12673.
- 59. Esmaeili S, Naddaf SR, Pourhossein B, Hashemi Shahraki A, Bagheri Amiri F, Gouya MM, et al. Seroprevalence of brucellosis, leptospirosis, and Q fever among butchers and slaughterhouse workers in south-eastern Iran. PloS one.

2016;11(1):e0144953. Doi:10.1371/journal.pone.0144953.

- 60. Islam MS, Islam MA, Rahman MM, Islam K, Islam MM, Kamal MM, et al. Presence of Brucella spp. in milk and dairy products: a comprehensive review and its perspectives. Journal of Food Quality. 2023;2023(1):2932883. Doi:10.1371/ journal.pone.0144953.
- Letchumanan V, Chan K-G, Lee L-H. Vibrio parahaemolyticus: a review on the pathogenesis, prevalence, and advance molecular identification techniques. Frontiers in microbiology. 2014;5:705. Doi:10.3389/fmicb.2014.00705.
- Odeyemi OA. Incidence and prevalence of Vibrio parahaemolyticus in seafood: a systematic review and meta-analysis. Springerplus. 2016;5(1):464. Doi:10.1186/s40064-016-2115-7.
- 63. Li Y, Xie T, Pang R, Wu Q, Zhang J, Lei T, et al. Food-Borne Vibrio parahaemolyticus in China: Prevalence, Antibiotic Susceptibility, and Genetic Characterization. Frontiers in microbiology. 2020;11:1670. Doi:10.3389/fmicb.2020.01670.
- 64. Shahin K, Bouzari M, Wang R, Yazdi M. Prevalence and molecular characterization of multidrug-resistant Shigella species of food origins and their inactivation by specific lytic bacteriophages. International journal of food microbiology. 2019;305:108252. Doi:10.1016/j.ijfoodmicro.2019.108252.
- 65. Gundappa M, Gaddad SM. Prevalence of Salmonella, Shigella and E. coli in Vegetables of Various Markets in Kalaburagi (India). Indian J Nat Sci. 2016;6(1):1-5. Doi:10.1007/s13197-017-2964-2.
- 66. Shahin K, Bouzari M. Bacteriophage application for biocontrolling Shigella flexneri in contaminated foods. Journal of food science and technology. 2018;55(2):550-9.
- Rahman S, Park J, Song KB, Al Harbi NA, Oh DH. Effects of slightly acidic low concentration electrolyzed water on microbiological, physicochemical, and sensory quality of fresh chicken breast meat. Journal of food science. 2012;77(1):M35-M41. Doi:10.1111/j.1750-3841.2011.02454.x.
- Sofos JN. Meat and meat products. Food safety management: Elsevier; 2014. p. 119-62. Doi:10.1016/B978-0-12-381504-0.00006-8.
- Pesavento G, Ducci B, Nieri D, Comodo N, Nostro AL. Prevalence and antibiotic susceptibility of Listeria spp. isolated from raw meat and retail foods. Food control. 2010;21(5):708-13. Doi:10.1016/j.foodcont.2009.10.012.
- Centers for Disease Control and Prevention [Internet].
 2011. Available from: https://www.cdc.gov/foodborneburden/2011-foodborne-estimates.html.
- 71. Tack DM. Preliminary incidence and trends of infections with pathogens transmitted commonly through food—Food-

borne Diseases Active Surveillance Network, 10 US sites, 2015–2018. MMWR Morbidity and mortality weekly report. 2019;68.

- 72. Rahimi Z, Mahmoudi R, Ghajarbeyg P, Mosavi S, Mehrabi A. The Prevalence of Salmonella Enteritidis in Packaged and Tray Eggs Samples, Qazvin, Iran. Journal of Chemical Health Risks. 2021. Doi:10.22034/jchr.2021.1922123.1250.
- 73. Nazari Moghadam M, Rahimi E, Shakerian A, Momtaz H. Prevalence of Salmonella Typhimurium and Salmonella Enteritidis isolated from poultry meat: virulence and antimicrobial-resistant genes. BMC microbiology. 2023;23(1):1-8. Doi:10.1186/s12866-023-02908-8.
- 74. Saram M, Nosrati AC, Modiri L, Issazadeh K. Investigation of Genetic-molecular Sources of Multidrug Resistance of Salmonella Isolated from Eggs. Social Determinants of Health.8:1-11. Doi:10.22037/sdh.v8i1.39325.
- 75. Hassanzadeh P, Mohammadzadeh S. Antimicrobial Effects of Different Synbiotic Compounds against Pathogenic Bacteria Isolated from Beef, Mutton, and Chicken. Archives of Razi Institute. 2022;77(6):2105. Doi: 10.22092/ ARI.2022.357834.2107.
- 76. Shayegh J, Shahbazi G, Ghazaei S, Movassagh Ghazani MH, Hanifian S. Prevalence, virulence factor and antibiotics susceptibility patterns of Salmonella spp. from poultry products in Ardabil province. 2022.
- 77. Rahimi Z, Ghajarbeygi P, Mahmoudi R, Mosavi S, Mehrabi A. PREVALENCE OF SALMONELLA STRAINS ISO-LATED FROM INDUSTRIAL QUAIL EGGS AND LOCAL DUCK EGGS, IRAN. Carpathian Journal of Food Science & Technology. 2022;14(2). Doi:10.34302/crpjfst/2022.14.2.14.
- Mir R, Salari S, Najimi M, Rashki A. Determination of frequency, multiple antibiotic resistance index and resistotype of Salmonella spp. in chicken meat collected from southeast of Iran. Veterinary Medicine and Science. 2022;8(1):229-36. Doi:10.1002/vms3.647.
- Kazemeini H. Molecular detection of Campylobacter species and Salmonella spp. In cattle raw milk specimens in Mazandaran province. Journal of food science and technology (Iran). 2022;19(125):101-8. Doi:10.22034/FSCT.19.125.101.
- 80. Norbakhsh F, Behshood P. Determination of molecular properties of Salmonella isolates isolated from red meat in Shahrekord, Iran. Journal of Zoonosis. 2022;2(1):18-25.
- Azizpour A. Prevalence and antibiotic resistance of Salmonella serotypes in chicken meat of Ardabil, Northwestern Iran. Iranian Journal of Medical Microbiology. 2021;15(2):232-46. Doi: 10.30699/ijmm.15.2.232.
- 82. Bahramianfard H, Derakhshandeh A, Naziri Z, Khaltabadi Farahani R. Prevalence, virulence factor and antimi-

crobial resistance analysis of Salmonella Enteritidis from poultry and egg samples in Iran. BMC veterinary research. 2021;17(1):196. Doi:10.1186/s12917-021-02900-2.

- Sabzali S, Bouzari M. Isolation, identification and some characteristics of two lytic bacteriophages against Salmonella enterica serovar Paratyphi B and S. enterica serovar Typhimurium from various food sources. FEMS Microbiology Letters. 2021;368(7):fnab037. Doi:10.1093/femsle/fnab037.
- 84. Afshari A, Tavassoli M, Ram M, Ranjbar G. Assessment of Microbial and Chemical Quality of a Ready to Eat Food, Olivier Salad, in Mashhad City. Journal of Nutrition, Fasting and Health. 2019;7(4 (Spe):175-81.
- Forouhar M, Harzandi N. Molecular Survey of Quinolone Resistance in Salmonella spp. Isolated From Poultry Products in Karaj, Iran. Int J Enteric Pathog. 2019;7(2):55-9. Doi: 10.15171/ijep.2019.13.
- Mahdavi M, Jalali M, Safaei HG, Shamloo E. Microbial quality and prevalence of Salmonella and Listeria in eggs. International Journal of Environmental Health Engineering. 2012;1(1):48.
- Hamzeh Pour S, Vaziri S, Molaee Aghaee E. Survey on the contamination rate and determination of antibiotic resistance of Staphylococcus aureus, Escherichia coli and Salmonella strains isolated from traditional cheeses distributed in Mahabad, Iran. Iranian Journal of Health and Environment. 2019;11(4):465-76.
- Mahdavi S, Azizi Dehbokri M, Isazadeh A. Contamination of chicken meat with salmonella spp distributed in mahabad city, iran. Int J Enteric Pathog. 2018;6(3):65-8. Doi: 10.15171/ ijep.2018.18
- Hashemi M, Baygan A, Balouchzehi Z, Dousti Nouri M, Afshari A. Evaluation of microbial quality of traditional sweets, Zoolbia and Bamieh, during Ramadan in Mashhad, Iran. Journal of Nutrition, Fasting and Health. 2018;6(3):145-9. Doi:10.22038/JNFH.2018.34900.1147.
- Karimiazar F, Soltanpour MS, Aminzare M, Hassanzadazar H. Prevalence, genotyping, serotyping, and antibiotic resistance of isolated Salmonella strains from industrial and local eggs in Iran. Journal of Food Safety. 2019;39(1):e12585. Doi:10.1111/jfs.12585.
- Rezaee R, Mohammadi S, Faizee S. A study of salmonella spp. contamination of eggs in Sanandaj in 2017. Zanko Journal of Medical Sciences. 2018;19(61):43-9.
- 92. Khedmati Morasa H, Mahmoudi R, Ghajarbeygi P, Abasi N, Mousavy S. Effects of cold-water egg shell washing on Salmonella contamination in the shell and its contents. The Journal of Qazvin University of Medical Sciences. 2018;22(2):83-9.
- 93. Sana M, Hosseini Siahi Z. The determination of total micro-

bial count and prevalence of Salmonella in the shrimp supply in Khuzestan province. Iranian Journal of Health and Environment. 2018;11(2):149-56.

