



Intra-Hippocampal Injection of Abscisic Acid Attenuates Learning and Memory Deficits, and Changes Oxidative Stress Indices in REM Sleep Deprived Rats

Alireza Fekrat^a, Mahzad Abbasnejad^b, Razieh Kooshki^c, Parastoo Nikkhrouy^b, Mehdi Abbasnejad^b

^a Department of Psychiatry, Nourah hospital, Kerman University of Medical Sciences, Kerman, Iran.

^b Department of Biology, Faculty of Sciences, Shahid Bahonar University of Kerman, Kerman, Iran.

^c Department of Biology, Faculty of Sciences, Lorestan University, Khorramabad, Iran.

ABSTRACT

This study evaluated whether intra-hippocampal administration of ABA can modulate learning and memory performance and oxidative stress biomarker activities in the cerebral cortex of rats exposed to rapid eye movement (REM) sleep deprivation. Adult male Wistar rats were cannulated in the CA1 area of the hippocampus. After recovery, the rats were subjected to REM sleep deprivation for 4 days. Then, the groups of REM sleep-deprived (SD) rats were treated with ABA (5, 10, and 15 µg) and ABA (10 µg) + bicuculline (Bic), a competitive GABAA receptor antagonist. Memory and learning were evaluated with the Morris water maze (MWM) and shuttle box tests. Moreover, alterations in catalase levels as an antioxidant enzyme, MDA, and H₂O₂ as oxidant biomarkers were determined in rat brain cortex. REM SD rats indicated noteworthy learning and memory deficits in both the MWM and shuttle box tests when compared to control rats. However, intra-CA1 injection of ABA (10 µg) decreased cognitive impairment in REM SD rats. Bic (1 µg/rat) could not change ABA (10 µg) effects. In addition, an increase in catalase activity and a decrease in MDA and H₂O₂ were indicated in the brain cortex of ABA (10 µg) and ABA+ Bic treated groups. Overall, the data showed ABA's aptitude to attenuate REM sleep deprivation-induced learning and memory disruption and oxidative damage in rats. Manipulation of the GABAA receptor failed to inhibit ABA effects in REM SD rats.

Keywords

sleep deprivation , abscisic acid , bicuculline, learning and memory , Rat

Number of Figures: 4
Number of Tables: 0
Number of References: 45
Number of Pages: 9

Abbreviations

CAT: Catalase
REM: Rapid eye movement
STZ: Streptozotocin
Bic: Bicuculline

MWM: Morris water maze
MDA: Malondialdehyde
H₂O₂: Hydrogen peroxide
SD: Sleep deprivation

Introduction

Sleep deprivation (SD) is a condition of inadequate sleep that can be considered a physiological disorder or a result of an inappropriate lifestyle [1, 2]. Sleep quality has a significant impact on the regulation of other physiological processes, including learning and memory [3, 4]. It has been shown that SD disrupts memory retrieval and consolidation by changing hippocampus structural constancy [5]. The patterns of rhythmic brain waves in non-rapid eye movement sleep also show a relationship with hippocampal activities [6]. Hippocampal-mediated learning and memory, as well as neurotransmitters, are affected by sleep quality [7, 8]. REM sleep deprivation could decline motor and sensory learning experiences in animals [9, 10].

Abscisic acid (ABA) is produced in all parts of plants and plays notable roles in their physiological functions, especially the regulation of stress responses [11, 12]. ABA is synthesized from pro-vitamin A carotenoids [13], which are found in high concentrations in plants [12]. Moreover, in animals, ABA is found in various brain areas including the hippocampus, cerebral cortex, and cerebellum [14, 15]. ABA receptors are peroxisome proliferator-activated receptors (PPARs) and lanthionine synthetase C-like protein 2 [16, 17]. ABA signalling shows variation, but changes in calcium concentration and activation of cyclic ADP-ribose are the most mutual pathways [18-20].

ABA exerts modulatory effects on a variety of physiological functions including nociception, anxiety and depression like behavior, sleep and learning and memory performances in rats [15, 21]. Central administration of ABA exhibited analgesic effect which is facilitated by the PPAR β/δ and opioid signalling [22]. Moreover, ABA meaningfully improved the pentobarbital-related sub hypnotic effects and also endorsed sleep induction. Such effects showed dependency with GABAA receptors and PPAR β /PPAR γ signalling [23].

The main goal of the present study was to evaluate if intra-hippocampal treatment of ABA can alter learning and memory performance in rats exposed to REM-SD. Moreover, bicuculline was used to assess the possible association of ABA with the GABA A receptor. In a previous study, pretreatment with bicuculline was found to block ABA's ability to extend sleep duration in a rat model of pentobarbital-induced sleep.

Abbreviations-Cont'd

PPARs: Peroxisome proliferator-activated receptors

ABA: Abscisic acid

ROS: Reactive oxygen species

CGRP: Calcitonin gene-related peptide

Bicuculline is a competitive GABAA receptor antagonist that blocks GABA's inhibitory effects by preventing chloride ion influx, leading to increased neuronal excitability and potential seizure activity[24]. The alteration of pro-oxidant/antioxidant biomarkers was also assessed in the cerebral cortex of SD rats.