- 94. Eslami A, Gholami Z, Nargesi S, Rostami B, Avazpour M. Evaluation of microbial contamination of ready-to-eat foods (pizza, frankfurters, sausages) in the city of Ilam. Environmental Health Engineering and Management Journal. 2017;4(2):117-22. Doi:10.15171/EHEM.2017.16.
- Rahimi E, Shakeriyan A. Salmonella contamination of camel meat in various stages of destruction in Isfahan and Chaharmahal va Bakhtiari. Journal of Food Microbiology. 2017;4(2):29-36.
- 96. Sharafati Chaleshtori R, Mazroii Arani N, Taghizadeh M, Sharafati Chaleshtori F. Antibiotic resistance pattern of salmonella isolated from hamburgers and detection of their sensitivity to some essential oils. Journal of Mazandaran University of Medical Sciences. 2017;27(148):136-42.
- 97. Mojaddar Langroodi A, Ehsani A, Ebadi A. Comparison of bacteria in beef meat and poultry in terms of packaging and without packaging in Gilan province. Journal of Food Microbiology. 2016;3(1):33-42.
- 98. Dousandeh S, Asaadi Tehrani G, Amini B, Maziri P. Simultaneous detection of invA, STM4497, and fliC183 genes in Salmonella typhimurium by multiplex PCR method in poultry meat samples in Zanjan, Iran. Infection, Epidemiology and Microbiology. 2016;2(4):20-3.
- 99. Vahabi AN, Abbasvali M. Determination of microbial contamination of olovie salads consumed in Isfahan. 2016.
- 100. Khammar F, Alipour Eskandari M, Saadati D. Study of Salmonella contamination of traditional ice creams in Zabol City, Iran. Iranian Journal of Medical Microbiology. 2017;11(1):83-9.
- 101. Asadi S, Maram ZR, Kooshk F. Evaluation of microbial contamination of pastry cream in Arak city of Iran. Journal of Food Safety and Hygiene. 2015;1(1):26-9.
- 102. Tajbakhsh F, Tajbakhsh E, Momeni M. Detection of Staphylococcus aureus and Salmonella typimurium in traditional and industrial Olivier salads in shahrekord city. 2015.
- 103. Khodadadipour T, Amini K, Mahmoudi R. Evaluation of virulence and enterotoxin genes in Salmonella enteritidis strains isolated from Meat and Egg samples by Multiplex-PCR. Journal of Food Microbiology. 2016;3(2):25-33.
- 104. Ghorbani Ranjbary A. Study of Drug Resistance in Salmonella spp. Isolated from Native Eggs of Iran's Southern Region. Journal of pure and applied microbiology. 2015;9(2):175-9.
- 105. Khoramrooz S, Sarikhani M, Khosravani S, Farhang Falah M,

- 106. Doosti A, Doosti E, Rahimi E, Ghasemi-Dehkordi P. Frequency of antimicrobial-resistant genes in Salmonella enteritidis isolated from traditional and industrial iranian white cheeses. Proceedings of the National Academy of Sciences, India Section B: Biological Sciences. 2017;87(1):73-80. Doi:10.1007/s40011-015-0572-3.
- 107. Moosavy M-H, Esmaeili S, Amiri FB, Mostafavi E, Salehi TZ. Detection of Salmonella spp in commercial eggs in Iran. Iranian journal of microbiology. 2015;7(1):50.
- 108. Kheyri A, Fakhernia M, Haghighat-Afshar N, Hassanzadazar H, Kazemi-Ghoshchi B, Zeynali F, et al. Microbial contamination of meat products produced in the factories of West Azerbaijan Province, North West of Iran. Global Veterinaria. 2014;12(6):796-802. Doi: 10.5829/idosi.gv.2014.12.06.83145.
- 109. Soltan Dallal MM, Emadi Koochak H, Sharifi Yazdi MK, Taheri Mirghaed A, Choobineh H. Determination of yersinia Spp. and Salmonella paratyphi B isolated from possibly contaminated cream samples in the city of Tehran. Journal of Payavard Salamat. 2014;8(1):34-43.
- 110. Dallal MMS. Prevalence of Salmonella spp. in packed and unpacked red meat and chicken in south of Tehran. Jundishapur Journal of Microbiology. 2014;7(4).
- 111. Sodagari HR, Mashak Z, Ghadimianazar A. Prevalence and antimicrobial resistance of Salmonella serotypes isolated from retail chicken meat and giblets in Iran. The Journal of Infection in Developing Countries. 2015;9(05):463-9. Doi: 10.3855/jidc.5945.
- 112. Zahedi M, Rahimi E, Zahedi M, Momtaz H. Prevalence of Salmonella enteritidis and S. typhimurium in marketed meat in Shahrekord in 2014. Journal of Shahrekord Uuniversity of Medical Sciences. 2017;19.
- 113. Momtaz H, Dehkordi FS, Rahimi E, Asgarifar A. Detection of Escherichia coli, Salmonella species, and Vibrio cholerae in tap water and bottled drinking water in Isfahan, Iran. BMC public health. 2013;13(1):556. Doi:10.1186/1471-2458-13-556.
- 114. Bidaki MZ, Pour AT, Dadpour S, Gholizadeh H. Prevalence of Salmonella in poultry carcasses serotypes in Birjand industrial slaughterhouses. Journal of Birjand University of Medical Sciences. 2013;20(02).
- 115. Monadi M, Kargar M, Naghiha A, Mohammadi R. Salmonella contamination of eggs of native Kohgiluyeh va Boyerahmad using PCR1 techniques and the evaluation of drug resistance. Armaghane danesh. 2014;19(2):179-87.
- 116. Amirmozaffari N, Rahmani Z, Iesazadeh K. Evaluation of the level of contamination with Salmonella spp. in red meat,

IRANIAN JOURNAL OF VETERINARY SCIENCE AND TECHNOLOGY

chicken, and domestic and industrial eggs produced in Talesh city and assessment of their antibiotic resistance pattern, Iran. Qom University of Medical Sciences Journal. 2013;7(5):60-5.

- 117. Kaseb F, Shiranian M, Abdar M, Aminalroayaei H, Fallahzadeh H. The prevalence of salmonella and Staphylococcus aureus in industrial Olivier salad in Yazd in 2013. Tolooebehdasht. 2015;14(3):51-9.
- 118. Rahimi E, Shakerian A, Falavarjani AG. Prevalence and antimicrobial resistance of Salmonella isolated from fish, shrimp, lobster, and crab in Iran. Comparative Clinical Pathology. 2013;22(1):59-62. Doi:10.1007/s00580-011-1368-3.
- 119. Rahimi E. Prevalence and antimicrobial resistance of Salmonella spp isolated from retail chicken, turkey and ostrich by-products in Iran. Revue de MédecineVétérinaire. 2012;163(52):271-5. Doi:10.1007/s00580-011-1368-3.
- 120. Karimi Darehabi H, Esmailneshad F, Ebrahimi Mohammadi K. Contamination of Fresh Beef to Salmonella typhimurium and Salmonella enteritidis in Sanandaj during 2012. Food Hygiene. 2012;2(3 (7)):41-8.
- 121. Vaskas AT, Dahesht AE, Seifi S, Rahmani M, Motaghifar A, Safanavaee R. Study and comparison of the bacterial contamination outbreak of chicken meat consumed in some cities of Mazandaran Province (Iran). African Journal of Microbiology Research. 2012;6(33):6286-90. Doi: 10.5897/AJMR12.984.
- 122. Zarei M, Maktabi S, Ghorbanpour M. Prevalence of Listeria monocytogenes, Vibrio parahaemolyticus, Staphylococcus aureus, and Salmonella spp. in seafood products using multiplex polymerase chain reaction. Foodborne Pathogens and Disease. 2012;9(2):108-12. Doi:10.1089/fpd.2011.0989.
- 123. Tajbakhsh F, Tajbakhsh E, Momeni M, Rahimi E, Sohrabi R. Occurrence and Antibiotic Resistance of Salmonella spp isolated from raw Cow's Milk from Shahahrekord. Iran Inter J Microbiol Res. 2012;3:242-5. Doi: 10.5829/idosi. ijmr.2012.3.3.61302.
- 124. Safaei HG, Jalali M, Hosseini A, Narimani T, Sharifzadeh A, Raheimi E. The prevalence of bacterial contamination of table eggs from retails markets by Salmonella spp., Listeria monocytogenes, Campylobacter jejuni and Escherichia coli in Shahrekord, Iran. Jundishapur Journal of Microbiology. 2011;4(4).
- 125. Dallal MMS, Doyle MP, Rezadehbashi M, Dabiri H, Sanaei M, Modarresi S, et al. Prevalence and antimicrobial resistance profiles of Salmonella serotypes, Campylobacter and Yersinia spp. isolated from retail chicken and beef, Tehran, Iran. Food Control. 2010;21(4):388-92. Doi:10.1016/j.food-cont.2009.06.001.
- 126. Rahimi E, Ameri M, Kazemeini H, Elbagi M. PREVALENCE AND ANTIMICROBIAL RESISTANCE OF SALMONEL-LA ISOLATED FROM RETAIL RAW TURKEY, OSTRICH, AND PARTRIDGE MEAT IN IRAN. Bulgarian Journal of

Veterinary Medicine. 2010;13(1).

- 127. Jamshidi A, Kalidari GA, Hedayati M. Isolation and identification of Salmonella Enteritidis and Salmonella Typhimurium from the eggs of retail stores in Mashhad, Iran using conventional culture method and multiplex PCR assay. Journal of food safety. 2010;30(3):558-68. Doi:10.1111/j.1745-4565.2010.00225.x.
- 128. Jamshidi Ae, basami m, afshari ns. Identification of Salmonella spp. and Salmonella typhimurium by a multiplex PCRbased assay from poultry carcasses in Mashhad-Iran. 2009.
- 129. Shapouri R, Rahnema M, Eghbalzadeh S. PREVALENCE OF SALMONELLA SEROTYPES IN POULTRY MEAT AND EGG AND DETERMINE THEIR ANTIBIOTIC SENSIVITY IN ZANJAN CITY. 2009.
- 130. Soltan Dmm, Taremi M, Gachkar L, MODARESI S, Sanaei M, Bakhtiari R, et al. Characterization of antibiotic resistant patterns of Salmonella serotypes isolated from beef and chicken samples in Tehran. 2009.
- 131. Namaei M, Ziaee M, Kafi MG. Prevalence of salmonella contamination in locally (non-industrially) produced eggs in Birjand (2006). Journal of Birjand University of medical sciences. 2009;16(02).
- 132. Jalali M, Abedi D, Pourbakhsh SA, Ghoukasin K. Prevalence of salmonella spp. in raw and cooked foods in Isfahan Iran. Journal of food safety. 2008;28(3):442-52. Doi:10.1111/j.1745-4565.2008.00122.x.
- 133. AR B. Qualitative and quantitative assessment of poultry carcasses contaminated with salmonella in Tehran industrial slaughter houses. 2008.
- 134. Daneshmand A, Rasooli M, Kargar M, Kyani S. A survey on bacterial contamination of fresh traditional cheeses with Salmonella spp. and Staphylococcus aureus in Jahrom, Fars Province. ISMJ. 2007;10(1):19-26.
- 135. Bonyadian M, Ale AS, MOTAHARI FA. Isolation and identification of Salmonellae from chicken carcasses in processing plants in Yazd province, central Iran. 2007.
- 136. Namaei M, ZIAEI M, GHANAD KM. Prevalence of salmonella contamination in locally (non-industrially) produced eggs in Birjand (2006). 2009.
- 137. Nourbakhsh SA, Rahimi E. The occurrence of some foodborne pathogens recovered from poultry meat in Shahrekord, Iran. Journal of Advanced Veterinary and Animal Research. 2023;10(2):205. Doi: 10.5455/javar.2023.j670.
- 138. Azad A, Shakerian A. Evaluation of Staphylococcus Aureus, Escherichia Coli, and Salmonella Contamination in Sausages and Bologna Sold in Tehran City, Iran. Journal of Alternative

Veterinary Medicine | Spring. 2023;6(16).

- 139. Azimirad M, Nadalian B, Alavifard H, Panirani SN, Bonab SMV, Azimirad F, et al. Microbiological survey and occurrence of bacterial foodborne pathogens in raw and ready-to-eat green leafy vegetables marketed in Tehran, Iran. International Journal of Hygiene and Environmental Health. 2021;237:113824. Doi:10.1016/j.ijheh.2021.113824.
- 140. Doudaran HO, Mahsk Z, Kohdar V. Distribution Of Antibiotic Resistance Genes Amongst The Staphylococcus Aureus Strains Isolated From Raw Milk And Traditional Dairy Products. Journal of Pharmaceutical Negative Results. 2022:9735-41.
- 141. Feizolahnezhad A, Mashak Z. Study The Prevalence And Antibiotic Resistance Of Staphylococcus Aureus Bacteria Isolated From Street Ready To Eat Food Samples. Journal of Pharmaceutical Negative Results. 2022:9714-20. Doi:10.47750/ pnr.2022.13.S09.1135.
- 142. Kabiri E, Mashak Z. Antimicrobial resistance properties and enterotoxigenic gene profile of methicillin-resistant and methicillin-susceptible Staphylococcus aureus isolates from raw milk. Journal of Positive School Psychology. 2022;6(8):1244-52.
- 143. Pajohesh R, Tajbakhsh E, Momtaz H, Rahimi E. Relationship between Biofilm Formation and Antibiotic Resistance and Adherence Genes in Staphylococcus aureus Strains Isolated from Raw Cow Milk in Shahrekord, Iran. International Journal of Microbiology. 2022;2022. Doi:10.1155/2022/6435774.
- 144. Hassani S, Pakbin B, Allahyari S, Mahmoudi R, Mousavi S, Ghajarbeygi P. PREVALENCE AND ANTIMICROBI-AL SUSCEPTIBILITY OF FOODBORNE BACTERIAL PATHOGENS ISOLATED FROM BAGHLAVA AN IRANI-AN EXPORTING PASTRY SWEET. Carpathian Journal of Food Science & Technology. 2022;14(3). Doi:10.34302/crpjfst/2022.14.3.5.
- 145. Shariatifar M, Salavati-Hamedani M, Rezaeian-Doloei R, Rezaeigolestani M, Mohsenzadeh M. Antibiotic resistance and enterotoxin gene profiles of Staphylococcus aureus isolated from raw milk in Iran. Journal of the Hellenic Veterinary Medical Society. 2023;74(1):5433-40. Doi:10.12681/ jhvms.29671.
- 146. Bastam MM, Jalili M, Pakzad I, Maleki A, Ghafourian S. Pathogenic bacteria in cheese, raw and pasteurised milk. Veterinary Medicine and Science. 2021;7(6):2445-9. Doi:10.1002/vms3.604.
- 147. Abbasi K, Tajbakhsh E, Momtaz H. Antimicrobial resistance and biofilm encoding genes amongst the Staphylococcus aureus bacteria isolated from meat and meat products. Egyptian Journal of Veterinary Sciences. 2021;52(1):55-62. Doi:10.21608/EJVS.2020.39385.1186.
- 148. Maktabi S, Gharibi D, Hormozi T. Detection of Enterotoxin A

Prevalence of Foodborne Bacteria in Food in Iran

- 149. Haghi F, Zeighami H, Hajiloo Z, Torabi N, Derakhshan S. High frequency of enterotoxin encoding genes of Staphylococcus aureus isolated from food and clinical samples. Journal of Health, Population and Nutrition. 2021;40(1):1-6. Doi:10.1186/s41043-021-00246-x.
- 150. Mesbah A, Mashak Z, Abdolmaleki Z. A survey of prevalence and phenotypic and genotypic assessment of antibiotic resistance in Staphylococcus aureus bacteria isolated from readyto-eat food samples collected from Tehran Province, Iran. Tropical Medicine and Health. 2021;49:1-12. Doi:10.1186/ s41182-021-00366-4.
- 151. Dadmehr A, Sadighara P, Zeinali T. A study on microbial and chemical characterization of mechanically deboned chicken in Tehran, Iran. International Journal of Environmental Health Research. 2022;32(11):2396-405. Doi.org/10.1080/09 603123.2021.1967889.
- 152. Saadati A, Mashak Z, Yarmand MS. Prevalence of Staphylococcal Cassette Chromosome mec and Panton-Valentine Leukocidin Gene Amongst the Methicillin-resistant Staphylococcus aureus Strains Isolated From Fowl Meat. Int J Enteric Pathog. 2019;7(3):93-8. Doi:10.15171/ijep.2019.20.
- 153. Mahdavi S, Isazadeh A. Investigation of contamination rate and determination of pattern of antibiotic resistance in coagulase positive staphylococcus aureus isolated from domestic cheeses in Maragheh, Iran. Pathobiology Research. 2019;22(2):85-9.
- 154. Dehkordi FS, Gandomi H, Basti AA, Misaghi A, Rahimi E. Phenotypic and genotypic characterization of antibiotic resistance of methicillin-resistant Staphylococcus aureus isolated from hospital food. Antimicrobial Resistance & Infection Control. 2017;6(1):104. Doi:10.1186/s13756-017-0257-1.
- 155. Goudarzi B, Alipour V, Rezaei L, Dindarlu K, Heidari M, Rahmaniyan O. Bacteriological quality of ready to use salads at restaurants in Bandar Abbas Abstract. Journal of Preventive Medicine. 2016;3(3):38-1.
- 156. Varmazyar-najafi M, Pajohi-alamoti M, Mohammadzadeh A, Mahmoodi P. Detection of methicillin-resistance gene in Staphylococcus aureus isolated from traditional white cheese in Iran. Archives of Hygiene Sciences. 2016;5(4):302-9.
- 157. Arfatahery N, Davoodabadi A, Abedimohtasab T. Characterization of toxin genes and antimicrobial susceptibility of Staphylococcus aureus isolates in fishery products in Iran. Scientific reports. 2016;6(1):1-7. Doi:10.1038/srep34216.
- 158. Arabestani MR, Kazemian H, Tabar ZK, Hosseini SM. Prevalence of virulence genes, agr and antimicrobial resistance of Staphylococcus aureus isolated from food and dairy products

in Hamadan, Iran. Der Pharmacia Lettre. 2016;8(8):62-7.

- 159. Dehkordi AA, Tajbakhsh E, Tajbakhsh F, Khamesipour F, Shahraki MM, Momeni H. Molecular typing of staphylococcus aureus strains from Iranian raw milk and dairy products by coagulase gene polymorphisms. Adv Stud Biol. 2015;7(4):169-77. Doi:10.12988/asb.2015.516.
- 160. Arfatahery N, Mirshafiey A, Abedimohtasab T, Zeinolabedinizamani M. Study of the prevalence of Staphylococcus aureus in marine and farmed shrimps in Iran aiming the future development of a prophylactic vaccine. Procedia in Vaccinology. 2015;9:44-9. Doi:10.1016/j.provac.2015.05.008.
- 161. Jamali H, Paydar M, Radmehr B, Ismail S, Dadrasnia A. Prevalence and antimicrobial resistance of Staphylococcus aureus isolated from raw milk and dairy products. Food Control. 2015;54:383-8. Doi:10.1016/j.foodcont.2015.02.013.
- 162. Sadeghi M. The prevalence of Enterotoxigenic Isolates of Staphylococcus aureus in Bovine and Sheep Bulk Tank Milk Samples of Maku. 2015. Doi:10.1016/j.foodcont.2015.02.013.
- 163. Eslami M, Koohi M, Zadehashem E, Khadiri B, Keshavarz H. Survey the presence of coagulase positive staphylococcus aureus in cottage cheeses produced from sheep milk and sold in Marand county. Journal of Food Science & Technology (2008-8787). 2014;12(46).
- 164. Madahi H, Rostami F, Rahimi E, Dehkordi FS. Prevalence of enterotoxigenic Staphylococcus aureus isolated from chicken nugget in Iran. Jundishapur journal of microbiology. 2014;7(8). Doi: 10.5812/jjm.10237.
- 165. Khoshkholgh MR, Isazadeh K. Evaluation of Microbial quality of food samples collected in the East region of Gilan. Journal of Food Microbiology. 2015;2(1):27-37.
- 166. Shabani S, Sadeghi Mahoonak A, Jalali H. Microbial contamination of pastry cream supplied in Gorgan. Medical Laboratory Journal. 2014;8(2):62-6.
- 167. Saadat YR, Fooladi AAI, Shapouri R, Hosseini MM, Khiabani ZD. Prevalence of enterotoxigenic Staphylococcus aureus in organic milk and cheese in Tabriz, Iran. Iranian journal of microbiology. 2014;6(5):345.
- 168. Fadaei A. Bacteriological quality of raw cow milk in Shahrekord, Iran. Veterinary World. 2014;7(4):240-3.
- 169. Al-musawi MHJ, Al-Rubaye MTS, Hosseini M. Culture and Molecular Detection of Staphylococcus aureus in Dairy Products of Ahwaz. Al-Mustansiriyah Journal for Pharmaceutical Sciences. 2014;14(2):103-7.
- 170. Farajvand N, Alimohammadi M. Prevalence of Staphylococcus aureus in Four Famous Brand ofDoogh Produced in Iran. Iranian Journal of Health and Environment. 2014;7(1):85-

94.

- 171. Shojaei M, KARIMI DH, Javadi A. Distribution of genes encoding biofilm production in S. aureus isolated from raw milk in Kurdistan. 2014.
- 172. Sepidarkish M, Ghane M. Isolation, identification and the presence of enterotoxin A gene in Staphylococcus aureus from meat products. 2014.
- 173. Shanehbandi D, Baradaran B, Sadigh-Eteghad S, Zarredar H. Occurrence of methicillin resistant and enterotoxigenic Staphylococcus aureus in traditional cheeses in the North West of Iran. International Scholarly Research Notices. 2014;2014. Doi:10.1155/2014/129580.
- 174. Rahimi E. Enterotoxigenicity of Staphylococcus aureus isolated from traditional and commercial dairy products marketed in Iran. Brazilian Journal of Microbiology. 2013;44(2):393-9.
- 175. Rahimi E, Alian F. Presence of enterotoxigenic Staphylococcus aureus in cow, camel, sheep, goat, and buffalo bulk tank milk. Veterinarski arhiv. 2013;83(1):23-30.
- 176. Mirzaei H, Farhoudi H, Tavassoli H, Farajli M, Monadi A. Presence and antimicrobial susceptibility of methicillin-resistant Staphylococcus aureus in raw and pasteurized milk and ice cream in Tabriz by culture and PCR techniques. African Journal of Microbiology Research. 2012;6(32):6224-9. Doi: 10.5897/AJMR12.1701.
- 177. Tavakoli H, Farhang K, AA KZ HE. Bacteriological quality of ready to eat food in four military restaurants. Journal Mil Med. 2012;13(4):207-12.
- 178. Shahraz F, Dadkhah H, Khaksar R, Mahmoudzadeh M, Hosseini H, Kamran M, et al. Analysis of antibiotic resistance patterns and detection of mecA gene in Staphylococcus aureus isolated from packaged hamburger. Meat science. 2012;90(3):759-63. Doi:10.1016/j.meatsci.2011.11.009.
- 179. Alian F, Rahimi E, Shakerian A, Momtaz H, Riahi M, Momeni M. Antimicrobial resistance of Staphylococcus aureus isolated from bovine, sheep and goat raw milk. Global Veterinaria. 2012;8(2):111-4.
- 180. Sharafati Chaleshtori R, Sharafati Chaleshtori F, Karimi A. Antibiotic resistance pattern of staphylococcus strains isolated from orange and apple juices in Shahre-kord, Iran. Pakistan Journal of Medical Sciences. 2010;26(3):615-8.
- 181. Tavakoli HR, Riazipour M. Microbial quality of cooked meat foods in Tehran University's Restaurants. Pakistan Journal of Medical Sciences. 2008;24(4):595-9.
- 182. Basti AA, Misaghi A, Salehi TZ, Kamkar A. Bacterial pathogens in fresh, smoked and salted Iranian fish. Food control. 2006;17(3):183-8. Doi:10.1016/j.foodcont.2004.10.001.