Results

PA test

The SD group showed an increase in the number of acquisition trials when compared with the control group ($p < 0.001$) (Fig. 1A). However, the number of acquisition trials was significantly decreased in SD groups post-treated with ABA (10 μg and 15 μg) ($p < 0.001$). No major alteration in acquisition trials was found in SD rats post-treated with Bic + ABA (10 μg) as compared to SD+ABA (10 μg) group. In addition, an increase in the step-through latency and a decline in time spent in the dark cavity were determined in the SD group ($p < .001$). ABA (10 μg) was able to increase the step-through latency and decrease time spent in a dark chamber in SD rats ($p < 0.001$). In addition, no significant alteration was found in the SD rats' response infused with Bic + ABA (10 μg) as compared with the ABA (10 μg) group (Fig. 1B and 1C).

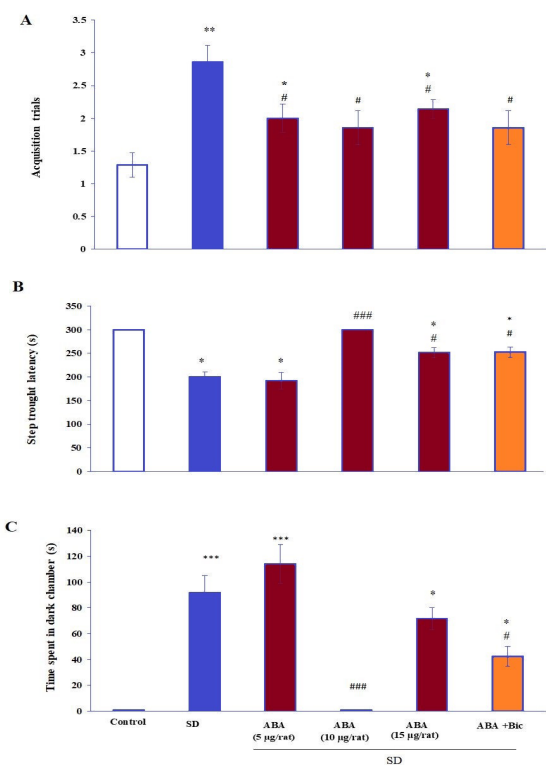
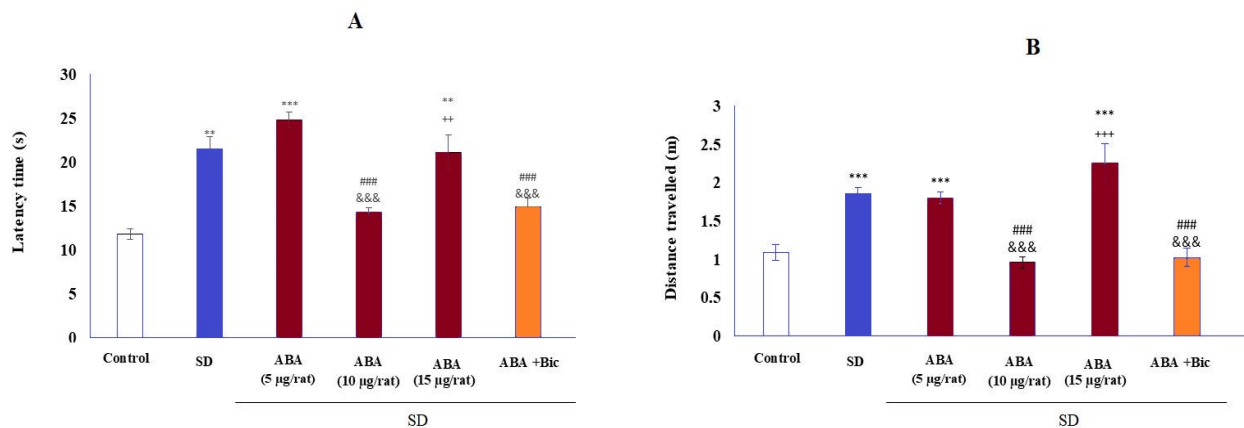


Figure 1.

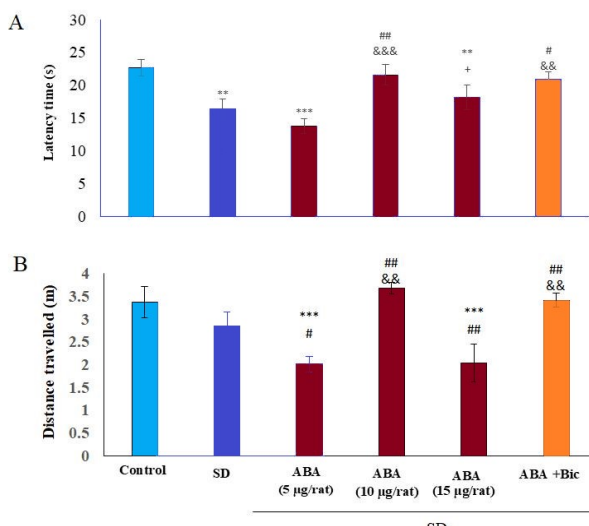
The effect of intra-hippocampal administration of ABA (5, 10, and 15 $\mu\text{g}/\text{rat}$) or Bic+ABA (10 $\mu\text{g}/\text{rat}$) on the number of acquisition trials (A), step through latency (B), and time spent in a dark chamber (C) in passive avoidance test in SD rats. Values are expressed as mean \pm SEM. * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$ versus control groups, # $p < 0.05$, ## $p < 0.01$, and ### $p < 0.001$ versus SD group

MWM test

In acquisition trials, the latency time to catch the concealed platform was pointedly increased in the SD group in comparison to the control group ($p < 0.001$). Intra-hippocampal infusion of ABA (10 μg / rat) expressively decreased the latency time to catch the concealed stage in the SD rats ($p < 0.01$) (Fig. 2A). Moreover, SD+Bic+ABA (10 μg /rat) and SD+ ABA (10 μg /rat) treated groups show no change the latency to discover the hidden platform. Moreover, the groups showed major differences in space moved to touch the concealed platform on the acquisition test. As shown in Fig. 2B, the distance trekked to touch the hidden stage was meaningfully increased in the SD group ($p < 0.001$).