- 183. Raeisi M, Afshari A, Shirzad-Aski H, Seifi S, Hashemi M, Khoshbakht R, et al. The Occurrence of Serotypes and Virulence Genes of Listeria monocytogenes in Various Food Products. Journal of food quality and hazards control. 2023. Doi:10.18502/jfqhc.10.2.12673.
- 184. Kiyanpour Berjoee R, Momtaz H, Lotfollahi L, Bamzaheh Z. Detection and antibiotic resistance pattern of Listeria monocytogenes strains isolated from curd and cheese. Journal of Food Microbiology. 2022;9(4):18-29. Doi:10.30495/jfm.2022.1943942.1741.
- 185. Farahbakhsh S, Kariminik A. Detection of Listeria Monocytogenes in non-Pasteurized Milk in Kerman City by Phenotypic and Molecular Techniques. Journal of Quality and Durability of Agricultural Products and Food Stuffs. 2021;1(1):30-9. Doi:10.30495/qafj.2021.685560.
- 186. Dehnavi M, Khanjari A, Rezaei E. Prevalence of Listeria monocytogenes in traditional cheeses obtained from food sale centers of Tehran, Iran. Journal of Food Safety and Hygiene. 2021;7(3):135-40. Doi:10.18502/jfsh.v7i3.9133.
- 187. Faridi Z, Dallal MMS, Koohy-Kamaly P, Javadi NHS, Hadian Z, Fard RMN. Prevalence and Antimicrobial Resistance of Listeria monocytogenes in Raw Milk in Tehran, Iran. Doi:10.22037/afb.v8i3.34379.
- 188. Mashak Z, Banisharif F, Banisharif G, Reza Pourian M, Eskandari S, Seif A, et al. Prevalence of listeria species and serotyping of Listeria monocytogenes bacteria isolated from seafood samples. Egyptian Journal of Veterinary Sciences. 2021;52(1):1-9. Doi:10.21608/EJVS.2020.17893.1105.
- 189. Farhoumand P, Hassanzadazar H, Soltanpour MS, Aminzare M, Abbasi Z. Prevalence, genotyping and antibiotic resistance of Listeria monocytogenes and Escherichia coli in fresh beef and chicken meats marketed in Zanjan, Iran. Iranian Journal of Microbiology. 2020;12(6):537. Doi: 10.18502/ijm. v12i6.5028.
- 190. Soleimani M, Khalili Sadrabad E, Hamidian N, Heydari A, Akrami Mohajeri F. Prevalence and Antibiotic Resistance of Listeria Monocytogenes in chicken meat retailers in Yazd, Iran. Journal of Environmental Health and Sustainable Development. 2019;4(4):895-902. Doi:10.18502/jehsd.v4i4.2022.
- 191. Akrami-Mohajeri F, Derakhshan Z, Ferrante M, Hamidiyan N, Soleymani M, Conti GO, et al. The prevalence and antimicrobial resistance of Listeria spp in raw milk and traditional dairy products delivered in Yazd, central Iran (2016). Food and Chemical Toxicology. 2018;114:141-4. Doi:10.1016/j. fct.2018.02.006.
- 192. Lotfollahi L, Chaharbalesh A, Rezaee MA, Hasani A. Prevalence, antimicrobial susceptibility and multiplex PCR-serotyping of Listeria monocytogenes isolated from humans, foods and livestock in Iran. Microbial pathogenesis. 2017;107:425-9. Doi:10.1016/j.micpath.2017.04.029.

- 193. Zeinali T, Jamshidi A, Bassami M, Rad M. Isolation and identification of Listeria spp. in chicken carcasses marketed in northeast of Iran. International Food Research Journal. 2017;24(2):881.
- 194. Pournajaf A, Rajabnia R, Sedighi M, Kassani A, Moqarabzadeh V, Lotfollahi L, et al. Prevalence, and virulence determination of Listeria monocytogenes strains isolated from clinical and non-clinical samples by multiplex polymerase chain reaction. Revista da Sociedade Brasileira de Medicina Tropical. 2016;49(5):624-7.
- 195. Abdollahzadeh E, Ojagh SM, Hosseini H, Ghaemi EA, Irajian G, Heidarlo MN. Antimicrobial resistance of Listeria monocytogenes isolated from seafood and humans in Iran. Microbial pathogenesis. 2016;100:70-4. Doi:10.1016/j.micpath.2016.09.012.
- 196. Rahimi E, Jahanmard M, SAFARI S, Ansari M, TORKI Z. PREVALENCE AND ANTIMICROBIAL RESISTANCE OF LISTERIA SPECIES ISOLATED FROM FILLETED AR-GYROSOMUS HOLOLEPIDOTUS, SCOMBEROMORUS COMMERSON AND ALBURNUS SPP. 2016.
- 197. Abbasinejad B, Neyriz-Nagadehi M, Taher Talatappeh N. Prevalence and antimicrobial susceptibility of Listeria monocytogenes in Koozeh cheeses of Urmia retails. Food Hygiene. 2015;5(1 (17)):27-34.
- 198. Maktabi S, Pourmehdi M, Zarei M, Moalemian R. Occurrence and antibiotic resistance of Listeria monocytogenes in retail minced beef distributed in Ahvaz, South-West of Iran. Journal of food quality and hazards control. 2015;2(3):101-6.
- 199. Jamali H, Paydar M, Ismail S, Looi CY, Wong WF, Radmehr B, et al. Prevalence, antimicrobial susceptibility and virulotyping of Listeria species and Listeria monocytogenes isolated from open-air fish markets. BMC microbiology. 2015;15(1):144. Doi:10.1186/s12866-015-0476-7.
- 200. Haghi F, Zeighami H, Naderi G, Samei A, Roudashti S, Bahari S, et al. Detection of major food-borne pathogens in raw milk samples from dairy bovine and ovine herds in Iran. Small Ruminant Research. 2015;131:136-40. Doi:10.1016/j.smallrumres.2015.08.005.
- 201. Abdimoghadam Z, Shamloo E, Atefi M. Frequency of Listeria species in raw milk and traditional dairy products in Isfahan, Iran. Iranian Journal of Nutrition Sciences & Food Technology. 2015;10(3):101-7.
- 202. Mansouri-Najand L, Kianpour M, Sami M, Jajarmi M, editors. Prevalence of Listeria monocytogenes in raw milk in Kerman, Iran. Veterinary Research Forum; 2015: Faculty of Veterinary Medicine, Urmia University, Urmia, Iran.
- 203. Rahimi E, Shakerian A. Prevalence of Listeria Species in Ready-to-Eat Food in Shahrekord Restaurants. Medical Laboratory Journal. 2014;8(2):83-7.

- 204. Haj Hosseini A, Sharifan A, Tabatabaee A. Isolation of Listeria monocytogenes from meat and dairy products. Journal of Medical Microbiology and Infectious Diseases. 2014;2(4):159-62.
- 205. Rahimi E, Momtaz H, Behzadnia A, Baghbadorani ZT. Incidence of Listeria species in bovine, ovine, caprine, camel and water buffalo milk using cultural method and the PCR assay. Asian Pacific Journal of Tropical Disease. 2014;4(1):50-3. Doi:10.1016/S2222-1808(14)60313-3.
- 206. Rahimi E. Prevalence and antimicrobial resistance of Listeria species isolated from smoked and salted fish. Iran J Med Microbiol: Volume. 2014;8(3).
- 207. Jamileh N, SIASIE TE, Baharvand R. Isolation of Listeria monocytogenes from meat products and study on prfA gene in bacteria isolated and comparing with clinical and standard samples. 2013.
- 208. Khamesipour F, Khodadoustan Shahraki A, Moumeni M, Khadivi Boroujeni R, Yadegari M. Prevalence of Listeria monocytogenes in the crayfish (Astacus leptodactylus) by polymerase chain reaction in Iran. International Journal of Biosciences. 2013;3(10):160-9. Doi:10.12692/ijb/3.10.160-169.
- 209. Sohrabi R, Rashedi M, Dorcheh MP, Shahbazi AM. Prevalence of Listeria species in raw milk in Esfahan Province, Iran. African Journal of Microbiology Research. 2013;7(19):2057-60. Doi: 10.5897/AJMR12.1463.
- 210. Jamali H, Paydar M, Looi CY, Wong WF. Prevalence of Listeria species and Listeria monocytogenes serotypes in ready mayonnaise salads and salad vegetables in Iran. African Journal of Microbiology Research. 2013;7(19):1903-6. Doi: 10.5897/AJMR2013.5658.
- 211. Fallah AA, Saei-Dehkordi SS, Mahzounieh M. Occurrence and antibiotic resistance profiles of Listeria monocytogenes isolated from seafood products and market and processing environments in Iran. Food control. 2013;34(2):630-6. Doi:10.1016/j.foodcont.2013.06.015.
- 212. Jamali H, Radmehr B, Thong KL. Prevalence, characterisation, and antimicrobial resistance of Listeria species and Listeria monocytogenes isolates from raw milk in farm bulk tanks. Food Control. 2013;34(1):121-5. Doi:10.1016/j.foodcont.2013.04.023.
- 213. Akya A, Najafi F, Moradi J, Mohebi Z, Adabagher S. Prevalence of food contamination with Listeria spp. in Kermanshah, Islamic Republic of Iran. EMHJ-Eastern Mediterranean Health Journal. 2013; 19 (5), 474-477.
- 214. Momtaz H, Yadollahi S. Molecular characterization of Listeria monocytogenes isolated from fresh seafood samples in Iran. Diagnostic Pathology. 2013;8(1):1-6. Doi:10.1186/1746-1596-8-149

- 215. Dorcheh MP, Sohrabi R, Salajegheh M. Prevalence of Listeria species in retail quail products from Isfahan, Iran. Journal of Veterinary Medicine and Animal Health. 2013;5(1):16-9. Doi: 10.5897/JVMAH12.019.
- 216. Khalili Borujeni F, Moshtaghi H, Bonyadian M. Study on contamination of sheep meat in Shahrekord area with Listeria ivanovii and determination its antibiotic resistance pattern. Iranian Journal of Medical Microbiology. 2013;7(1):15-21.
- 217. Rahimi E, Yazdi F, Farzinezhadizadeh H. Prevalence and antimicrobial resistance of Listeria species isolated from different types of raw meat in Iran. Journal of food protection. 2012;75(12):2223-7. Doi:10.4315/0362-028X.JFP-11-565.
- 218. Fallah AA, Saei-Dehkordi SS, Rahnama M, Tahmasby H, Mahzounieh M. Prevalence and antimicrobial resistance patterns of Listeria species isolated from poultry products marketed in Iran. Food Control. 2012;28(2):327-32. Doi:10.1016/j.foodcont.2012.05.014.
- 219. Rahimi E, Shakerian A, Raissy M. Prevalence of Listeria species in fresh and frozen fish and shrimp in Iran. Annals of Microbiology. 2012;62(1):37-40. Doi:10.1007/s13213-011-0222-9.
- 220. Modaresi R, Mardani K, Tukmechi A, Ownagh A. Prevalence of Listeria spp. in fish obtained from Urmia fish markets. African Journal of Microbiology Research. 2011;5(30):5398-401. Doi: 10.5897/AJMR11.888.
- 221. Rahimi E, Ameri M, Momtaz H. Prevalence and antimicrobial resistance of Listeria species isolated from milk and dairy products in Iran. Food Control. 2010;21(11):1448-52. Doi:10.1016/j.foodcont.2010.03.014.
- 222. Jami S, Jamshidi A, Khanzadi S. The presence of Listeria monocytogenesin raw milk samples in Mashhad, Iran. Iranian Journal of Veterinary Research. 2010;11(4):363-7.
- 223. Mahmoodi MM. Occurrence of Listeria monocytogenes in raw milk and dairy products in Noorabad, Iran. Journal of Animal and Veterinary Advances. 2010;9(1):16-9.
- 224. Jalali M, Abedi D. Prevalence of Listeria species in food products in Isfahan, Iran. International journal of food microbiology. 2008;122(3):336-40. Doi:10.1016/j.ijfoodmicro.2007.11.082.
- 225. Rahimi E, Momtaz H, Hemmatzadeh F. The prevalence of Escherichia coli O157: H7, Listeria monocytogenes and Campylobacter spp. on bovine carcasses in Isfahan, Iran. Iranian Journal of Veterinary Research. 2008;9(4):365-70. Doi: 10.22099/IJVR.2008.2620.
- 226. Shahbazi N, Javadi B, Afshari A, Jamshidi A, Ghavidel M. Identification of Coxiella Burnetti and Mycobacterium SPP through Touch-down PCR Examination in Unpasteurized

Camel Milk in North-East of Iran. Journal of Nutrition, Fasting & Health. 2022;10(4). Doi: 10.22038/JNFH.2022.64095.1382.

- 227. Mokarizadeh K, Ownagh A, Tajik H, editors. Molecular detection of Coxiella burnetii in Kope cheese and cattle milk in West Azerbaijan, Iran. Veterinary Research Forum; 2023: Faculty of Veterinary Medicine, Urmia University, Urmia, Iran. Doi:10.30466/vrf.2022.544750.3323.
- 228. Mobarez AM, Mostafavi E, Khalili M, Esmaeili S. Identification of Coxiella burnetii in raw milk of livestock animal in Iran. International Journal of Microbiology. 2021;2021. Doi:10.1155/2021/6632036.
- 229. Kazemeini H, Partovi R, Nazaktabar A, Shokri H. Detection of Coxiella burnetii in raw milk samples collected from dairy farms in Mazandaran province, north of Iran. Journal of Food Safety and Hygiene. 2022. Doi:10.18502/jfsh.v7i3.9137.
- 230. Nourozhaghi NA, RAHIMI E, SHAKERIAN A. Prevalence of Coxiella burnetii in bovine bulk milk by Nested-PCR in Gilan province. 2020.
- 231. Khademi P, Ownagh A, Ataei B, Kazemnia A, Enferadi A, Khalili M, et al. Prevalence of C. burnetii DNA in sheep and goats milk in the northwest of Iran. International Journal of Food Microbiology. 2020;331:108716. Doi:10.1016/j.ijfood-micro.2020.108716.
- 232. Esmaeili S, Mobarez AM, Khalili M, Mostafavi E. High prevalence and risk factors of Coxiella burnetii in milk of dairy animals with a history of abortion in Iran. Comparative immunology, microbiology and infectious diseases. 2019;63:127-30. Doi:10.1016/j.cimid.2019.01.015.
- 233. Khademi P, Ownagh A, Mardani K, Khalili M. Prevalence of Coxiella burnetii in milk collected from buffalo (water buffalo) and cattle dairy farms in Northwest of Iran. Comparative immunology, microbiology and infectious diseases. 2019;67:101368. Doi:10.1016/j.cimid.2019.101368.
- 234. Reisi M, Rahimi E, Momeni M. Prevalence of Coxiella burnetii in Traditional and Industrial Butter and Cream Using Nested Polymerase Chain Reaction in Shahrekord, Iran. Avicenna Journal of Clinical Microbiology and Infection. 2019;6(2):61-5. Doi: 10.34172/ajcmi.2019.12
- 235. Norouzian H, Diali HG, Azadpour M, Afrough P, Shakib P, Mosavi SM, et al. PCR detection of Coxiella burnetii in milk samples of ruminants, Iran. Journal of Medical Bacteriology. 2018;7(1-2):31-5.
- 236. Rahmdel S, Sadat Moezzi M, Azimzadeh N, Hosseinzadeh S. PCR Detection of Coxiella Burnetii in bovine bulk tank milk samples in Shiraz, southern Iran. International Journal of Nutrition Sciences. 2018;3(4):198-201. Doi: 10.30476/ IJNS.2019.82644.1022.
- 237. Mosleh N, Moslehishad M, Moosakhani F. Detection of Cox-

iella burnetii by Real-Time PCR in Raw Milk and Traditional Cheese Distributed in Tehran Province. Journal of Pharmaceutical & Health Sciences. 2018;6(2):149-55.

- 238. Lorestani S, Jaydari A, Maleki S, Khademi P. Genomic detection of Coxiella burnetii in sheep milk samples by Nested-PCR method in Khorramabad, Iran. 2016.
- 239. Kargar M, Rashidi A, Doosti A, Najafi A, Ghorbani-Dalini S. The sensitivity of the PCR method for detection of Coxiella burnetii in the milk samples. Zahedan Journal of Research in Medical Sciences. 2015;17(6). Doi:10.17795/zjrms988.
- 240. Ahmadizadeh C, Moosakhani F, Jamshidian M. Detection and Identification of Coxiella burnetii in Milk Cattles of Tehran Province. Advances in Bioresearch. 2015;6(4).
- 241. Khalili M, Diali HG, Mirza HN, Mosavi SM. Detection of Coxiella burnetii by PCR in bulk tank milk samples from dairy caprine herds in southeast of Iran. Asian Pacific Journal of Tropical Disease. 2015;5(2):119-22. Doi:10.1016/S2222-1808(14)60638-1.
- 242. Khademi MP. Genomic detection of Coxiella burnetii in cattle milk samples by Nested-PCR method, Iran. Iran J Med Microbiol: Volume. 2015;9(2).
- 243. Khanzadi S, Jamshidi A, Razmyar J, Borji S. Identification of Coxiella burnetii by touch-down PCR assay in unpasteurized milk and dairy products in North-East of Iran. Iranian Journal of Veterinary Medicine. 2014;8(1):15-9.
- 244. Borji S, Jamshidi A, Khanzadi S, Razmyar J. Detection of Coxiella burnetii and sequencing the IS1111 gene fragment in bulk tank milk of dairy herds. Iranian Journal of Veterinary Science and Technology. 2014;6(2):21-8. Doi:10.22067/ VETERINARY.V6I2.26330.
- 245. Amin J, Peyman K, Samira L, Shahram M. DEVELOPMENT OF NESTED-PCR METHOD FOR DETECTING COXIEL-LA BURNETII IN GOAT MILK SAMPLES. Iranian Journal of Public Health. 2014;43(2):171.
- 246. Ghalyanchi Langeroudi A, Babkhani N, Zolfaghari MR, Majidzadeh Arbadili K, Morovvati A, Soleimani M. Detection of Coxeilla brunetii in bulk tank milk samples from dairy bovine farms using nested-PCR in Qom, Iran, 2011. Iranian Journal of Veterinary Medicine. 2013;7(3):207-11.
- 247. Kargar M, Rashidi A, Doosti A, Ghorbani-Dalini S, Najafi A. Prevalence of Coxiella burnetii in bovine bulk milk samples in southern Iran. Comparative clinical pathology. 2013;22(3):331-4. Doi:10.1007/s00580-012-1406-9.
- 248. Rahimi E, Ameri M, Karim G, Doosti A. Prevalence of Coxiella burnetii in bulk milk samples from dairy bovine, ovine, caprine, and camel herds in Iran as determined by polymerase chain reaction. Foodborne pathogens and disease.

2011;8(2):307-10. Doi:10.1089/fpd.2010.0684.

- 249. Rahimi E. Coxiella burnetii in goat bulk milk samples in Iran. African Journal of Microbiology Research. 2010;4(21):2324-6.
- 250. Rahimi E, Doosti A, Ameri M, Kabiri E, Sharifian B. Detection of Coxiella burnetii by nested PCR in bulk milk samples from dairy bovine, ovine, and caprine herds in Iran. Zoonoses and public health. 2010;57(7 8):e38-e41. Doi:10.1111/ j.1863-2378.2009.01289.x.
- 251. Torki Baghbadorani S, Rahimi E, Shakerian A. Investigation of Virulence and Antibiotic-Resistance of Bacillus cereus Isolated from Various Spices. Canadian Journal of Infectious Diseases and Medical Microbiology. 2023;2023. Doi:10.1155/2023/8390778.
- 252. Zeighami H, Nejad-dost G, Parsadanians A, Daneshamouz S, Haghi F. Frequency of hemolysin BL and non-hemolytic enterotoxin complex genes of Bacillus cereus in raw and cooked meat samples in Zanjan, Iran. Toxicology reports. 2020;7:89-92. Doi:10.1016/j.toxrep.2019.12.006.
- 253. Heydarzadeh M, Javadi A, Rad MG. Detection of emetic toxin (ces) gene and antimicrobial susceptibility of Bacillus cereus isolates from Iranian traditional dairy products. 2020. Doi:10.21203/rs.3.rs-61769/v1.
- 254. Deilami Khiabani Z, Noori E, Rahnama M, Shapouri R, Bigdelo Y, Ghamari D, et al. Detection of NHE Complex Genes in Bacillus cereus Isolated from Rice Samples from Zanjan, Iran by Multiplex PCR. Pathobiology Research. 2012;14(4):89-97.
- 255. Heydarzadeh M, Javadi A. Isolation and enumeration of Bacillus cereus in raw milk distributed in Tabriz, Iran and detection of ces gene among the isolates. Food Hygiene. 2018;8(2 (30)):35-42.
- 256. Ghourchian S, Douraghi M, Baghani A, Soltan Dallal M. Bacillus cereus Assessment in Dried Vegetables Distributed in Tehran, Iran. Journal of food quality and hazards control. 2018;5(1):29-32. Doi: 10.29252/jfqhc.5.1.29.
- 257. Hassanzadazar H, Taami B, Abbasi Z, Aminzare M. Microbial Contamination of Cream filled Pastries supplied in Confectioneries of Zanjan, Iran. Journal of Nutrition, Fasting and Health. 2018;6(Issue):30-4. Doi:10.22038/ jnfh.2018.33470.1119.
- 258. Soleimani M, Hosseini H, Pilevar Z, Mehdizadeh M, Carlin F. Prevalence, molecular identification and characterization of Bacillus cereus isolated from beef burgers. Journal of Food Safety. 2018;38(1):e12414. Doi:10.1111/jfs.12414.
- 259. Ranjbar R, Shahreza MHS. Prevalence, antibiotic-resistance properties and enterotoxin gene profile of Bacillus cereus strains isolated from milk-based baby foods. Tropical Journal of Pharmaceutical Research. 2017;16(8):1931-7. Doi:10.4314/

tjpr.v16i8.25.