**Figure 2.**

The effect of intra-hippocampal administration of ABA (5, 10, and 15 $\mu\text{g}/\text{rat}$) or Bic+ABA (10 $\mu\text{g}/\text{rat}$) on the escape latency time (A) and distance travelled to find the hidden platform in the MWM test in SD rats. Values are expressed as mean \pm SEM. ** $p < 0.01$ and *** $p < 0.001$ versus control groups, ### $p < 0.001$ versus SD group, &&& $p < 0.001$ versus SD + ABA (5 $\mu\text{g}/\text{rat}$) group, ++ $p < 0.01$, +++ $p < 0.001$ versus SD + ABA (10 $\mu\text{g}/\text{rat}$) group

**Figure 3**

The effect of intra-hippocampal administration of ABA (5, 10, and 15 $\mu\text{g}/\text{rat}$) or Bic+ABA (10 $\mu\text{g}/\text{rat}$) on the duration time (A), and distance travelled in target zone in SD rats in probe trial of MWM test. Values are expressed as mean \pm SEM. ** $p < 0.01$ and *** $p < 0.001$ versus control groups, # $p < 0.05$, ## $p < 0.01$ versus SD group, && $p < 0.05$, &&& $p < 0.001$ versus SD + ABA (5 $\mu\text{g}/\text{rat}$) group, + $p < 0.05$ versus SD + ABA (10 $\mu\text{g}/\text{rat}$) group

$p < 0.001$). Besides, the SD group treated with ABA (10 $\mu\text{g}/\text{rat}$) traveled a lower distance to reach the hidden platform as compared to the SD group ($p < 0.001$). In the SD group injected with Bic+ ABA (10 $\mu\text{g}/\text{rat}$) the distance traveled to find the platform showed no difference as compared to the ABA (10 $\mu\text{g}/\text{rat}$) group (Fig. 2B).

Fig.3 indicates the results of the probe trial. The figure indicated that time spent and the traveled distance in the object zone significantly decreased in the SD group than the control group ($p < 0.001$) (Fig. 3A). Moreover, ABA weakened the effects of SD on the time spent in the object area ($p < 0.05$) (Fig. 3A). Further, ABA meaningfully improved distance traveled in the

object area in SD-treated rats ($p < 0.01$) (Fig. 3B). As notated in Fig. 3, SD rats infused with Bic + ABA show no significant difference in spent time and distance traveled in the target quadrat in comparison to ABA (10 $\mu\text{g}/\text{rat}$) group ($p < 0.05$).

Biochemical assay

The activity of the antioxidant enzyme CAT was significantly decreased in the SD group when compared with control rats. As shown in Fig. 4A, ABA at 10 $\mu\text{g}/\text{rat}$ and Bic+ABA (10 $\mu\text{g}/\text{rat}$) were able to increase CAT activity in the SD group. Moreover, there were significant increases in the activity of pro-oxidant biomarker H₂O₂ and MDA concentration in the cerebral cortex of the SD group as compared to the control group. However, post-treatment of SD rats with ABA (10 $\mu\text{g}/\text{rat}$) or Bic+ABA significantly attenuated H₂O₂ activity and MDA level in the cerebral cortex (Fig. 4B and 4C).

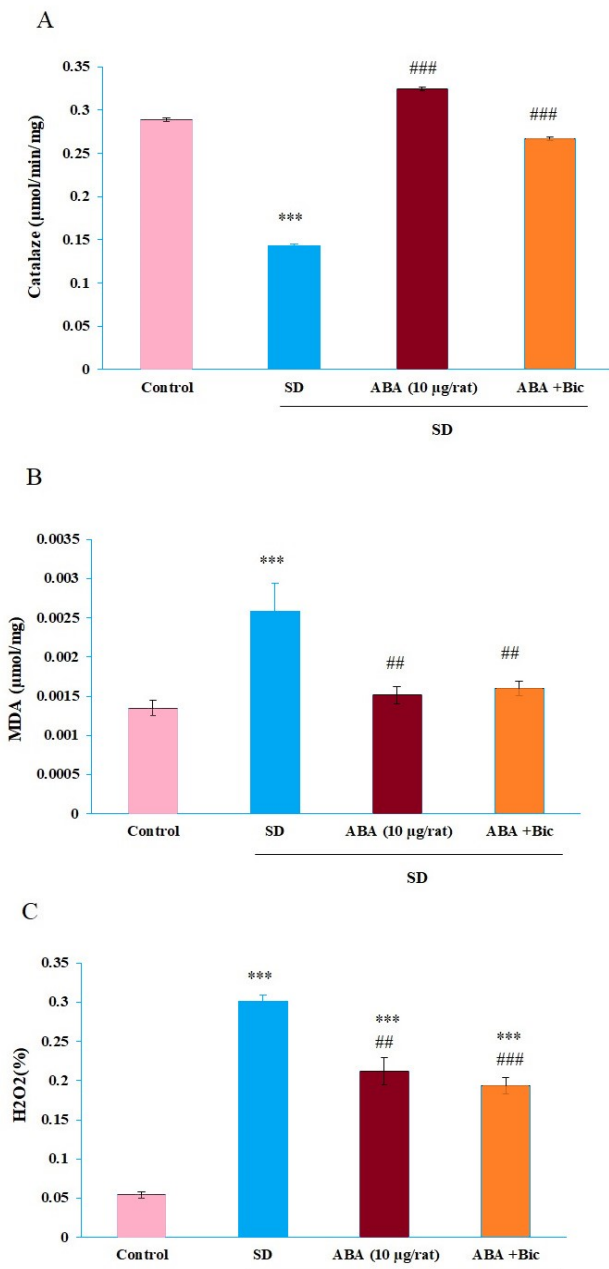


Figure 4.

The effect of intra-hippocampal administration of ABA (10 µg/rat) or Bic+ABA (10 µg/rat) on the activity of CAT enzyme (A), MDA concentration (B) and H₂O₂ activity in the cerebral cortex of rats. Values are expressed as mean ± SEM. ** $p < 0.01$ and *** $p < 0.001$ versus control groups, # $p < 0.05$, ## $p < 0.01$ versus SD group, && $p < 0.05$, &&& $p < 0.001$ versus SD + ABA (5 µg/rat) group, + $p < 0.05$ versus SD + ABA (10 µg/rat) group

Discussion

The present study showed the deteriorating effects of REM sleep deprivation on the memory and learning performance of rats assessed in the MWM and shuttle box tests. However, intra-CA1 microinjection of ABA decreased SD-induced learning and memory deficiency in rats. Moreover, the sleep-deprived rats

indicated a disruption in oxidant/antioxidant biomarkers verified by a decrease in CAT activity and increases in lipid peroxidation and H₂O₂ production in the cerebral cortex, which was prevented by ABA (10 µg) treatment. The ABA effects in behavioral and biochemical experiments did not diminish with the GABA receptors antagonist bicuculline.

The importance of sleep quality on cognitive performance, especially hippocampal-dependent learning and memory has been strongly supported by evidence from clinical and experimental studies [25]. In this study, the rats' learning and memory performances were assessed after a continuous 72 h period of REM SD. The 72-hour REM sleep deprivation period in rats reflects severe sleep loss but is not directly equivalent to 72 hours in humans due to differences in metabolism and sleep architecture. In humans, this time-frame would likely correspond to several days of significant sleep restriction or chronic sleep disruption rather than total sleep deprivation. Rodent models typically involve more intense and compressed sleep deprivation protocols compared to human studies. To bridge the gap between rodent and human studies, future research could explore the effects of varying durations of REM sleep deprivation in animal models and attempt to correlate these findings with human studies involving partial sleep restriction or chronic sleep fragmentation. Learning and memory changes following SD are highly dependent on the lasting duration of the SD. In line with our result, most studies showed the highest detrimental effects of SD on memory performance when it lasted for 72 h. Nevertheless, in some cases, shorter terms of SD lasting for 24 or 48 hours have been associated with no alteration or even increases in hippocampal synaptic plasticity and memory impairment [26–28]. The mechanism(s) underlying different effects of SD lasting on learning and memory function are complex and still not well understood.

For the first time, this study shows ABA's ability to increase learning and memory performance in SD rats. The efficacy of ABA interventions on sleep, learning, and memory has been demonstrated in previous studies conducted on rodents. It has been indicated that ABA decreases onset time and prolongs sleep duration in a rat model of pentobarbital-induced sleep [23]. Moreover, ABA treatment reduced learning and memory deficits in rat models of STZ-induced Alzheimer's disease [29]. In addition, ABA infusion decreased learning and memory deficits in MWM and shuttle box tasks in STZ diabetic rats [30]. The mechanism(s) of ABA involvement to attenuate sleep deprivation weakening effects on learning and memory is not understood. It is postulated the effects might be intended by manipulation of related neurotransmit-

ters and distinct neural networks within the brain.

The data showed pharmacological blockage of the GABAA receptor with bicuculline did not inhibit ABA efficiency on learning and memory performances in sleep-deprived rats. In a related study, pretreatment with bicuculline could obstruct ABA impending to prolong sleep duration in a rat model of pentobarbital-induced sleep [23]. This duality suggests that ABA could engage different pathways—supporting both sleep recovery and neurocognitive resilience—depending on the physiological or experimental conditions.

GABAergic synapses are profoundly founded on hippocampus CA1 pyramidal neurons [31, 32]. While the baseline GABA levels in the hippocampus improve learning and memory performance, an increase in GABAA receptor activity has been shown to decline network excitability and reduce synaptic plasticity in the CA1 area [33, 34]. Indeed, memory retrieval is impeded by the glutamate and GABA concentration balance in the brain [35]. In the rats subjected to SD impairment of memory performance has been associated with imbalances in Glu/GABA ratio [36]. Although this study did not find ABA interfering with the GABAA receptor, however, more data are still required to describe the details of ABA's impact on the GABAergic system to modulate the learning and memory of SD-exposed rats.