- 260. Soltan Dallal MM, Nezamabadi S, Mardaneh J, Rajabi Z, Sirdani A. Detection of toxigenic bacillus cereus strains in powdered infant formula (PIF) milk by PCR assay. Tehran University Medical Journal TUMS Publications. 2017;75(3):179-86.
- 261. Rasoulpour T, Mahdavi S. Study of frequency of NHE complex genes in Bacillus cereus isolated from milk in Tabriz city in spring, 2017. 2020.
- 262. Molayi Ks, deilami Kz, Ghasemian A, SHAPOURI R, Taghinejad J, Eslami M, et al. Detection of hbla and bal genes in Bacillus cereus isolates from cheese samples using the polymerase chain reaction. 2016.
- 263. Keshtkar M, Momtaz H, Ghavamizadeh M, Rahimi E. Detection of Enterotoxin Genes Profile of Bacillus cereus in Pasteurized and Sterile Milk, Baby Food and Dairy Products. International Medical Journal. 2016;23(2).
- 264. Mashak Z, Langroodi AM, Ehsani A, Ilkhanipoor A, Fathabad AE. Microbiological quality of ready-to-eat foods of Tehran province. African Journal of Food Science. 2015;9(5):257-61. Doi: 10.5897/AJFS2015.1260.
- 265. Teymori R, Ghazanfarirad N, Dehghan K, Asadzadeh J, Hajigholizadeh G, Bahmani M. A survey of bacterial and mold contamination of imported rice into West Azerbaijan Province, northwest of Iran. Asian Pacific Journal of Tropical Disease. 2014;4:S833-S5. Doi:10.1016/S2222-1808(14)60737-4.
- 266. Rahimi E, Abdos F, Momtaz H, Torki Baghbadorani Z, Jalali M. Bacillus cereus in infant foods: prevalence study and distribution of enterotoxigenic virulence factors in Isfahan Province, Iran. The Scientific World Journal. 2013;2013. Doi:10.1155/2013/292571.
- 267. Hosseini H, Hippe B, Denner E, Kollegger E, Haslberger A. Isolation, identification and monitoring of contaminant bacteria in Iranian Kefir type drink by 16S rDNA sequencing. Food Control. 2012;25(2):784-8. Doi:10.1016/j.food-cont.2011.12.017.
- 268. Rahimifard N, Fatholahzadeh B, Noory Z, Saadati S, Zavar M, Pirouz B, et al. Bacillus cereus contamination in infant formula: a study in food and drug control laboratory. Tehran University Medical Journal TUMS Publications. 2007;65(8):64-8.
- 269. Yazdi SS, Mobasseri B, Bonab PT, Yazdi SS, Mirbagheri Z, Dallal MMS. Prevalence and Characteristics of Yersinia Enterocolitica and Yersinia Pseudotuberculosis from Raw Milk Supplied in Tehran. Journal of Nutrition and Food Security. 2022. Doi:10.18502/jnfs.v8i1.11776.
- 270. Najjar Asiabani F, Bazaei M, Rais Sadat SA. Study of Yersinia enterocolitica contamination in red meat supplied In

Shiraz, Iran. Journal of Zoonotic Diseases. 2021;5(3):28-33. Doi:10.22034/jzd.2021.13491.

- 271. Movafagh F, Zeinali T, Jamshidi A. Contamination Rate of Bovine Raw Milk with Yersinia enterocolitica Biotypes in Mashhad, Iran. Journal of Human Environment and Health Promotion. 2021;7(1):30-4. Doi: 10.29252/jhehp.7.1.5.
- 272. Tavassoli M, Jamshidi A, Ranjbar G, Torabi MR, Moghddam AA. Prevalence, Biotyping, and Antimicrobial Resistance of Yersinia enterocolitica Isolated from Traditional Iranian Cheeses-Evaluation of Yersinia enterocolitica in Traditional Cheeses. City. 2021;1(1B):5.
- 273. Tavassoli M, Jamshidi A, Movafagh F, Afshari A. Virulence Characteristics of Yersinia enterocolitica Isolated from Dairy Products in the Northeast of Iran. Journal of Human, Environment and Health Promotion. 2019;5(2):72-8. Doi:10.29252/jhehp.5.2.5.
- 274. Sirghani K, Zeinali T, Jamshidi A. Detection of Yersinia enterocolitica in retail chicken meat, Mashhad, Iran. Journal of pathogens. 2018. Doi:10.1155/2018/1286216.
- 275. Alavi S, Rahimi E, Tajbakhsh E, Branch S. Prevalence of Yersinia enterocolitica in raw small ruminant milk in Shahrekord, Iran. Bulgarian J Vet Med. 2018;21(3):364-70. Doi: 10.15547/bjvm.1088.
- 276. Hadad A, Tajbakhsh E, Reiysi M. Identification of virulence genes in serotype O: 3 Yersinia enterocolitica isolated from turkey meat by PCR method in Shahrekord. Journal of Food Microbiology. 2018;5(2):1-10.
- 277. Dabiri H, Aghamohammad S, Goudarzi H, Noori M, Hedayati MA, Ghoreyshiamiri SM. Prevalence and antibiotic susceptibility of Campylobacter species isolated from chicken and beef meat. International Journal of Enteric Pathogens. 2016;2(2):6-17087. Doi:10.3168/jds.2014-8853.
- 278. Jamali H, Paydar M, Radmehr B, Ismail S. Prevalence, characterization, and antimicrobial resistance of Yersinia species and Yersinia enterocolitica isolated from raw milk in farm bulk tanks. Journal of dairy science. 2015;98(2):798-803. Doi:10.3168/jds.2014-8853.
- 279. Rahimi E, Sepehri S, Dehkordi FS, Shaygan S, Momtaz H. Prevalence of Yersinia species in traditional and commercial dairy products in Isfahan Province, Iran. Jundishapur Journal of Microbiology. 2014;7(4). Doi: 10.5812/jjm.9249
- 280. Saberianpour S, Tajbakhsh E, Khamesipour F, Branch S. Prevalence of virulence genes and biotyping of Yersinia enterocolitica isolated from chicken meat in Shahrekord, Iran. Vidyabharati International Interdisciplinary Research Journal. 2014;3:71-6.
- 281. Mehdi SDM, Hamid EK, Kazem SYM, Ali TM, Hamid C. Determination Of Yersinia Spp. And Salmonella Paratyphi B

Isolated From Possibly Contaminated Cream Samples In The City Of Tehran. Payavard Salamat. 2014;8(1).

- 282. Momtaz H, Rahimian MD, Dehkordi FS. Identification and characterization of Yersinia enterocolitica isolated from raw chicken meat based on molecular and biological techniques. Journal of Applied Poultry Research. 2013;22(1):137-45. Doi:10.3382/japr.2012-00549.
- 283. Hanifian S, Khani S. Prevalence of virulent Yersinia enterocolitica in bulk raw milk and retail cheese in northern-west of Iran. International Journal of Food Microbiology. 2012;155(1-2):89-92. Doi:10.1016/j.ijfoodmicro.2012.01.012.
- 284. Mahdavi S, Farshchian MR, Amini K, Rad MAG, Ebadi AR. Survey of Yersinia enterocolitica contamination in distributed broiler meats in Tabriz City, Iran. African Journal of Microbiology Research. 2012;6(12):3019-23. Doi: 10.5897/ AJMR12.094.
- 285. MK SY, Soltan-Dallal M, Bakhtiari R. Incidence and antibiotic susceptibilities of Yersinia enterocolitica and other Yersinia species recovered from meat and chicken in Tehran, Iran. African Journal of Microbiology Research. 2011;5(18):2649-53. Doi: 10.5897/AJMR11.248.
- 286. Mirmoeini SS, Sari AA, Goudarztalejerdi A, Alamoti MP, Staji H. Prevalence of Campylobacter spp. Among Broiler Carcasses at Industrial Slaughterhouses in Hamedan, Iran. Archives of Hygiene Sciences. 2023;12(2). Doi: 10.34172/ AHS.12.2.1.393.
- 287. Mousavinafchi SB, Rahimi E, Shakerian A. Campylobacter spp. isolated from poultry in Iran: Antibiotic resistance profiles, virulence genes, and molecular mechanisms. Food Science & Nutrition. 2023;11(2):1142-53. Doi:10.1002/ fsn3.3152.
- 288. Emami MS, Shakerian A, Chaleshtori RS, Rahimi E. Distribution of Virulence Genes in Campylobacter spp. Isolated from Agaricus Mushrooms in Iran. BioMed research international. 2023;2023. Doi:10.1155/2023/1872655.
- 289. Hadiyan M, Momtaz H, Shakerian A. Prevalence, antimicrobial resistance, virulence gene profile and molecular typing of Campylobacter species isolated from poultry meat samples. Veterinary Medicine and Science. 2022;8(6):2482-93. Doi:10.1002/vms3.944.
- 290. Sadeghi A, Ganji L, Fani F, Pouladfar G, Eslami P, Doregiraee F, et al. Prevalence, species diversity, and antimicrobial susceptibility of Campylobacter strains in patients with diarrhea and poultry meat samples: one-year prospective study. Iranian Journal of Microbiology. 2022;14(3):362. Doi: 10.18502/ijm.v14i3.9775.
- 291. Shahreza MS, Dehkordi NG, Nassar MF, Al-Saedi R. Genotyping of Campylobacter jejuni isolates from raw meat of animal species. Academic Journal of Health Sciences: Medicina balear. 2022;47(4):52-7. Doi: 10.3306/AJHS.2022.37.04.52.