In the present study, REM sleep deprivation increased oxidative stress damage defined by increases in lipid peroxidation and H₂O₂ levels, and a decrease in CAT activity in the cerebral cortex of rats. However, post-treatment with ABA (10 µg/rat), which was the most effective dose to increase learning and memory behaviors, could inhibit oxidative stress imbalances in SD rats. This data is supported by many previous studies that display ABA antioxidant capacity in rodents. Oral treatment with ABA in drinking water increased antioxidant defence systems indices and decreased MDA levels in many tissues of rats [37]. Moreover, intra-lateral ventricles infusion of ABA increased feeding behavior and increased the antioxidant enzymes activity, while attenuated stress oxidative enzymes [38]. In a mouse model of thioacetamide-induced hepatic fibrosis ABA treatment decreased oxidative stress enlargements and inflammation by induction of NF-κB signaling path [39]. Indeed, this study data support an association between ABA antioxidant properties and reduction of REM-SD induced learning and memory deficits.

It has been shown that as a isoprenoid plant hormone compound, ABA binds to PPARs and activates several intracellular signaling molecules essential in the regulation of learning and memory performance [16]. Pretreatment with PPAR β/δ antagonist was able

to suppress ABA anti-nociceptive effects in rats [16]. Moreover, ABA decreased diabetes-induced learning and memory deficit in rats via intonation of PPARγ receptors [30]. In addition, PPARγ receptors antagonist prevented the ability of ABA to increase sleep duration in a rat model of pentobarbital-induced sleep [23]. On the other hand, motivation of PPARγ receptors with ABA modifies calcium channel activity and induces PI3K/PKC pathway in rat's brain to modulate learning and memory and anxiety-like behavior [40]. Possibly ABA efficiency on learning and memory responses in SD rats is at least partially mediated by manipulations of the PPARs system and induction of the downstream signaling molecules involved in learning and memory performance.

Our study primarily focused on learning and memory performance using specific behavioral tests (e.g., acquisition trials). While these tests provide valuable insights, they may not fully capture the broader spectrum of cognitive functions affected by sleep deprivation or ABA treatment. While oxidative stress biomarkers (catalase, MDA, H₂O₂) were evaluated, other potential mechanisms (e.g., neuroinflammation, synaptic plasticity) were not explored, leaving gaps in understanding ABA's comprehensive effects.

Conclusions

Overall, the data of this study showed the potential of intra-hippocampal administration of ABA to increase antioxidant indices in the brain and attenuate learning and memory deficits in RAM-SD rats. Pretreatment infusion with GABAA receptors antagonist did not change ABA-induced responses.

Materials & Methods

Animals

Adult male Wistar rats (2 months) weighing 230–270 grams were used in this study. The animals were contained four per cage in a room with a temperature of 23 ± 2 °C under a 12-h light/dark cycle with limitless entrance to food and water. All trial procedures were permitted by the Animal Research Ethics Committee of Kerman University of Medical Sciences, Kerman, Iran.

Surgery and microinjection

Rats were profoundly anesthetized with a mixture of ketamine (100 mg) and xylazine (5 mg) and placed in a stereotaxic apparatus (Estotling CO, USA). Guide cannulae were bilaterally inserted in the CA1 region (3.8 mm posterior to the bregma, 2.2 mm lateral from the midline, and 3.2 mm depth to the cortical surface). Afterward, rats were kept separately and endorsed for 1 week to recover from surgery before treatments [41]. The drugs (1 µL each side) were delivered using a 27-gauge stainless steel needle devoted to a Hamilton micro-syringe.

Experimental design

The animals were randomly separated into six experimental groups

(n=7) as follows: control (untreated rats); sleep deprivation (SD): located in small platform; SD + ABA groups: treated intra-CA1 with ABA (5, 10, and 15 µg/ rat) and then located on small platform; SD + ABA (10 µg/ rat) + bicuculline (1 µg/rat): treated intra-CA1 with ABA and bicuculline and then located on small platform. The groups were exposed to SD procedures for 72 h and then injected with specific treatments. Ten minutes after intra-hippocampal injection, the rats were verified in MWM and Shuttle box tests, respectively.

Sleep deprivation (SD)

In the first tests, the single small platform method of SD was used. Animals were sited on a single stage in the center of a water cistern. The water reached up to 2 cm under the shallow of the stage. Based on the multiple small platform method, five stages (each 5 cm diameter) were used. In this method, the stages were spread out (8–10 cm apart) so that animals were able to simply travel amongst them but could not lie through any two. The control group was tested using the single large stage method, where the size of the stage was enlarged to 15.2 cm to ensure sleep. All the treatments lasted 72 hours [42].

Learning and memory assessment

1. MWM

In this study, all the experimental groups were subjected to four days of training trials in the MWM, as defined previously [43]. A video camera was attached straight overhead the water maze pool, and the tracking system of Any maze was provided to assess the time to reach the concealed platform (the escape latency) and the length of the swim (traveled distance) of each rat in training time. Twenty-four hours later, the rats were evaluated in the probe trial, in which the escape platform was detached from the pool, and the animal was permitted to swim for 60 sec. The total time spent and the number of visits across the past position of the platform were measured to appraise spatial memory.

2. Shuttle box test

The apparatus encompassed identical-sized light and dark partitions that were separated by a sliding guillotine door. The floor of the dark and light partitions consisted of a stainless-steel shock grid. This test was divided into training and memory stages. In the instruction phase, each animal was positioned in the lightened partition, and after 5 seconds, the gate was unlocked and the rats were indorsed to transfer freely into the dark space. Upon entry into the dark chamber, the door was barred and the rat was assumed 1 mA electrical shock in 1 second. The instruction trial was completed when the rat endured in the light hall for 5 continuous min. Twenty-four hours later, in stage 2 (retrieval session), each rat was positioned on the light side of the box. Ensuing 30 s acclimatization, the door was raised. The number of electrical shock trial, latency to enter the dark chamber initial time spent to wholly enter the dark room (STL), as well as whole time consumed in the dark box were important in the passive avoidance test [44].

Biochemical assay

The rats were euthanized with CO₂. The brains were detached, and the separated brain regions, hippocampus and prefrontal lobe, were kept in liquid nitrogen for assessment of biochemical parameters. Brain malondialdehyde (MDA) and hydrogen peroxide (H₂O₂) assay were evaluated as lipid peroxidation products and oxidative stress index [26, 45]. Moreover, the measurement of catalase enzyme activity was done as an index of antioxidant activity [46].

Statistical analysis

The results are expressed as the mean ± SEM. The statistical analyses were performed using SPSS (version 22) software. One-way analysis of variance (ANOVA) was used to evaluate significant variations among groups. Tukey's Post hoc assessment was conducted to ex-

plore the differences between the groups. A significance level of $p < 0.05$ was adopted for all tests.

Authors' Contributions

Md. Taimur Islam conceived the idea, designed the experiments and drafted the first version of the manuscript; Mohosina Mou, Nusrat Binte Rafique, Minhaz Ahmed and Md. Selim Jahangir Saurov performed the sample collection and laboratory experiment; Robius Sani Sadi edited the manuscript; Anup Kumar Talukder participated in the data analysis and edited the manuscript; Ziban Chandra Das and Md. Golam Haider reviewed the manuscript. All authors read and approved the final version of the manuscript.

Acknowledgements

This study was granted by Shahid Bahonar University of Kerman (grant number: 99.7).

Conflict of interest

The authors have no competing interests to declare.

References

- Gong L, Liu Q. Mobile phone addiction and sleep quality: the mediating role of anxiety and the moderating role of emotion regulation. *Behav Sci.* 2023;13(3):250. Doi: 0.3390/bs13030250.
- Chattu VK, Manzar MD, Kumary S, Burman D, Spence DW, Pandi-Perumal SR. The global problem of insufficient sleep and its serious public health implications. *Healthcare*; 2018: MDPI. Doi: 10.3390/healthcare7010001.
- Tobaldini E, Costantino G, Solbiati M, Cogliati C, Kara T, Nobili L, et al. Sleep, sleep deprivation, autonomic nervous system and cardiovascular diseases. *Neurosci Biobehav Rev.* 2017;74:321-9. Doi: 10.1016/j.neubiorev.2016.07.004.
- Shen Y, Lv Q-k, Xie W-y, Gong S-y, Zhuang S, Liu J-y, et al. Circadian disruption and sleep disorders in neurodegeneration. *Trans Neurodegeneration.* 2023;12(1):8. Doi: 10.1186/s40035-023-00340-6.
- Guan Z, Peng X, Fang J. Sleep deprivation impairs spatial memory and decreases extracellular signal-regulated kinase phosphorylation in the hippocampus. *Brain Res.* 2004;1018(1):38-47. Doi: 10.1016/j.brainres.2004.05.032.
- Latchoumane C-F V, Ngo H-V V, Born J, Shin H-S. Thalamic spindles promote memory formation during sleep through triple phase-locking of cortical, thalamic, and hippocampal rhythms. *Neuron.* <http://dx.doi.org/10.1016/j.neuron.2017.06.025> (2017). Doi: 10.1016/j.neuron.2017.06.025.