- 292. Jahromi RR, Moradi F, Erfanian S, Pourahmadi M. Evaluation of the Contamination of Poultry Carcasses with Campylobacter jejuni and Campylobacter coli in Southern Iran: A Molecular Study. Jundishapur Journal of Health Sciences. 2021;13(3). Doi:10.5812/jjhs.116991.
- 293. Shafiei A, Rahimi E, Shakerian A. Prevalence, Virulence, and Antimicrobial Resistance of Campylobacter Species Isolated From Carcasses of Camels Slaughtered in Slaughterhouses of Chaharmahal and Bakhtiari Province, 2018-2019. Epidemiology and Health System Journal. 2021;8(3):115-21. Doi:10.34172/ijer.2021.22.
- 294. Far hang Zargar M. Identification of Campylobacter species in Jahrom slaughterhouse chickens in 2016-17. Journal of Jahrom University of Medical Sciences. 2019;17(3):1-6. Doi:10.52547/JMJ.17.3.1.
- 295. Maktabi S, Ghorbanpoor M, Hossaini M, Motavalibashi A, editors. Detection of multi-antibiotic resistant Campylobacter coli and Campylobacter jejuni in beef, mutton, chicken and water buffalo meat in Ahvaz, Iran. Veterinary Research Forum; 2019: Faculty of Veterinary Medicine, Urmia University, Urmia, Iran. Doi: 10.30466/vrf.2019.34310.
- 296. Fani F, Aminshahidi M, Firoozian N, Rafaatpour N. Prevalence, antimicrobial resistance, and virulence-associated genes of Campylobacter isolates from raw chicken meat in Shiraz, Iran. Iranian journal of veterinary research. 2019;20(4):283.
- 297. Modirrousta S, Shapouri R, Rezasoltani S, Molaabaszadeh H. Prevalence of Campylobacter spp. and their Common Serotypes in 330 Cases of Red-meat, Chicken-meat and Egg-shell in Zanjan City, Iran. 2016. Doi: 10.7508/iem.2016.01.003.
- 298. Hosseinzadeh S, Mardani K, Aliakbarlu J, Ghorbanzadehghan M. Occurrence of Campylobacter in chicken wings marketed in the northwest of Iran. International Food Research Journal. 2015;22(1).
- 299. Zendehbad B, Khayatzadeh J, Alipour A. Prevalence, seasonality and antibiotic susceptibility of Campylobacter spp. isolates of retail broiler meat in Iran. Food Control. 2015;53:41-5. Doi:10.1016/j.foodcont.2015.01.008.
- 300. Dabiri H, Aghamohammad S, Goudarzi H, Noori M, Ahmadi Hedayati M, Ghoreyshiamiri SM. Prevalence and antibiotic susceptibility of Campylobacter species isolated from chicken and beef meat. Int J Enteric Pathog. 2014;2(2):1-4.
- 301. Rahimi E, Sepehri S, Momtaz H. Prevalence of Campylobacter species in milk and dairy products in Iran. Revue de Médecine Vétérinaire. 2013;164(5):283-8.
- 302. Kazemeini H, Valizade Y, Parsaei P, Nozarpour N, Rahimi E. Prevalence of Campylobacter species in raw bovine milk in Isfahan, Iran. Middle-East J Sci Res. 2011;5:664-6.

- 303. Rahimi E, Ameri M. Antimicrobial resistance patterns of Campylobacter spp. isolated from raw chicken, turkey, quail, partridge, and ostrich meat in Iran. Food Control. 2011;22(8):1165-70. Doi:10.1016/j.foodcont.2011.01.010.
- 304. Rahimi E, Kazemeini HR, Safaei S, Allahbakhshi K, Momeni M, Riahi M. Detection and identification of Campylobacter spp. from retail raw chicken, turkey, sheep and goat meat in Ahvaz, Iran. African journal of microbiology research. 2010;4(15):1620-3.
- 305. Rahimi E, Ameri M, Kazemeini HR. Prevalence and antimicrobial resistance of Campylobacter species isolated from raw camel, beef, lamb, and goat meat in Iran. Foodborne pathogens and disease. 2010;7(4):443-7. Doi:10.1089/ fpd.2009.0421.
- 306. Rahimi E, Tajbakhsh E. Prevalence of Campylobacter species in poultry meat in the Esfahan city, Iran. Bulg J Vet Med. 2008;11(4):257-62.
- 307. Asadi S, Rahimi E, Shakerian A. Helicobacter pylori Strains Isolated from Raw Poultry Meat in the Shahrekord Region, Iran: Frequency and Molecular Characteristics. Genes. 2023;14(5):1006. Doi:10.3390/genes14051006.
- 308. Mashak Z, Jafariaskari S, Alavi I, Shahreza MS, Dehkordi FS. Phenotypic and genotypic assessment of antibiotic resistance and genotyping of vacA, cagA, iceA, oipA, cagE, and babA2 alleles of Helicobacter pylori bacteria isolated from raw meat. Infection and drug resistance. 2020;13:257. Doi:10.2147/ IDR.S233612.
- 309. Ranjbar R, Yadollahi Farsani F, Safarpoor Dehkordi F. Antimicrobial resistance and genotyping of vacA, cagA, and iceA alleles of the Helicobacter pylori strains isolated from traditional dairy products. Journal of Food Safety. 2019;39(2):e12594. Doi:10.1111/jfs.12594.
- 310. Gilani A, Razavilar V, Rokni N, Rahimi E, editors. VacA and cagA genotypes of Helicobacter pylori isolated from raw meat in Isfahan province, Iran. Veterinary Research Forum; 2017: Faculty of Veterinary Medicine, Urmia University, Urmia, Iran.
- 311. Talimkhani A, Mashak Z. Prevalence and genotyping of helicobacter pylori isolated from meat, Milk and vegetable in Iran. Jundishapur Journal of Microbiology. 2017;10(11). Doi: 10.5812/jjm.14240.
- 312. Hemmatinezhad B, Momtaz H, Rahimi E. VacA, cagA, iceA and oipA genotypes status and antimicrobial resistance properties of Helicobacter pylori isolated from various types of ready to eat foods. Annals of clinical microbiology and antimicrobials. 2016;15(1):1-9. Doi:10.1186/s12941-015-0115-z.
- 313. Saeidi E, Sheikhshahrokh A. VacA genotype status of Helicobacter pylori isolated from foods with animal origin. BioMed research international. 2016;2016. Doi:10.1155/2016/8701067.

- 314. Gilani A, Razavilar V, Rokni N, Rahimi E. VacA and cagA genotypes status and antimicrobial resistance properties of Helicobacter pylori strains isolated from meat products in Isfahan province, Iran. Iranian journal of veterinary research. 2017;18(2):97.
- 315. Ranjbar R, Khamesipour F, Jonaidi Jafari N, Rahimi E. Helicobacter pylori isolated from Iranian drinking water: vacA, cagA, iceA, oipA and babA2 genotype status and antimicrobial resistance properties. FEBS Open Bio. 2016;6(5):433-41. Doi:10.1002/2211-5463.12054.
- 316. Ghorbani F, Gheisari E, Dehkordi FS. Genotyping of vacA alleles of Helicobacter pylori strains recovered from some Iranian food items. Tropical Journal of Pharmaceutical Research. 2016;15(8):1631-6. Doi:10.4314/tjpr.v15i8.5.
- 317. Talaei R, Souod N, Momtaz H, Dabiri H. Milk of livestock as a possible transmission route of Helicobacter pylori infection. Gastroenterology and hepatology from bed to bench. 2015;8(Suppl1):S30.
- 318. Esmaeiligoudarzi D, Tameshkel FS, Ajdarkosh H, Arsalani M, Sohani MH, Behnod V. Prevalence of Helicobacter pyloriinIranian milk and dairy products using culture and ureC based-PCR techniques. Biomedical and Pharmacology Journal. 2015;8(1):179-83. Doi:10.13005/bpj/597.
- 319. Atapoor S, Dehkordi FS, Rahimi E. Detection of Helicobacter pylori in various types of vegetables and salads. Jundishapur Journal of Microbiology. 2014;7(5). Doi: 10.5812/jjm.10013.
- 320. Yahaghi E, Khamesipour F, Mashayekhi F, Safarpoor Dehkordi F, Sakhaei MH, Masoudimanesh M, et al. Helicobacter pylori in vegetables and salads: genotyping and antimicrobial resistance properties. BioMed Research International. 2014;2014. Doi:10.1155/2014/757941.
- 321. Rahimi E, Kheirabadi EK. Detection of Helicobacter pylori in bovine, buffalo, camel, ovine, and caprine milk in Iran. Foodborne Pathogens and Disease. 2012;9(5):453-6. Doi:10.1089/ fpd.2011.1060.
- 322. Ansarian Barezi A, Shakerian A, Rahimi E, Esfandiari Z. Examining the Extent of Contamination, Antibiotic Resistance, and Genetic Diversity of Clostridioides (Clostridium) difficile Strains in Meat and Feces of Some Native Birds of Iran. BioMed Research International. 2023;2023. Doi:10.1155/2023/3524091.
- 323. Ghorbani Filabadi P, Rahimi E, Shakerian A, Esfandiari Z. Prevalence, antibiotic resistance, and genetic diversities of Clostridium difficile in meat nuggets from various sources in Isfahan, Iran. Journal of Food Quality. 2022;2022. Doi:10.1155/2022/9919464.
- 324. Hassani S, Mahmoudi R, Kabudari A, Ghajarbeygi P, Peymani A, Moosavi S, et al. The Prevalence of Clostridium difficile and Clostridium perfringens in Minced and Ground Beef in Iran. Journal of Chemical Health Risks. 2022;12(4):575-80.