7. Abel T, Havekes R, Saletin JM, Walker MP. Sleep, plasticity and memory from molecules to whole-brain networks. *Curr Biol*. 2013;23(17):R774-R88. Doi: 10.1016/j.cub.2013.07.025.
8. Havekes R, Canton DA, Park AJ, Huang T, Nie T, Day JP, et al. Gravin orchestrates protein kinase A and β 2-adrenergic receptor signaling critical for synaptic plasticity and memory. *J Neurosci*. 2012;32(50):18137-49. Doi: 10.1523/JNEUROSCI.3612-12.2012.
9. Henderson MX, Wirak GS, Zhang Y-q, Dai F, Ginsberg SD, Dolzhanskaya N, et al. Neuronal ceroid lipofuscinosis with DNAJC5/CSP α mutation has PPT1 pathology and exhibit aberrant protein palmitoylation. *Acta Neuropathologica*. 2016;131(4):621-37. Doi: 10.1007/s00401-015-1512-2.
10. Rolls A, Colas D, Adamantidis A, Carter M, Lanre-Amos T, Heller HC, et al. Optogenetic disruption of sleep continuity impairs memory consolidation. *Proc Natl Academy Sci*. 2011;108(32):13305-10. Doi: 10.1073/pnas.101563108.
11. Matthys D, Gielis J, Debergh P. Ethylene. Automation and environmental control in plant tissue culture. Springer; 1995. p. 473-91.
12. Magnone M, Ameri P, Salis A, Andraghetti G, Emionite L, Murialdo G, et al. Microgram amounts of abscisic acid in fruit extracts improve glucose tolerance and reduce insulinemia in rats and in humans. *FASEB J*. 2015;29(12):4783-93. Doi: 10.1096/fj.15-277731.
13. Finkelstein RR, Gampala SS, Rock CD. Abscisic acid signaling in seeds and seedlings. *Plant Cell*. 2002;14(suppl 1):S15-S45. Doi: 10.1105/tpc.010441.
14. Qi C-C, Zhang Z, Fang H, Liu J, Zhou N, Ge J-F, et al. Antidepressant effects of abscisic acid mediated by the downregulation of corticotrophin-releasing hormone gene expression in rats. *Int J Neuropsychopharmacol*. 2015;18(4). Doi: 10.1093/ijnp/pyu006.
15. Qi C-C, Ge J-F, Zhou J-N. Preliminary evidence that abscisic acid improves spatial memory in rats. *Phys Behav*. 2015;139:231-9. Doi: 10.1016/j.physbeh.2014.11.053.
16. Bassaganya-Riera J, Guri AJ, Lu P, Climent M, Carbo A, Sobral BW, et al. Abscisic acid regulates inflammation via ligand-binding domain-independent activation of peroxisome proliferator-activated receptor γ . *J Biol Chem*. 2011;286(4):2504-16. Doi: 10.1074/jbc.M110.160077.
17. Sturla L, Fresia C, Guida L, Bruzzone S, Scarfi S, Usai C, et al. LANCL2 is necessary for abscisic acid binding and signaling in human granulocytes and in rat insulinoma cells. *J Biol Chem*. 2009;284(41):28045-57. Doi: 10.1074/jbc.M109.035329.
18. Magnone M, Bruzzone S, Guida L, Damonte G, Millo E, Scarfi S, et al. Abscisic acid released by human monocytes activates monocytes and vascular smooth muscle cell responses involved in atherogenesis. *J Biol Chem*. 2009;284(26):17808-18. Doi: 10.1074/jbc.M809546200.
19. Zocchi E, Basile G, Cerrano C, Bavestrello G, Giovine M, Bruzzone S, et al. ABA-and cADPR-mediated effects on respiration and filtration downstream of the temperature-signaling cascade in sponges. *J Cell Sci*. 2003;116(4):629-36. Doi: 10.1242/jcs.00277.
20. Scarfi S, Ferraris C, Fruscione F, Fresia C, Guida L, Bruzzone S, et al. Cyclic ADP-ribose-mediated expansion and stimulation of human mesenchymal stem cells by the plant hormone abscisic acid. *Stem Cells*. 2008;26(11):2855-64. Doi: 10.1634/stemcells.2008-0488.
21. Freitag CM, Miller RJ. Peroxisome proliferator-activated receptor agonists modulate neuropathic pain: a link to chemokines? *Front Cell Neurosci*. 2014;8:238. Doi: 10.3389/fncel.2014.00238.
22. Mollashahi M, Abbasnejad M, Esmaeili-Mahani S. Phytohormone abscisic acid elicits antinociceptive effects in rats through the activation of opioid and peroxisome proliferator-activated receptors β/δ . *Eur J Pharmacol*. 2018;832:75-80. Doi: 10.1016/j.ejphar.2018.05.013.
23. Madadzadeh M, Abbasnejad M, Mollashahi M, Pourrahimi AM, Esmaeili-Mahani S. Phytohormone abscisic acid boosts pentobarbital-induced sleep through activation of GABA-A, PPAR β and PPAR γ receptor signaling. *Arq de Neuro-Psiquiatria*. 2021;79:216-21. Doi: 10.1590/0004-282X-A NP-2019-0393.
24. Alhola P, Polo-Kantola P. Sleep deprivation: impact on cognitive performance. *Neuropsychiatric disease and treatment*. 2007;3(5):553-67.
25. Vaseghi S, Arjmandi-Rad S, Kholghi G, Nasehi M. Inconsistent effects of sleep deprivation on memory function. *EXCLI journal*. 2021;20:1011. Doi: 10.17179/excli2021-3764.
26. Jiao Q, Dong X, Guo C, Wu T, Chen F, Zhang K, et al. Effects of sleep deprivation of various durations on novelty-related object recognition memory and object location memory in mice. *Behav Brain Res*. 2022;418:113621. Doi: 10.1016/j.bbr.2021.113621.
27. Ma Y, Liang L, Zheng F, Shi L, Zhong B, Xie W. Association between sleep duration and cognitive decline. *JAMA network open*. 2020;3(9):e2013573-e. Doi: 10.1001/jamanetworkopen.2020.13573.
28. Khorasani A, Abbasnejad M, Esmaeili-Mahani S. Phytohormone abscisic acid ameliorates cognitive impairments in streptozotocin-induced rat model of Alzheimer's disease through PPAR β/δ and PKA signaling. *Int J Neurosci*. 2019;129(11):1053-65. Doi: 10.1080/00207454.2019.1634067.
29. Kooshki R, Anaieigoudari A, Abbasnejad M, Askari-Zahabi K, Esmaeili-Mahani S. Abscisic acid interplays with PPAR γ receptors and ameliorates diabetes-induced cognitive deficits in rats. *Avicenna J Phytomed*. 2021;11(3):247.
30. Swanwick CC, Murthy NR, Mtchedlishvili Z, Sieghart W, Kapur J. Development of γ -aminobutyric acidergic syn-