- 325. Bacheno H, Ahmadi M, Fazeli F, Ariaii P. Antibiotic Resistance Of Clostridioides Difficile Isolated From Bovine, Vone And Caprine Raw Meat. Journal of Pharmaceutical Negative Results. 2022:9721-6. Doi: 10.47750/pnr.2022.13.S09.1136.
- 326. Hassani S, Pakbin B, Brück WM, Mahmoudi R, Mousavi S. Prevalence, antibiotic resistance, toxin-typing and genotyping of Clostridium perfringens in raw beef meats obtained from Qazvin city, Iran.Antibiotics.2022;11(3):340.Doi:10.3390/an-tibiotics11030340.
- 327. Ram M, Tavassoli M, Ranjbar G, Afshari A. The Microbial and Chemical Quality of Ready-to-Eat Olivier Salad in Mashhad, Iran. Journal of Nutrition, Fasting and Health. 2019;7(4):175-81. Doi: 10.22038/jnfh.2019.40572.1199.
- 328. Hosseinzadeh S, Bahadori M, Poormontaseri M, Dehghani M, Fazeli M, Nazifi S. Molecular characterization of Clostridium perfringens isolated from cattle and sheep carcasses and its antibiotic resistance patterns in Shiraz slaughterhouse, southern Iran. Veterinarski arhiv. 2018;88(5):581-91. Doi:10.24099/vet.arhiv.0009.
- 329. Shahdadnejad N, Mohammadabadi M, Shamsadini M. Typing of clostridium perfringens isolated from broiler chickens using multiplex PCR. Genetics in the 3rd millennium. 2016;14(4):4368-74.
- 330. Doosti A, Pasand M, Mokhtari-Farsani A, Ahmadi R, Chehelgerdi M. Prevalence of Clostridium perfringens type a isolates in different tissues of broiler chickens. Bulgarian Journal of Veterinary Medicine. 2017;20(1). Doi: 10.15547/bjvm.919.
- 331. Hajimohammadi B, Dehghani A, Javadzadeh M, Zandi H, Eslami G. Clostridium botulinum spores and fungal contamination in honeys of Iran. Tolooebehdasht. 2017;15(6):1-9.
- 332. Shakerian A, Rahimi E, Mesbah J, Mousavi M. Molecular Detection of Clostridium Perfringens in Some Raw Animal Food Origin Products in Shahrekord, 2014. Journal of Payavard Salamat. 2017;11(4):391-9.
- 333. Afshari A, Jamshidi A, Razmyar J, Rad M, editors. Genotyping of Clostridium perfringens isolated from broiler meat in northeastern of Iran. Veterinary Research Forum; 2015: Faculty of Veterinary Medicine, Urmia University, Urmia, Iran.
- 334. Afshari A, Jamshidi A, Razmyar J, Rad M. Molecular typing of Clostridium perfringens isolated from minced meat. Iranian Journal of Veterinary Science and Technology. 2015;7(1):32-9. Doi:10.22067/VETERINARY.V711.39040.
- 335. Poormontaseri M, Hosseinzadeh S, Shekarforoush S. Characterization of Clostridium botulinum spores and its toxin in honey. Iranian Journal of Veterinary Research. 2014;15(1):36-9.
- 336. Sadeghi Sarvestani MV, Hosseinzadeh S, Poormontaseri M, Fazeli M. Monitoring the Various Types of Clostridium botulinumin in Four Kinds of Food Stuffs Using Multiplex PCR.

IRANIAN JOURNAL OF VETERINARY SCIENCE AND TECHNOLOGY

Journal of Fasa University of Medical Sciences. 2014;3(4):380-6.

- 337. Shahcheraghi F, Nobari S, Asl HM, Aslani MM. Identification of botulinum toxin type in clinical samples and foods in Iran. Archives of Iranian medicine. 2013;16(11):0-.
- 338. Hosseini H, Tavakoli HR, Meshgi MA, Khaksar R, Hosseini M, Khakpour M. Survey of Clostridium botulinum toxins in Iranian traditional food products. Comparative clinical pathology. 2010;19(3):247-50. Doi:10.1007/s00580-009-0865-0.
- 339. Majzobi MM, Karami P, Khodavirdipour A, Alikhani MY. Brucellosis in humans with the approach of brucella species contamination in unpasteurized milk and dairy products from Hamadan, Iran. Iranian Journal of Medical Microbiology. 2022;16(4):282-7.
- 340. Asgari MH, Ahmadi E. Contamination of bovine milk with Brucella spp.: a current public health menace in Kurdistan province of Iran. Avicenna Journal of Clinical Microbiology and Infection. 2021;8(1):17-22. Doi: 10.34172/ajcmi.2021.04.
- 341. Majzoobi MM, Karami P, Khodavirdipour A, yousef Alikhani M. The contamination rate of Brucella spp. in unpasteurized dairy products used in urban and rural areas of Hamadan County, west of Iran. 2020. Doi:10.21203/rs.2.22703/v1.
- 342. Alamian S, Dadar M. Brucella abortus contamination of camel milk in two Iranian regions. Preventive veterinary medicine. 2019;169:104708. Doi:10.1016/j.prevetmed.2019.104708.
- 343. Mirnejad R, Jazi FM, Mostafaei S, Sedighi M. Molecular investigation of virulence factors of Brucella melitensis and Brucella abortus strains isolated from clinical and non-clinical samples. Microbial pathogenesis. 2017;109:8-14. Doi:10.1016/j. micpath.2017.05.019.
- 344. Shirazi Z, Khalili M, Sadeghi B, Sharifi H, Ashrafganjooyi S. Detection of Brucella spp. in the sheep and goats milks from southeastern Iran using culture and PCR. Journal of Medical Microbiology and Infectious Diseases. 2017;5(3):40-2.
- 345. Ashrafganjooyi S, Saedadeli N, Alamian S, Khalili M, Shirazi Z. Isolation and biotyping of Brucella spp. from sheep and goats raw milk in southeastern Iran. Trop Biomed. 2017;34(3):507-11.
- 346. Majid Y, Somayeh N, Parisa S, Behrooz A, Reza FN, Javad R, et al. Prevalence of Brucella melitensis and Brucella abortus in raw milk and dairy product by real time PCR technique. The Ulutas Medical Journal. 2016;2(1):7-11. Doi:10.5455/ umj.20151004054235.
- 347.Khalili M, Aflatoonian MR, Aliabadi FS, Abshenas J. Brucella contamination in raw milk by polymerase chain reaction. Tehran University Medical Journal TUMS Publications. 2016;74(7):517-21.

- 348. Shakerian A, Deo P, Rahimi E, Shahjavan A-R, Khamesipour F. Molecular detection of Brucella melitensis in sheep and goat milk in Iran. Tropical Journal of Pharmaceutical Research. 2016;15(5):913-8. Doi:10.4314/tjpr.v15i5.3.
- 349. Izadi A, Moslemi E, Tabatabaei Panah A, Kheiri Manjili H. Brucella spp. detection in dairy products using nested and hemi nested PCR techniques. Ann Biol Res. 2014;5(1):124-31.
- 350. Gholizadeh SS, Zali M, Hashempour A, AHMADI EM. Investigation of brucellosis in cattle and sheep in Urmia-Iran. Yüzüncü yıl Üniversitesi Veteriner Fakültesi Dergisi. 2013;24(3):133-4.
- 351. Karimi Alavigeh A, Sharifzadeh A. Study on the Frequency of Vibrio species in fish from Isfahan. Journal of Food Microbiology. 2021;8(4):47-55.
- 352. Shohreh P, Azizkhani M, Tooryan F. Food Poisoning Bacteria from Commercially Important Fish Species Collected from Markets in Mazandaran Province. Fisheries Science and Technology. 2019;8(1):1-6.
- 353. Asgarpoor D, Haghi F, Zeighami H. Detection and molecular characterization of Vibrio parahaemolyticus in shrimp samples. The Open Biotechnology Journal. 2018;12(1).
- 354. Mashak Z, Khalajzadeh A, Koohdar V. Study the bacterial and fungal quality and physicochemical properties of cold smoked salted fishes of Caspian sea, İran. Biosciences Biotechnology Research Asia. 2016;13(3):1811-20. Doi: 10.13005/bbra/2334.
- 355. Raissy M, Rahimi E, Azargun R, Moumeni M, Sohrabi H. Molecular detection of Vibrio spp. in fish and shrimp from the persian gulf. Journal of Food Biosciences and Technology. 2015;5(2):49-52.
- 356. Hosseini S, Safarpoor Dehkordi F, Rahimi E, Shakerian A. Prevalence study of Vibrio species and frequency of the virulence genes of Vibrio parahaemolyticus isolated from fresh and salted shrimps in Genaveh seaport. Food Hygiene. 2014;4(2 (14)):17-26.
- 357. Khamesipour F, Noshadi E, Moradi M, Raissy M. Detection of Vibrio spp. in shrimp from aquaculture sites in Iran using polymerase chain reaction (PCR). Aquaculture, Aquarium, Conservation & Legislation. 2014;7(1):1-7.
- 358. Dehkordi FS, Hosseini S, Rahimi E, Momeni M, Yahaghi E, Darian EK. Study the distribution of virulence genes of Vibrio parahaemolyticus isolated from fish, lobster and crab caught from Persian Gulf. Iran J Med Microbiol: Volume. 2014;8(2).
- 359. Raissy M, Khamesipour F, Rahimi E, Khodadoostan A. Occurrence of Vibrio spp., Aeromonas hydrophila, Escherichia coli and Campylobacter spp. in crayfish (Astacus leptodactylus) from Iran. Iranian Journal of Fisheries Sciences. 2014;13(4):944-54.

- 360. Momtaz H, Dehkordi FS, Rahimi E, Asgarifar A. Detection of Escherichia coli, Salmonella species, and Vibrio cholerae in tap water and bottled drinking water in Isfahan, Iran. BMC public health. 2013;13(1):1-7. Doi:10.1186/1471-2458-13-556.
- 361. Raissy M, Moumeni M, Ansari M, Rahimi E. Occurrence of Vibrio spp. in lobster and crab from the Persian Gulf. Journal of Food Safety. 2012;32(2):198-203. Doi:10.1111/j.1745-4565.2012.00368.x.
- 362. Rahimi E, Ameri M, Doosti A, Gholampour AR. Occurrence of toxigenic Vibrio parahaemolyticus strains in shrimp in Iran. Foodborne pathogens and disease. 2010;7(9):1107-11. Doi:10.1089/fpd.2010.0554.
- 363. Hosseini H, Cheraghali AM, Yalfani R, Razavilar V. Incidence of Vibrio spp. in shrimp caught off the south coast of Iran. Food control. 2004;15(3):187-90. Doi:10.1016/S0956-7135(03)00045-8.
- 364. Pakbin B, Didban A, Brück WM, Alizadeh M. Phylogenetic analysis and antibiotic resistance of Shigella sonnei isolates. FEMS microbiology letters. 2022;369(1):fnac042. Doi:10.1093/femsle/fnac042.
- 365. Pakbin B, Amani Z, Allahyari S, Mousavi S, Mahmoudi R, Brück WM, et al. Genetic diversity and antibiotic resistance of Shigella spp. isolates from food products. Food Science & Nutrition. 2021;9(11):6362-71. Doi:10.1002/fsn3.2603.
- 366. Pakbin B, Didban A, Monfared YK, Mahmoudi R, Peymani A, Modabber MR. Antibiotic susceptibility and genetic relatedness of Shigella species isolated from food and human stool samples in Qazvin, Iran. BMC research notes. 2021;14(1):1-6. Doi:10.1186/s13104-021-05554-3.
- 367. Tehrani ST, Harzandi N, Jabalameli L. Molecular detection of Shigella spp. contamination in ready-to-eat salad samples in west of Tehran. International Journal of Enteric Pathogens. 2018;6(2):41-4. Doi: 10.15171/ijep.2018.11.
- 368. Bahador N, Ajampour I. Phenotypic and genotypic characterization of isolated Shigella spp. from food based on IpaH gene and evaluation of their antibiotic patterns. Journal of Food Microbiology. 2019;5(4):55-65.
- 369. Yam BAZ, Khomeiri M, Mahounak AS, Jafari SM. Hygienic quality of camel milk and fermented camel milk (chal) in Golestan Province, Iran. Journal of Microbiology Research. 2014;4(2):98-103. Doi: 10.5923/j.microbiology.20140402.09.

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