- apses in cultured hippocampal neurons. *J Comp Neurol.* 2006;495(5):497-510. Doi: 10.1002/cne.20897.
31. Tyzio R, Represa A, Jorquera I, Ben-Ari Y, Gozlan H, Aniksztejn L. The establishment of GABAergic and glutamatergic synapses on CA1 pyramidal neurons is sequential and correlates with the development of the apical dendrite. *J Neurosci.* 1999;19(23):10372-82. Doi: 10.1523/JNEUROSCI.19-23-10372.1999.
 32. Makkar SR, Zhang SQ, Cranney J. Behavioral and neural analysis of GABA in the acquisition, consolidation, reconsolidation, and extinction of fear memory. *Neuropsychopharmacology.* 2012;37(7):1793. Doi: 10.1038/npp.2010.53.
 33. Whissell PD, Eng D, Lecker I, Martin LJ, Wang D-S, Orser BA. Acutely increasing δ GABAA receptor activity impairs memory and inhibits synaptic plasticity in the hippocampus. *Front Neural Circuits.* 2013;7:146. Doi: 10.3389/fncir.2013.00146.
 34. Wimmer GE, Poldrack RA. Reward learning and working memory: Effects of massed versus spaced training and post-learning delay period. *Mem Cogn.* 2022;50(2):312-24. Doi: 10.3758/s13421-021-01233-7.
 35. Chen P, Ban W, Wang W, You Y, Yang Z. The Devastating Effects of Sleep Deprivation on Memory: Lessons from Rodent Models. *Clock Sleep.* 2023;5(2):276-94. Doi: 10.3390/clockssleep5020022.
 36. Celik I, Turker M, Tuluçe Y. Abscisic acid and gibberellic acid cause increased lipid peroxidation and fluctuated antioxidant defense systems of various tissues in rats. *J Hazardous Mat.* 2007;148(3):623-9. Doi: 10.1016/j.jhazmat.2007.03.018.
 37. Soti M, Abbasnejad M, Kooshki R, Esmaili-Mahani S. Central microinjection of phytohormone abscisic acid changes feeding behavior, decreases body weight, and reduces brain oxidative stress in rats. *Nutr Neurosci.* 2019;22(10):678-87. Doi: 10.1080/1028415X.2018.1431093.
 38. Chen X, Ding C, Liu W, Liu X, Zhao Y, Zheng Y, et al. Abscisic acid ameliorates oxidative stress, inflammation, and apoptosis in thioacetamide-induced hepatic fibrosis by regulating the NF- κ B signaling pathway in mice. *Eur J Pharmacol.* 2021;891:173652. Doi: 10.1016/j.ejphar.2020.173652.
 39. Naderi R, Esmaili-Mahani S, Abbasnejad M. Phosphatidylinositol-3-kinase and protein kinase C are involved in the pro-cognitive and anti-anxiety effects of phytohormone abscisic acid in rats. *Biomed Pharmacother.* 2017;96:112-9. Doi: 10.1016/j.biopha.2017.09.089.
 40. Kooshki R, Abbasnejad M, Esmaili-Mahani S, Raouf M. The Modulatory Role of Orexin 1 Receptor in CA1 on Orofacial Pain-induced Learning and Memory Deficits in Rats. *Basic Clin Neurosci.* 2017;8(3):213. Doi: 10.18869/nirp.bcn.8.3.213.
 41. McDermott CM, LaHoste GJ, Chen C, Musto A, Bazan NG, Magee JC. Sleep deprivation causes behavioral, synaptic, and membrane excitability alterations in hippocampal neurons. *J Neurosci.* 2003;23(29):9687-95. Doi: 10.1523/JNEUROSCI.23-29-09687.2003.
 42. Morris RG, Garrud P, Rawlins Ja, O'Keefe J. Place navigation impaired in rats with hippocampal lesions. *Nature.* 1982;297(5868):681-3.
 43. Kameyama T, Nabeshima T, Kozawa T. Step-down-type passive avoidance-and escape-learning method: Suitability for experimental amnesia models. *J Pharmacological Met.* 1986;16(1):39-52. Doi: 10.1016/0160-5402(86)90027-6.
 44. Hodges DM, DeLong JM, Forney CF, Prange RK. Improving the thiobarbituric acid-reactive-substances assay for estimating lipid peroxidation in plant tissues containing anthocyanin and other interfering compounds. *Planta.* 1999;207(4):604-11. Doi: 10.1007/s004250050524
 45. Velikova V, Yordanov I, Edreva A. Oxidative stress and some antioxidant systems in acid rain-treated bean plants. *Plants Sci.* 2000; 59-66. Doi: 10.1016/S0168-9452(99)00197-1.

COPYRIGHTS

©2025 The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution (CC BY 4.0), which permits unrestricted use, distribution, and reproduction in any medium, as long as the original authors and source are cited. No permission is required from the authors or the publishers.

**How to cite this article**

Fekrat A, Abbasnejad M, Kooshki R, Nikkhoy P, Abbasnejad M. Intra-Hippocampal Injection of Abscisic Acid Attenuates Learning and Memory Deficits, and Changes Oxidative Stress Indices in REM Sleep Deprived Rats . Iran J Vet Sci Technol.2025; 17(2): 29-37.

DOI: <https://doi.org/10.22067/ijvst.2025.86932.1355>.

URL:https://ijvst.um.ac.ir/article_46690.html